Rare Gastric Lesions Associated with Helicobacter pylori Infection: A Histopathological Review

Epstein-Barr Virus–Associated Lymphoproliferative Disorders: Review and Update on 2016 WHO Classification
Aims & Scope

The Journal of Pathology and Translational Medicine is an open venue for the rapid publication of major achievements in various fields of pathology, cytopathology, and biomedical and translational research. The Journal aims to share new insights into the molecular and cellular mechanisms of human diseases and to report major advances in both experimental and clinical medicine, with a particular emphasis on translational research. The investigations of human cells and tissues using high-dimensional biological techniques such as genomics and proteomics will be given a high priority. Articles on stem cell biology are also welcome. The categories of manuscript include original articles, review and perspective articles, case studies, clinical reports, and letters to the editor.

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Rare Gastric Lesions Associated with *Helicobacter pylori* Infection: A Histopathological Review

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*Helicobacter pylori* is a common gastric pathogen that causes gastritis, peptic ulcer disease, gastric adenocarcinoma, and mucosa-associated lymphoid tissue (MALT) lymphoma. *H. pylori* infection has been linked to a number of rare gastric mucosal lesions with distinctive histological features, including rare forms of gastritis, such as lymphocytic gastritis (LG)\(^1\)-\(^5\) or granulomatous gastritis (GG),\(^6\)\(^,\)\(^7\) and abnormal immunoglobulin deposits, such as Russell body gastritis (RBG)\(^8\) or crystal-storing histiocytosis (CSH).\(^9\)\(^,\)\(^10\) These lesions are easily diagnosed based on their distinctive histological features; however, their development can be attributed to various factors besides *H. pylori* infection. Therefore, the role of *H. pylori* as a causative organism remains debatable, and it has been suggested that *H. pylori* may be an “innocent bystander.”\(^11\)\(^,\)\(^12\) Nevertheless, *H. pylori* eradication therapy has been linked to clinical and histological improvements in a subset of these lesions,\(^2\)\(^-\)\(^21\) which supports the role of *H. pylori* infection in their development and implies that *H. pylori* eradication therapy could be an effective treatment. Therefore, in this review, we investigate the relationship between these rare gastric lesions and *H. pylori* infection, describe their characteristic histological features, and evaluate the role of *H. pylori* infection in their pathogenesis.

**Key Words:** Helicobacter pylori; Stomach; Gastritis; Rare; Immunoglobulins

**RARE FORMS OF GASTRITIS**

LG and GG are not distinct clinicopathological entities, but rather morphologic patterns of injury that can be secondary to a variety of underlying etiologies.\(^2\)\(^-\)\(^4\)\(^,\)\(^6\) The histological identification of intraepithelial lymphocytosis and granuloma formation are key diagnostic features of LG and GG, respectively.\(^1\)\(^,\)\(^7\) However, a morphological diagnosis of LG and GG should elicit clinical and laboratory workups to identify the underlying etiology. In Western countries, LG and GG are generally classified as special forms of *H. pylori*-negative gastritis.\(^3\)\(^,\)\(^4\) However, there is convincing evidence that *H. pylori* infection contributes to the pathogenesis of both LG\(^1\)\(^,\)\(^5\) and GG,\(^6\)\(^,\)\(^7\)\(^,\)\(^22\) and that *H. pylori* eradication therapy may be an effective treatment.\(^7\)\(^,\)\(^15\)\(^-\)\(^17\)\(^,\)\(^23\)\(^,\)\(^24\) Thus, it is possible that subsets of LG and GG could be categorized as *H. pylori*-associated gastritis. The potential role of *H. pylori* infection in the pathogenesis of each of these lesions is described below, along with their histopathological characteristics.

**Lymphocytic gastritis**

LG, first described by Haot *et al.*\(^25\) in 1988, is a rare form of chronic gastritis that is characterized by a dense lymphocytic infiltration of the surface and pit gastric epithelium known as “intraepithelial lymphocytosis.” LG was initially considered to be...
related to varioliform gastritis, which manifests as thickened mu-
cosa with “octopus-sucker” targetoid erosions; however, the endoscopic features of LG vary according to its severity, ranging from normal to hypertrophic. Histologically, LG is defined by the presence of ≥ 25 intraepithelial lymphocytes (IELs) per 100 epithelial cells (Fig. 1). Gastric intraepithelial lymphocytosis is associated with a variety of conditions, including celiac disease, H. pylori infection, Crohn disease, syphilis, hypertrophic gastropathy, Ménétrier’s disease, human immunodeficiency virus, and lymphoma. However, celiac disease and H. pylori infection are the main causes of LG, accounting for 38% and 29% of cases, respectively. The association between LG and celiac disease is relatively well established. Mild intraepithelial lymphocytosis with a low cut-off value (≥ 8 IELs/100 epithelial cells) has been observed in 84% of patients with celiac disease, and LG occurs in up to 45% of patients, with resolution of LG in response to a gluten-free diet.

However, questions remain regarding the role of H. pylori infection in the development of LG due to the discrepancy between the prevalence of H. pylori in the general population and the incidence of LG among patients with H. pylori–associated gastritis. Given the high global prevalence of H. pylori infection, it is unclear why the proportion of H. pylori–infected patients presenting with LG morphology (< 5%) is so low? Nevertheless, H. pylori eradication therapy can resolve H. pylori–associated LG by reducing IEL levels, and has been shown to improve symptoms and/or lead to regression of the gastritis, which supports a causal role of H. pylori infection in the development of LG. Previous studies have reported that H. pylori–associated LG frequently exhibits significant neutrophilic activity in addition to intraepithelial lymphocytosis, which is distinct from celiac disease-associated LG that exhibits intraepithelial lymphocytosis without neutrophilic infiltration. In this context, Nielsen et al. argued that H. pylori–associated intraepithelial lymphocytosis accompanied with significant neutrophilic infiltration should be considered “chronic active gastritis,” rather than LG. However, considering the fact that LG is a morphological diagnosis that is based on the presence of intraepithelial lymphocytosis (≥ 25 IELs per 100 epithelial cells), the use of the term “LG” remains relevant.

Interestingly, a considerable number of patients with LG and

Fig. 1. Lymphocytic gastritis. (A) The biopsy specimen shows a marked increase in intraepithelial lymphocytes (IELs) (over 25 IELs per 100 epithelial cells) with a top-heavy distribution. (B) Most IELs are positive for CD3 immunostaining.
positive *H. pylori* serology did not exhibit histological evidence of *H. pylori* infection.\(^1,13,15,22\) In addition, the beneficial effects of *H. pylori* eradication therapy in LG have been observed in patients who had positive serology but negative histology for *H. pylori*.\(^14,15\) Furthermore, even when *H. pylori* infection has been histologically confirmed in patients with LG, colonization tends to be mild and focal.\(^14,15\) These results imply that failure to histologically detect *H. pylori* may be related to sampling error due to low-level infection.\(^27\) Also, it is possible that LG development is a local, transient, or delayed immunological reaction to *H. pylori* infection, and is not a direct effect of the infection.\(^1,13,14,27\) Thus, if other causes can be excluded, *H. pylori* eradication therapy could be considered in symptomatic patients with LG who are histologically and/or serologically positive for *H. pylori*. This approach may be more appropriate in countries with a low prevalence of celiac disease and a high prevalence of *H. pylori* infection, even without histological detection of *H. pylori*.

IELs are functionally and phenotypically distinct from peripheral lymphocytes, and the majority of IELs in the gastrointestinal epithelium are CD3\(^+\)CD8\(^+\) T cells with cytotoxic potential.\(^11\) These IELs are thought to play an important role in mucosal immunity and have also been implicated in epithelial cell turnover by eliciting apoptosis through cytotoxic T-lymphocytes (CTLs).\(^31,32\) Recently, Han *et al.*\(^33\) studied IEL subpopulations and their cytotoxicity in *H. pylori*-infected gastric mucosa using T-cell–restricted intracellular antigen-1 (TIA-1; a marker for resting and activated CTLs) and granzyme B (GrB; a marker for activated CTLs). They found that the IELs consisted of a mixture of TIA-1\(^-\)/GrB\(^+\) CTLs, TIA-1\(^+\)/GrB\(^-\) CTLs, and CD4\(^+\) T cells in the infected mucosa. In addition, they found that *H. pylori*–associated LG was distinct from *H. pylori* gastritis, based on the increased IEL levels and changes in the cytotoxicity and distribution of the subpopulations: *H. pylori*–associated LG had a higher proportion of activated GrB\(^+\) CTLs, compared to *H. pylori* gastritis. There was also a parallel increase in epithelial apoptosis. Meanwhile, in a study by Oberhuber *et al.*,\(^34\) the proportion of GrB\(^+\) CTLs in *H. pylori*–associated LG (10.8%) was lower than that in idiopathic LG (12%) or celiac disease–associated LG (18.9%). Thus, although LG exhibits consistent histological features (regardless of etiology), IEL characteristics may vary depending on the underlying condition, which can lead to different clinical manifestations.

**Granulomatous gastritis**

GG is a rare disease that is characterized by the presence of granulomas, and is detected in 0.01%–0.35% of gastric biopsies.\(^6,7,17,35\) GG can be caused by a number of factors, including systemic disease (e.g., Crohn disease, sarcoidosis, or vasculitis), infection (e.g., tuberculosis, histoplasmosis, or syphilis), underlying malignancy, or foreign bodies.\(^4,6,7,36\) Crohn’s disease and gastric sarcoidosis are the two leading causes of GG, accounting for 20%–50% of cases in Western countries.\(^6,37\) Isolated or idiopathic granulomatous gastritis (IGG) was first described by Fahimi *et al.*\(^37\) in 1965, and is diagnosed by the exclusion of other granulomatous diseases. However, whether IGG can be considered a discrete condition remains controversial, as it is possible that a clear etiology could be identified through a more meticulous clinical work-up and long-term follow-up.

Dhillon and Sawyert\(^22\) first reported an association between GG and *H. pylori* infection in 1989. Since then a number of reports have been published that support their findings.\(^6,7,17,18,23,24,38-42\) These reports demonstrated that the mucosa surrounding granulomas in GG exhibits typical histological features of *H. pylori* gastritis, that features suggestive of other etiologies are absent, and that *H. pylori* eradication therapy can result in GG resolution. However, whether *H. pylori* plays a causative role in the pathogenesis of GG remains debatable. First, the incidence of GG is abnormally low relative to the *H. pylori* prevalence in the general population.\(^6,7,12,35\) Although Ectors *et al.*\(^6\) and Maeng *et al.*\(^1\) have reported the presence of *H. pylori* in 92% and 89% of GG cases, respectively, the overall incidence of GG was only 0.27% and 0.08%, respectively. Second, *H. pylori* organisms are rarely found within granulomas, implying *H. pylori* infection is a comorbidity rather than a cause in GG pathogenesis. Lastly, in cases of GG with *H. pylori* infection, granulomas often persist for 3–17 months after *H. pylori* eradication therapy,\(^17,24,39\) making its efficacy in GG questionable. It is plausible that although *H. pylori* can cause GG, granuloma formation is the result of a rare host response as opposed to a direct effect. Thus, *H. pylori* eradication therapy would be less effective in the resolution of GG, compared to its efficacy in conventional *H. pylori* gastritis.

Histologically, *H. pylori*–associated GG exhibits small non-necrotizing epithelioid granulomas with Langhans giant cells (Fig. 2), which are similar to those that are associated with sarcoidosis or Crohn disease. These granulomas tend to form in distinctive locations, such as the foveolar isthmus,\(^6\) with Maeng *et al.*\(^7\) reporting that the majority (66.7%) of granulomas were found there. They are also often in contact with a damaged pit (where *H. pylori* are commonly found) and are frequently accompanied by prominent neutrophil infiltration, which distinguishes them from the granulomas that are observed in Crohn disease or sarcoidosis.\(^6,7\) However, this characteristic morphology and local-
Immunoglobulin accumulations have not been consistently reported in subsequent studies. Therefore, there are no distinct histologic features that could be used to confirm H. pylori–associated GG. Rather, it appears that the conditions in the mucosa surrounding granulomas are more informative, as the presence of H. pylori and neutrophil-rich chronic active gastritis (associated with glandular atrophy or intestinal metaplasia) increase the likelihood of H. pylori infection. Taken together, although the presence of H. pylori in the vicinity of granulomas does not imply a causative association, H. pylori eradication therapy should be considered when there are no other underlying disease except H. pylori infection.

**INTRACELLULAR IMMUNOGLOBULIN ACCUMULATION IN ASSOCIATION WITH HELICOBACTER PYLORI INFECTION**

Immunoglobulin accumulations can be found in reactive and neoplastic plasma cells and include large intracytoplasmic spherical inclusions (Russell bodies), small intracytoplasmic monorular inclusions (Mott cells), intranuclear inclusions (Duchte bodies), and rare angular- or needle-shaped intracytoplasmic crystalline inclusions. These accumulations are associated with chronic inflammation with plasmacytosis, autoimmune disease, multiple myeloma, or other B-cell lymphomas. Although the exact mechanism of immunoglobulin accumulation is unclear, it may be due to simple over-production, altered production, abnormal secretion, or impaired excretion. In the gastric mucosa, diffuse plasma cell infiltration with immunoglobulin overproduction may result from chronic over-stimulation of plasma cells by mucosal pathogens, especially H. pylori. Scattered Russell bodies are often observed with H. pylori gastritis, whereas Dutcher bodies are frequently associated with low-grade MALT lymphoma. However, RBG and CSH are rarely reported in the stomach. As the incidence of these lesions is low relative to H. pylori–associated gastritis, the contribution of H. pylori infection to their development is questionable. Furthermore, careful evaluation of their underlying cause is essential, as they can be associated with

![Fig. 2. Granulomatous gastritis. (A) The biopsy specimen demonstrates diffuse chronic active gastritis with confluent granulomas including multinucleated giant cells. (B) A well-defined granuloma is noted just below the surface foveolar epithelium. Some Helicobacter pylori organisms are seen (arrows).](http://jpatholm.org/)

http://jpatholm.org/  
https://doi.org/10.4132/jptm.2017.04.03
monoclonal gammopathy or lymphoreticular neoplasms.\textsuperscript{20,43,48}
The possible connections between \textit{H. pylori} infection and these immunoglobulin accumulations, diagnostically relevant histological features, and biological significance are described below.

**Russell body gastritis**

The first case of RBG was reported by Tazawa and Tsutsumi in 1998.\textsuperscript{19} RBG is a rare form of chronic gastritis characterized by localized accumulation of Mott cells, which are plasma cells with a cytoplasm packed with small spherical inclusions (Fig. 3A, B). These lesions are rare, and only 30 cases of RBG have been published to date in the English literature.\textsuperscript{8,9,19-21,48-61} The clinical and pathological features of RBG are summarized in Table 1. Although its pathogenesis has not been fully elucidated, there is evidence to support a strong association between \textit{H. pylori} infection and RBG development. For example, \textit{H. pylori} is detected in approximately two-thirds of patients with RBG.\textsuperscript{8,9,19-21,48,52,57,60}\textsuperscript{61}

Few other infections that have been reported with RBG include human immunodeficiency virus (three patients),\textsuperscript{35,53,59} hepatitis C virus (one patient),\textsuperscript{58} and candida esophagitis (one case).\textsuperscript{49} In addition, more than 60% of patients with RBG exhibit lesion regression following \textit{H. pylori} eradication therapy.\textsuperscript{19-21,53,60} Furthermore, it has been reported that highly virulent \textit{H. pylori} genotypes (vacA and cagA) are associated with the formation of Russell bodies and Mott cells in the antral mucosa.\textsuperscript{62}

Immunoglobulin light chain restriction is generally considered to be proof of monoclonality and is an important indicator of B-cell neoplasia. Interestingly, light chain restriction was detected via immunohistochemistry in 12 out of 30 cases of RBG (kappa restriction in 11 cases and lambda restriction in one case).\textsuperscript{8,9,20,48,58,61}

![Fig. 3. Russell body gastritis with concomitant mucosa-associated lymphoid tissue lymphoma.](https://doi.org/10.4132/jptm.2017.04.03)

(A) Mott cells are plasma cells in which the cytoplasm is packed with multiple variable-sized Russell bodies. (B) The lamina propria of the gastric mucosa is expanded by extensive infiltration of Mott cells, consistent with Russell body gastritis. (C) Small- to intermediate-sized atypical lymphoid cells, morphologically consistent with centrocyte-like cells are admixed with Mott cells and destroy adjacent gastric glands to form a lymphoepithelial lesion (circle). (D) Immunostaining for cytokeratin highlights a lymphoepithelial lesion.
Ten of these cases were localized gastric lesions with no associated lymphoid malignancy or plasma cell disorder, although one case was associated with low-grade MALT lymphoma, and another with concomitant monoclonal gammopathy of undetermined significance. This could be explained by the findings of Girón and Shah, who reported that approximately 50% of patients with H. pylori infection exhibit either kappa or lambda light chain elevation, and suggested that H. pylori infection might contribute to immunoglobulin light chain dysfunction. Thus, RBG may be closely associated with H. pylori infection, and the majority of RBG cases may be reactive in nature, even when light chain monoclonality is detected. Nevertheless, pathologists should be aware of the possibility of concomitant lymphoid neoplasms.

Given the strong association with H. pylori infection, it is possible that RBG, gastric carcinoma, or MALT lymphoma might occur simultaneously in the same patient. Previous reports describe Mott cell proliferation (features of RBG) in association with gastric carcinoma, including two cases of Epstein-Barr virus–positive lymphoepithelioma-like carcinoma, and one case of signet ring cell carcinoma. In these cases, Mott cell proliferation was likely a reactive paraneoplastic event, given that the carcinoma and Mott cells did not mix and that the Mott cells were

Table 1. Clinical and pathologic findings of previously published cases of Russell body gastritis in the English literature

<table>
<thead>
<tr>
<th>Case</th>
<th>Study</th>
<th>Age (yr)/Sex</th>
<th>Endoscopic finding</th>
<th>Helicobacter pylori infection</th>
<th>Ig light chain of Mott cells</th>
<th>Gastric lesion coexisted</th>
<th>HPET/Resolution of RBG</th>
<th>Others</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Tazawa and Tsutsumi43</td>
<td>53/M</td>
<td>Multiple ulcer scars</td>
<td>Yes</td>
<td>Polyclonal</td>
<td>None</td>
<td>Done/Yes</td>
<td>-</td>
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<tr>
<td>2</td>
<td>Erbersdobler et al.44</td>
<td>80/F</td>
<td>Irregular mucosal swelling</td>
<td>No</td>
<td>Polyclonal</td>
<td>None</td>
<td>NS</td>
<td>Candida esophagitis</td>
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<tr>
<td>3</td>
<td>Ersari et al.45</td>
<td>70/M</td>
<td>Flattened gastric folds</td>
<td>Yes</td>
<td>Polyclonal</td>
<td>None</td>
<td>Done/NS</td>
<td>-</td>
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<tr>
<td>4</td>
<td>Paik et al.46</td>
<td>47/F</td>
<td>Erythematous swelling</td>
<td>Yes</td>
<td>Polyclonal</td>
<td>None</td>
<td>Done/NS</td>
<td>-</td>
</tr>
<tr>
<td>5</td>
<td>Paik et al.46</td>
<td>53/F</td>
<td>Yellowish raised lesion</td>
<td>Yes</td>
<td>Polyclonal</td>
<td>None</td>
<td>Done/NS</td>
<td>-</td>
</tr>
<tr>
<td>6</td>
<td>Woklersdörfer et al.47</td>
<td>54/M</td>
<td>Erythema and erosions</td>
<td>Yes</td>
<td>Monoclonal (λ)</td>
<td>None</td>
<td>Done/Yes</td>
<td>MGUS</td>
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<td>Drut and Olenchuk48</td>
<td>34/M</td>
<td>Elevation with central macule</td>
<td>No</td>
<td>Polyclonal</td>
<td>None</td>
<td>NS</td>
<td>HIV infection</td>
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<td>Pizzolitto et al.49</td>
<td>60/F</td>
<td>Minute-raised granular areas</td>
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<td>Polyclonal</td>
<td>None</td>
<td>Done/NS</td>
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<td>9</td>
<td>Licci et al.50</td>
<td>59/M</td>
<td>Hyperemia</td>
<td>Yes</td>
<td>Polyclonal</td>
<td>None</td>
<td>Done/Yes</td>
<td>HIV infection</td>
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<tr>
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<td>Habib et al.51</td>
<td>75/M</td>
<td>Nodular chronic active gastritis</td>
<td>No</td>
<td>Polyclonal</td>
<td>None</td>
<td>NS</td>
<td>-</td>
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<tr>
<td>11</td>
<td>Shinozaki et al.52</td>
<td>74/M</td>
<td>Centrally ulcerated bulky mass</td>
<td>Yes</td>
<td>Polyclonal</td>
<td>EBV-positive carcinoma</td>
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<td>29/F</td>
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<td>Del Gobbo et al.53</td>
<td>78/F</td>
<td>Hyperemic gastric mucosa</td>
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<td>Signet ring cell carcinoma</td>
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<td>49/M</td>
<td>Severe raised erosive gastritis</td>
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<td>Monoclonal (κ)</td>
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<td>NS</td>
<td>HCV infection</td>
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<td>16</td>
<td>Bhalla et al.56</td>
<td>82/M</td>
<td>Gastritis</td>
<td>No</td>
<td>Polyclonal</td>
<td>None</td>
<td>NS</td>
<td>HIV infection</td>
</tr>
<tr>
<td>17</td>
<td>Karabagli and Gokturk57</td>
<td>60/M</td>
<td>Large ulcerofungating mass</td>
<td>Yes</td>
<td>Polyclonal</td>
<td>None</td>
<td>Done/Yes</td>
<td>-</td>
</tr>
<tr>
<td>18</td>
<td>Yoon et al.58</td>
<td>57/M</td>
<td>Elevation with central depression</td>
<td>Yes</td>
<td>Polyclonal</td>
<td>None</td>
<td>Done/Yes</td>
<td>-</td>
</tr>
<tr>
<td>19</td>
<td>Yoon et al.58</td>
<td>43/M</td>
<td>Whitish flat lesion with nodularity</td>
<td>Yes</td>
<td>Polyclonal</td>
<td>None</td>
<td>Done/Yes</td>
<td>-</td>
</tr>
<tr>
<td>20</td>
<td>Araki et al.59</td>
<td>74/F</td>
<td>Ulcer</td>
<td>Yes</td>
<td>Monoclonal (κ)</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>21</td>
<td>Zhang et al.60</td>
<td>78/F</td>
<td>Gastritis with uneven mucosa</td>
<td>No</td>
<td>Monoclonal (κ)</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>22</td>
<td>Zhang et al.60</td>
<td>77/F</td>
<td>Gastritis with uneven mucosa</td>
<td>Yes</td>
<td>Monoclonal (κ)</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>23</td>
<td>Zhang et al.60</td>
<td>77/F</td>
<td>Punctiform erosion</td>
<td>Yes</td>
<td>Monoclonal (κ)</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>24</td>
<td>Zhang et al.60</td>
<td>56/M</td>
<td>Raised erosion</td>
<td>Yes</td>
<td>Monoclonal (κ)</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>25</td>
<td>Zhang et al.60</td>
<td>76/M</td>
<td>Erythema</td>
<td>Yes</td>
<td>Monoclonal (κ)</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>26</td>
<td>Zhang et al.60</td>
<td>50/M</td>
<td>Flat and raised erosions</td>
<td>Yes</td>
<td>Monoclonal (κ)</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>27</td>
<td>Zhang et al.60</td>
<td>28/M</td>
<td>Erythema</td>
<td>No</td>
<td>Monoclonal (κ)</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>28</td>
<td>Zhang et al.60</td>
<td>24/F</td>
<td>Erythema</td>
<td>No</td>
<td>Monoclonal (κ)</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>29</td>
<td>Zhang et al.60</td>
<td>66/M</td>
<td>Ulceration</td>
<td>No</td>
<td>NA</td>
<td>None</td>
<td>No</td>
<td>-</td>
</tr>
<tr>
<td>30</td>
<td>Joo61</td>
<td>56/M</td>
<td>Hyperemia and micronodularity</td>
<td>Yes</td>
<td>Monoclonal (κ)</td>
<td>MALT lymphoma</td>
<td>No</td>
<td>-</td>
</tr>
</tbody>
</table>

HPET, Helicobacter pylori eradication therapy; RBG, Russell body gastritis; M, male; F, female; NS, not stated; MGUS, monoclonal gammopathy of undetermined significance; HIV, human immunodeficiency virus; EBV, Epstein-Barr virus; HCV, hepatitis C virus; NA, not assessed; MALT, mucosa-associated lymphoid tissue.
polyclonal. However, in the previously described case of RBG with concomitant MALT lymphoma, the Mott cells were mixed with the neoplastic centrocyte-like cells (Fig. 3C) and exhibited IgM kappa monoclonality, which indicates the proliferating Mott cells were neoplastic components of MALT lymphoma.

Morphologically, Mott cells with eccentric nuclei and abundant eosinophilic cytoplasm are similar to poorly differentiated carcinoma cells or signet ring cells. In addition, in cases with abundant Mott cells and a few neoplastic cells, neoplastic cells may not be easily detected. Thus, immunostaining for cytokeratin should be conducted to exclude associated carcinoma. Furthermore, if light chain monoclonality is detected, pathologists should consider associated MALT lymphoma and perform ancillary immunostaining (e.g., for CD20 and cytokeratin) to identify centrocyte-like cells and lymphoepithelial lesions (Fig. 3D).

Crystal-storing histiocytosis

CSH is a rare condition, which often occurs with disorders such as monoclonal gammopathy, B-cell lymphoma, or plasma cell myeloma. Although many cases of CSH are systemic, organ-confined CSH has been described in the lung, lymph node, kidney, thyroid, thymus, parotid gland, and cornea. CSH is extremely rare in the stomach, and only eight cases of gastric CSH have been described to date in the English literature (Table 2). Among these, H. pylori infection was identified in four patients (50%) who did not exhibit concomitant gastric lesions (except for H. pylori gastritis) or a systemic disorder that might have caused monoclonal gammopathy. In the other four patients, there was no mention of H. pylori infection, and two of them were subsequently diagnosed with thymic lymphoma and plasma cell myeloma, respectively. Therefore, although overproduction of immunoglobulin due to H. pylori infection could be a plausible cause of isolated gastric CSH, clinical workup is needed to exclude the possibility that it is a manifestation of underlying lymphoma or plasma cell myeloma. Light chain restriction was detected in five of seven cases (kappa restriction in two cases and lambda restriction in three cases), all of which except one had no associated B-cell/plasmacytic neoplasms. Thus, light chain restriction detected in isolated gastric CSH does not necessarily mean that it is associated with B-cell or plasmacytic neoplasms. Meanwhile, because no studies have reported CSH responding to H. pylori eradication therapy, its effectiveness is unclear.

Histologically, CSH is characterized by diffuse infiltrations of large, oval, polygonal, and, occasionally, spindle cells, with abundant eosinophilic cytoplasm and small eccentric nuclei. The eosinophilic cytoplasm is filled with elongated, rectangular, and needle-shaped fibrillary crystalline inclusions (Fig. 4). These crystalline inclusions are approximately 5–20 nm long and are frequently arranged in parallel arrays. At low magnification, nodular aggregates of these cells can sometimes resemble adult rhabdomyomas or granular cell tumors in the way that they expand or displace normal structures, and proliferations of benign-looking histioyte-like cells can also resemble Gaucher disease or malakoplakia. Therefore, immunostaining for desmin, smooth muscle actin, S100 protein, and immunoglobulin light chains can facilitate an accurate diagnosis.

Table 2. Clinical and pathologic findings of six cases of gastric crystal-storing histiocytosis

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Study</th>
<th>Age (yr)/Sex</th>
<th>Endoscopic finding</th>
<th>Ig light chains</th>
<th>Crystal-storing cells</th>
<th>Helicobacter pylori infection</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Jones et al.</td>
<td>35/F</td>
<td>NS</td>
<td>Polyclonal</td>
<td>Histiocytes</td>
<td>NS</td>
</tr>
<tr>
<td>2</td>
<td>Stewart and Spagnolo</td>
<td>82/M</td>
<td>Gastritis</td>
<td>Monoclonal (IgA)</td>
<td>Plasma cells</td>
<td>Positive</td>
</tr>
<tr>
<td>3</td>
<td>Stewart and Spagnolo</td>
<td>81/M</td>
<td>Gastritis</td>
<td>NA</td>
<td>Plasma cells</td>
<td>Positive</td>
</tr>
<tr>
<td>4</td>
<td>Stewart and Spagnolo</td>
<td>52/F</td>
<td>Gastritis</td>
<td>Monoclonal (IgA)</td>
<td>Plasma cells</td>
<td>Positive</td>
</tr>
<tr>
<td>5</td>
<td>Joo et al.</td>
<td>56/F</td>
<td>Polyps (three)</td>
<td>Polyclonal</td>
<td>Plasma cells and histiocytes</td>
<td>Positive</td>
</tr>
<tr>
<td>6</td>
<td>Vaid et al.</td>
<td>NS</td>
<td>Submucosal tumor</td>
<td>Monoclonal (k)</td>
<td>Histiocytes</td>
<td>NS</td>
</tr>
</tbody>
</table>

F, female; NS, not stated; M, male; NA, not assessed.

CONCLUSION

This review examined several rare gastric lesions that have distinctive histological characteristics and are associated with a variety of conditions, including H. pylori infection. Although H. pylori may be a cause in many of these conditions, the association cannot be viewed as definite, given the low incidence of these lesions relative to the high prevalence of H. pylori infection, regional differences in the prevalence of H. pylori infection, and the possibility of other causative disorders. However, it is reasonable to consider H. pylori once other potential etiologies have been excluded. In addition, it is not advisable to consider H. pylori to be an “innocent bystander,” given the considerable proportion of these lesions that can be regressed or cured with H. pylori eradication therapy. Therefore, it is important that pathologists properly identify a lesion’s cause in order to ensure ap-
appropriate patient management. In this context, *H. pylori* should be considered as a possible cause in areas where it is prevalent. Further investigation is needed to confirm the role of *H. pylori* in the development of rare gastric lesions.

**Conflicts of Interest**

No potential conflict of interest relevant to this article was reported.

**REFERENCES**


[Fig. 4. Gastric crystal-storing histiocytosis. (A) The biopsy specimen demonstrates many large pinkish mononuclear cells in the lamina propria. (B) Higher magnification of mononuclear cells shows densely eosinophilic, refractile, needle-shaped, intracytoplasmic crystalline inclusions.]


Epstein-Barr virus (EBV) is classified as a γ-herpes virus and contains a linear DNA molecule about 172 kb in length, which affects more than 90% of the worldwide adult population. If the infection does not become clinically silent, infectious mononucleosis is experienced by the exposed persons. Although EBV infection is lifelong, a long latency and reactivation of EBV results in various lymphoproliferative lesions including hematologic malignancies. This article reviews the current understanding of EBV-associated lymphoproliferative disorders (LPDs) based on the 2016 classification of the World Health Organization.
(WHO). Some LPDs exhibit a predisposition in Asian populations, including Koreans. These entities often pose diagnostic challenges, both clinically and pathologically, and it is important to understand their unique pathophysiology for correct diagnoses and optimal management. Generally intrinsic defects and post-transplant lymphoproliferative disorders (PTLDs) are excluded from this review, these are treated separately as immunodeficiency disorders by the WHO.

**EPSTEIN-BARR VIRUS–ASSOCIATED B-CELL LYMPHOPROLIFERATIVE DISORDERS**

The spectrum of EBV-associated B-cell LPDs is broad, ranging from reactive lymphoproliferative lymphadenitis to lymphomas. All the related disease entities are shown in Table 1. Well-defined lymphoma entities such as Burkitt lymphoma, classical Hodgkin lymphoma, and plasmablastic lymphoma are not included. Infections mononucleosis

Primary EBV infection occurs most often in childhood and is generally asymptomatic. In adolescence, it is associated with a self-limiting infectious mononucleosis syndrome, manifested by fever, pharyngitis, malaise, and atypical lymphocytosis. Following primary infection, most individuals remain a life-long carrier of the virus without serious sequelae. However, a small population with the latent infection will develop various LPDs.

EBV is transmitted from the host by saliva and infected EBV replicates within oropharyngeal epithelium and is then exposed to circulating B-lymphocytes. Peripheral EBV-infected memory B-cells can return to the Waldeyer's ring, undergoing reactivation to produce an infectious virus that will be shed in the saliva. EBV-specific cytotoxic T-cells (CTL) destroy most infected cells. The histologic features of infectious mononucleosis vary during the course of the disease. Early in the disorder, follicular hyperplasia occurs with mononuclear B-cell aggregates and epithelioid histiocytes. Later, the expansion of the paracortex predominates. The immunoblasts resemble classical Reed-Sternberg (RS) cells. The immunoblasts of predominantly B-cell types and partly T-cell types often express CD30. In situ hybridization (ISH) of EBV-encoded small RNAs (EBERs) exhibit numerous positive immunoblasts in the paracortex but not in the germinal centers.

**Infectious mononucleosis**

**Chronic active EBV of B-cell types**

As first defined by Lekstrom-Himes et al., chronic active EBV (CAEBV) of B-cell types refers to a chronic or persistent EBV infection characterized by a severe illness lasting more than 6 months, persistent elevated EBV titers, and evidence of EBV-related organ damage. Currently, CAEBV is defined as (1) a severe progressive illness with a duration of more than 6 months, (2) lymphocytic infiltration of tissue (e.g., lymph nodes, lungs, liver, central nervous tissue, bone marrow, eye, and skin), (3) elevated EBV DNA and RNA in affected tissue, and (4) absence of any other immunosuppressive conditions. Histologically, the lymph nodes exhibits features resembling polymorphic PTLD, with paracortical expansion, plasmacytoid lymphoblastic proliferation, presence of plasma cells, and presence of occasional RS-like cells. EBV-ISH positive B-cells are noted in the paracortex. Among CAEBV patients, 63% had clonal immunoglobulin rearrangement.

**EBV-positive diffuse large B-cell lymphoma**

EBV-positive diffuse large B-cell lymphoma (DLBCL), not otherwise specified was originally described as “senile EBV-associated B-cell LPD,” or “EBV-positive DLBCL of the elderly” (older than 50 years) (WHO 4th edition). However, subsequent studies have shown that EBV-positive DLBCL is not limited to this older age group. In the elderly group, it is thought to be related to immunosenescence, which modifies T-cell homeostasis through a lack of thymic output of naïve T-cells and an accumulation of viral specific CD8+ T-cells. Histologically, four types have been described: monomorphic (DLBCL-like, monotonous sheets of large cells), polymorphic in the inflammatory background, T-cell/histiocyte-rich large cell lymphoma, and plasmacytoid differentiation. Immunophenotypically, the tumor cells express pan B-cell markers (CD20, PAX5, CD79a, OCT-2, and BOB-1), and are mostly CD30+, but lack CD15 expression.

This disease entity frequently involves extranodal sites including the skin, lung, tonsils, and stomach. Unfavorable prognostic factors include older age (> 70 years), high international prognostic index, and activated B-cell phenotype.

**EBV mucocutaneous ulcer**

This self-limited EBV positive B-cell proliferation is characterized by the presence of mucocutaneous ulcers with an indolent clinical course. This phenomenon is likely due to a more localized form of decreased immune surveillance, which is supported by a very low EBV viral load. The frequently involved sites includes the skin, oropharyngeal mucosa, and gastrointestinal tract.
with RS cell-like features. Phenotypically the large atypical cells are variably positive for CD20 and CD30, and uniformly positive for EBV-ISH and CD15+ in half of the cases (Fig. 1).

**DLBCL associated with chronic inflammation**

DLBCL associated with chronic inflammation develops in the setting of long-standing chronic inflammation with EBV association. It usually involves body cavities or enclosed spaces (like cysts). Pyothorax-associated lymphoma (PAL) represents the prototype of this entity.

Cases with PAL have long history of chronic pyothorax and may present with chest pain, fever, cough, dyspnea, and tumor mass. The prognosis of PAL is poor.

The morphology discloses a diffuse proliferation of large atypical lymphocytes with plasmacytoid cytomorphology. Immunohistochemistry reveals neoplastic cells that represent pan-B cell markers, usually positive for IRF4/MUM1, and CD13. An aberrant expression of T-cell phenotypes is also seen.

The unique genomic instability has been reported as follows: A20 deletion, interferon-inducible 27 (IFI27), TP53 mutation, and MYC amplification.13

**Lymphomatoid granulomatosis**

Katzenstein et al.14 initially described a rare angiocentric and angiodestructive EBV-associated LPD, which was distinct from Wegener’s granulomatosis.

Nearly all patients present with symptoms related to pulmonary involvement, followed by involved sites of the central nervous system, skin, liver, and kidney. Radiologically, bilateral variable sized lung nodules are noted in lymphomatoid granulomatosis (LYG).

The histologic features of LYG are observed in lung nodules. All the lesions are angioinvasive and angiocentric with fibrinoid necrosis of the vascular wall. The infiltrate is polymorphous with an admixture of small lymphocytes, histiocytes, and large lymphoid cells. The grading of LYG is based on the proportion of

| Table 1. Epstein-Barr virus–associated B-cell lymphoproliferative diseases |
|-----------------------------|-----------------------------|
| Disease                    |                             |
| Infectious mononucleosis   | Chronic active Epstein-Barr virus of B-cell type |
|                           | Epstein-Barr virus-positive diffuse large B-cell lymphoma |
|                           | Epstein-Barr virus mucocutaneous ulcer |
|                           | Diffuse large B-cell lymphoma associated with chronic inflammation |
|                           | Lymphomatoid granulomatosis |

**Fig. 1.** Mucocutaneous ulcer (Courtesy of Dr. J.H. Paik). (A) This 70-year-old female presented with a sore throat, painful swelling saliva, and tonsillar enlargement with a discrete ulcer. (B) The scanning power view shows a dense infiltrate beneath the ulcer. (C) Medium sized atypical lymphocytes are observed. (D) Epstein-Barr virus (EBV)-in-situ hybridization positive cells are aggregated in the ulcer bed. (E) CD20 immunostaining disclosed overlapping with EBV-positive cells. (F) The large atypical cells are diffusely and strongly positive for CD20.
EBV positive cells. In general, grade 1 and grade 2 lesions are approached using strategies that are designed to improve the host’s immune system, whereas grade 3 lesions require chemotherapy and do not respond to immunomodulatory therapies. Phenotypically, large atypical EBV-positive B cells express CD20, PAX5, CD79a, CD30+, and CD15.\textsuperscript{13}

**EBV-ASSOCIATED T-CELL AND NATURAL KILLER CELL LPDS**

EBV is a ubiquitous herpes virus with tropism for B cells, but the infection of T cells and natural killer (NK) cells may lead to several EBV-related LPDs. EBV-positive T/NK LPD encompasses disease entities with a broad clinicopathologic spectrum (Table 2).\textsuperscript{16}

**Table 2.** EBV-associated T-cell and NK cell lymphoproliferative diseases

<table>
<thead>
<tr>
<th>Disease</th>
<th>EBV-associated hyperinflammatory syndrome</th>
<th>EBV-associated hemophagocytic lymphohistiocytosis</th>
<th>CAEBV-type T/NK cell disease</th>
<th>Systemic chronic active EBV infection of T cell or NK cell type</th>
<th>Cutaneous forms of CAEBV</th>
<th>Severe mosquito bite allergy</th>
<th>Hydroa vacciniforme-like lymphoproliferative disease</th>
<th>Malignant T/NK cell disease</th>
<th>Extracranial NK/T cell lymphoma</th>
<th>Extranasal NK/T cell lymphoma</th>
<th>Aggressive NK cell leukemia</th>
<th>EBV-positive nodal NK/T cell lymphoma (provisional)</th>
</tr>
</thead>
</table>

EBV, Epstein-Barr virus; CAEBV, chronic active EBV; NK, natural killer.

**EBV-associated hemophagocytic lymphohistiocytosis**

Hemophagocytic lymphohistiocytosis (HLH) is a clinicopathologic syndrome encompassing a markedly dysregulated immune response and hypercytokinemia. HLH is characterized clinically by fever, splenomegaly, and cytopenias, and histologically by hemophagocytosis. The supportive laboratory findings for HLH are as follows: extremely high serum level of ferritin, lactate dehydrogenase, soluble CD25, and elevated viral capsid. EBV associated HLH accounts for 40\% of HLH.

Histologically, EBV-positive T-cells and hemophagocytosing histiocytes are scattered in the sinusoids of the bone marrow and liver. The T-cells express CD8 and granzyme B. Even though it is uncommon, NK cells can be infiltrated in HLH.\textsuperscript{17}

HLH can be effectively controlled in most patients (more than 90\%), but the other 10\% often die of fulminant disease.

**CAEBV infection of T-cell or NK-cell types, systemic**

CAEBV infection was initially defined as follows: (1) markedly abnormal EBV antibody titer; (2) histologic evidence of organ involved- interstitial pneumonia, hypoplasia of the bone marrow, uveitis, lymphadenitis, persistent hepatitis, or splenomegaly; and (3) increased EBV RNA in affected tissue.\textsuperscript{18}

Clinically, CAEBV-T/NK is a disease of children but is also detected in young adults and even in middle-aged older adults with a mean age of 11.3 years.\textsuperscript{19} The symptoms are usually prolonged fever, hepatomegaly, splenomegaly, thrombocytopenia, anemia, and lymphadenopathy.\textsuperscript{19} Life-threatening complications include hemophagocytic syndrome, interstitial pneumonia, malignant lymphoma, coronary aneurysms, and central nervous system involvement. All patients have elevated levels of EBV DNA in their blood.

![Fig. 2. Chronic active Epstein-Barr virus (EBV) infection of a T-cell or natural killer cell type, systemic (Courtesy of Dr. Y.H. Ko). (A) A 21-year-old man presented with severe oral ulcer, recurrent pneumonia, thrombocytopenia, and elevated liver enzymes for 2 years. Liver biopsy reveals atypical T lymphocytes infiltrating the sinusoidal and hepatic lobules. (B) EBV-encoded small RNA (EBER) in-situ hybridization exhibits positive signals in these T cells. (C) Bone marrow biopsy shows small lymphocytic infiltrate. (D) CD3 is expressed in most lymphocytes. (E) EBER in-situ hybridization also shows positive signals in T cells.](https://doi.org/10.4132/jptm.2017.03.15)
blood, which is well-correlated with clinical severity. Morphologically, in patients with CAEBV-T/NK the lymph nodes exhibit paracortical hyperplasia with polymorphic and polyclonal lymphoid proliferation and large numbers of EBER positive cells. The liver exhibits portal or sinusoidal infiltration by small lymphocytes with no definite atypia (Fig. 2).

Ohshima et al. proposed a three-tier classification as follows: category A1 is polymorphic LPD with polyclonal proliferation of EBV-infected T cells or NK cells; category A2 is polymorphic LPD with monoclonal T/NK cells; and category A3 is monomorphic LPD of monoclonal T cells.

Severe mosquito bite allergy
A severe mosquito bite allergy is a cutaneous manifestation of chronic EBV infection characterized by intense local skin symptoms, such as erythema, bullae, ulcers, and scarring. The systemic symptoms such as fever, lymphadenopathy, and liver dysfunction are developed after mosquito bites, vaccination, or injection.

The epidermis at the mosquito bite site exhibits necrosis and ulceration. The dermis reveals edema and infiltration of polymorphonuclear leukocytes, nuclear debris, and extravasated erythrocytes with fibrinoid necrosis of small vessels. The infiltrating small lymphocytes extend from the dermis to the subcutis in an angiocentric pattern. EBV-positive cells represent 3%-10% of infiltrating lymphocytes.

Hydroa vacciniforme-like LPD
Hydroa vacciniforme (HV)-like LPD is one of the cutaneous forms of CAEBV. It is initially described as an EBV positive polyclonal or monoclonal T/NK LPD, characterized by blistering photodermatoses in childhood and healed with vacciniform scarring. It is clinically divided into two types. The classic type is a self-limited disease with vesicles on sun-exposed areas in adolescence or young adulthood. Severe HV-type tends to exhibit more extensive skin lesions and systemic manifestations of fever, hepatomegaly, serologic abnormalities, and peripheral NK lymphocytosis. The severe type of HV often progresses to EBV-associated NK/T-cell malignancy.

Morphologic findings of HV are epidermal reticular degeneration to spongiotic vesiculation with perivascular and periappendicical lymphocytic infiltration with no definite cytologic atypia. Severe HV and HV-like T-cell lymphomas mimic those of classic HV, but the dermal infiltrates are more extensive and deeper, composed of variably atypical lymphocytes.

The immunophenotype of the classic HV is CD4+ or CD8+ T-cells, but the severe form/lymphoma exhibits predominantly CD8+ CTLs. The majority are αβ T cells and rarely NK cells.

Systemic EBV-positive T-cell lymphoma
Systemic EBV-positive T-cell lymphoma of childhood and young adulthood is a fulminant illness of EBV-infected T cells with clonal proliferation and cytotoxic phenotype. The clinical manifestation of this lesion is a rapid clinical progression with multiple organ failure, sepsis, and death. A hemophagocytic syndrome is nearly always associated. Systemic EBV-positive T-cell lymphoma arising in patients with a history of CAEBV-T/NK develops in a median time of 35 months.

Hyperplasia of histiocytes and marked hemophagocytic syndrome are also noted with increased small T-cells in the bone marrow, spleen, and liver. The paracortical zone of the lymph node is expanded with the depletion of B-cell areas. The degree of cytologic atypia in EBV-positive lymphocytes is variable.

Extranodal NK/T cell lymphoma, nasal type
Extranodal NK/T cell lymphoma is a prototype of EBV-associated T-cell LPD, which is characterized by frequent necrosis, angiocentric growth, cytotoxic phenotype and a strong association with EBV. Since the nasal cavity is the most commonly involved site, the nasopharynx and upper aerodigestive areas including the nasal cavity disclose progressively destructive and ulcerative lesions or obstructive symptoms due to mass effects.

Extranodal NK/T cell lymphoma
Extranodal NK/T cell lymphomas frequently involve the skin, gastrointestinal tract, testis, and soft tissue. Most patients present at a higher stage with multiple areas of involvement.

The morphology of the involved sites is frequently ulcerated and necrotic. The cytologic composition varies ranging from small, medium, and large. Angiocentric growth accompanies diffuse necrosis, and vascular damage.

Aggressive NK cell leukemia
Aggressive NK-cell leukemia is a neoplasm of NK cells, which primarily involves peripheral blood and bone marrow. In contrast to conventional leukemia, the tumor cells may not be abundant in the peripheral blood and bone marrow. The average age of patients exhibiting this disorder is 39 years. The typical presentations include fever, hepatosplenomegaly, lymphadenopathy, and is complicated by hemophagocytic syndrome.

In histologic sections, there are diffuse, destructive and permeative infiltrates of monomorphic cells with a round to moderate
rim of pale or amphophilic cytoplasm. Interspersed apoptotic bodies and zonal cell death are common. Angioinvasive and angiodestructive growth is also frequently noted. The clinical course is fatal.

**EBV-positive nodal NK/T cell lymphoma**

EBV-positive nodal T/NK cell lymphoma may involve a limited number of extranodal organs except for the nasal cavity, but the main bulk of the tumor is located in the lymph nodes. This entity is very rare with fewer than 100 cases being reported in the literature.27

The lymph nodes exhibit a diffuse infiltration of pleomorphic, variable sized cells. The cytomorphology of the tumor cells are more commonly centroblastoid, often anaplastic or plasmacytoid with some RS-like cells being noted. Some cases show extensive necrosis, many apoptotic bodies, and angiocentric growth patterns. The immunophenotype is as follows: CD3+, CD8+, TIA+, and granzyme B+.28

The disease course is very aggressive, with a median survival of only 4 months.

**CONCLUSION**

Many new entities and concepts for EBV-positive LPDs have been added to the 2016 WHO classification, based on growing knowledge in the field of genetics and molecular virology. With this increased understanding of LPD the clinical and pathologic entities, we can perform EBV-ISH indicative of (1) clinically bordered between infection and neoplastic conditions; (2) past history of recurrent inappropriate immune response, especially in children/young adults or old age; and (3) a pathologically polymorphous inflammatory background (but not Hodgkin lymphoma). It helps that the final destination is the achievement of appropriate diagnosis and management of LPDs. Some entities are provisional and must wait for confirmation of additional data.

**Conflicts of Interest**

No potential conflict of interest relevant to this article was reported.

**REFERENCES**


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Implication of PHF2 Expression in Clear Cell Renal Cell Carcinoma

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Background: Clear cell renal cell carcinoma (CCRCC) is presumed to be associated with adipogenic differentiation. Histone modification is known to be important for adipogenesis, and the function of histone demethylase plant homeodomain finger 2 (PHF2) has been noted. In addition, PHF2 may act as a tumor suppressor via epigenetic regulation of p53 and is reported to be reduced in colon cancer and stomach cancer tissues. In this study, we examined PHF2 expression in CCRCC specimens by immunohistochemistry. Methods: We studied 254 CCRCCs and 56 non-neoplastic renal tissues from patients who underwent radical or partial nephrectomy between 2000 and 2003 at the Seoul National University Hospital. Tissue microarray blocks were prepared, and immunohistochemical staining for PHF2 was performed. Results: Among 254 CCRCC cases, 150 cases (59.1%) showed high expression and 104 cases (40.1%) showed low expression. High expression of PHF2 was significantly correlated with a low Fuhrman nuclear grade (p < .001), smaller tumor size (p < .001), low overall stage (p = .003), longer cancer-specific survival (p = .002), and progression-free survival (p < .001) of the patients. However, it was not an independent prognostic factor in multivariate analysis adjusted for Fuhrman nuclear grade and overall stage. Conclusions: Our study showed that low expression of PHF2 is associated with aggressiveness and poor prognosis of CCRCC.

Key Words: PHF2 protein, human; Carcinoma, renal cell; Clear cell renal cell carcinoma; Immunohistochemistry; Adipogenesis

Renal cancer is one of the most common cancers in Korean males.1 Among the types of renal cancer, clear cell renal cell carcinoma (CCRCC) is the most common. CCRCC can exhibit various histologic features,2 but most tumors are golden yellow due to the abundant lipid contents of the cells, and microscopically, the tumor cells have a clear cytoplasm similar to adipocytes.3 The tumor cells have abundant cytoplasmic lipids in addition to glycogen, as revealed by an ultrastructural study.4 For this reason, CCRCC is presumed to be associated with adipogenic differentiation. Until recently, although three studies have shown that CCRCC is associated with adipogenic differentiation,5–7 the association of tumorigenesis with clear cell morphology is still required at the molecular level.

In recent studies, histone modification has been confirmed to be important in adipogenesis. In particular, the function of histone demethylase plant homeodomain finger 2 (PHF2), as it maps to human chromosome 9q22,8 has been noted.9,10 PHF2 is a dimethylated histone H3 lysine 9 (H3K9me2) demethylase10 that acts both in gluconeogenesis of hepatocytes and in signaling for regulation of immune and homeogenesis of macrophages in vitro.11,12 The physiological role of PHF2 in vitro is not yet clear, but it may be a co-activator with multiple transcription factors.10 To the best of our knowledge, only a few studies have investigated the association of PHF2 expression with human cancers, such as breast cancer, esophageal cancer and squamous cell carcinoma of the head and neck.13–15 A recent study has shown that PHF2 can act as a tumor suppressor through epigenetic regulation of p53 and shows reduced expression in colon cancer and stomach cancer tissues.16 However, there has been no report of PHF2 expression in CCRCC.

In this study, we investigated the relationship between the expression level of PHF2 and clinicopathological factors and prognosis in CCRCC patients using immunohistochemical staining of CCRCC specimens.

MATERIALS AND METHODS

Patients and tissue microarray

We examined 254 CCRCCs and 56 non-neoplastic renal tissues including the cortex and medulla from patients who underwent radical or partial nephrectomy between 2000 and 2003 in Seoul National University Hospital. Tissue microarray (TMA) blocks
containing representative tumor and non-neoplastic tissue core sections (2 mm in diameter) were prepared from each formalin-fixed paraffin block (SuperBioChips Laboratories, Seoul, Korea). All available hematoxylin and eosin–stained slides were reviewed to assess the validity of the diagnosis, and tumors were graded from 1 to 4 according to the Fuhrman nuclear grade system. Clinical and pathological information was collected from electronic medical records and pathological reports. The follow-up period ranged from 2 to 163 months, and the median follow-up period was 84.5 months. This study was approved by Seoul National University Hospital's Institutional Review Board (IRB).

Immunohistochemistry

Immunohistochemical staining for PHF2 was performed on 4 μm-thick sections taken from TMA. Immunohistochemistry (IHC) was performed using the Ventana Benchmark XT automated dyeing system (Ventana Medical Systems, Tucson, AZ, USA). Polyclonal rabbit anti-PHF2 antibody (Novus Biologicals, Littleton, CO, USA) was diluted 1:200. Immunohistochemically stained TMA slides were individually reviewed by two pathologists without clinicopathological information.

In non-neoplastic renal tissue, PHF2 was commonly expressed in the nucleus and cytoplasm of the proximal tubular epithelium, but only in the nucleus of the distal tubular epithelium and podocytes of the glomerulus (Fig. 1). The expression of PHF2 was assessed in both the nucleus and cytoplasm of CCRCCs and classified as high when nuclear expression was observed in more than 10% of the tumor cells, regardless of cytoplasmic expression (Fig. 2).

Statistical analysis

Statistical analysis was performed using the statistical program SPSS ver. 21.0 (IBM Corp., Armonk, NY, USA). The chi-squared test was used to analyze the relationship between PHF2 expression and clinicopathological characteristics. Survival curves were generated using the Kaplan-Meier method, and survival rate differences were compared using the log-rank test.

Fig. 1. Plant homeodomain finger 2 (PHF2) expression in non-neoplastic renal tissue. PHF2 is expressed in the nucleus and cytoplasm of the proximal tubular epithelium and in the nucleus of the distal tubular epithelium and podocytes in the glomerulus. (A) Cortex. (B) Medulla.

Fig. 2. Plant homeodomain finger 2 expression in clear cell renal cell carcinoma. (A) High expression (B) Low expression.
The multivariate Cox proportional hazard model was used to analyze the importance of various variables for survival. P-values less than .05 were considered statistically significant.

RESULTS

Basic clinicopathological characteristics

Of the 254 patients in this study, 187 (73.6%) were males and 67 (26.4%) were females. The age at diagnosis was between 28 and 82 years old. The mean age±standard deviation (SD) and median age were 56.2±11.3 years and 57 years, respectively. We have classified them into two groups: older than 57 years or not. The Fuhrman nuclear grade distribution was 16 cases in grade I (6.3%), 113 cases in grade II (44.5%), 93 cases in grade III (36.6%), and 32 cases in grade IV (12.6%). The tumor size ranged from 1.0 to 22.0 cm and the mean±SD size was 5.6±3.5 cm. At the time of diagnosis, 23 patients (9.1%) had metastatic cancers and 231 (90.9%) did not. According to the prognostic classification of the American Joint Committee on Cancer 7th edition, the T stage was as follows: T1 was 164 cases (64.6%), T2 was 38 cases (15.0%), T3 was 50 cases (19.7%), and T4 was two cases (0.8%). Additionally, the overall stage was 162 cases in stage I (63.8%), 31 cases in stage II (12.2%), 37 cases in stage III (14.6%), and 24 cases in stage IV (9.4%) (Table 1).

Immunohistochemical result of PHF2 and relationship with clinicopathological characteristics

Of the 254 cases, 150 cases (59.1%) showed high expression, and 104 cases (40.1%) showed low expression. The high expression of PHF2 was associated with a younger age group (p = .006) and significantly correlated with low Fuhrman nuclear grade, small tumor size and low overall stage (p < .001, p < .001, and p = .003, respectively) (Table 1). However, there was no association with the patients’ sex, T stage, or metastasis at diagnosis.

The association between PHF2 expression and survival time of the patients

To assess prognostic value, we evaluated the association between PHF2 expression and patient survival by a log-rank test using Kaplan-Meier analysis. We observed that high expression of PHF2 was significantly associated with longer cancer-specific survival (p = .002) and progression-free survival (p < .001) in CCRCC patients (Fig. 3).

Univariate and multivariate Cox regression analysis of cancer-specific survival and progression-free survival

Univariate Cox regression analysis showed significant correlation with age (less than 57 or not), Fuhrman nuclear grade, tumor size (less than 5 cm or not), T stage, metastasis at diagnosis, overall stage, PHF2 expression status, cancer-specific survival (p = .009 for age, p = .003 for PHF2 expression status, and p < .001 for the others), and progression-free survival (p = .001 for age and p < .001 for the others) (Table 2). However, multivariate analysis adjusted for age, Fuhrman nuclear grade and overall stage showed that high expression of PHF2 was not an independent prognostic factor in CCRCC patients with cancer-specific survival (p = .566) and progression-free survival (p = .099) (Table 2).

DISCUSSION

In this study, we analyzed the association of PHF2 expression with other clinicopathological parameters and survival time of the CCRCC patients. High expression of PHF2 was associated with a young age group and correlated significantly with low Fuhrman nuclear grade and overall stage. We also observed that high expression of PHF2 was significantly correlated with longer cancer-specific survival and progression-free survival in patients.
Multivariate analysis adjusted for age, Fuhrman nuclear grade and overall stage showed that high expression of PHF2 was not an independent prognostic factor for cancer-specific survival and progression-free survival in CCRCC patients.

PHF2 is a type of histone demethylase, and histone modification is known to be important in adipogenesis. PHF2 has been shown to play a role in gluconeogenesis in hepatocytes of fasted mice and proinflammatory gene regulation in macrophages in vitro. In a recent study, PHF2 was shown to play an important role in adipogenesis in transgenic mice. PHF2 demethylates the AT-rich interactive domain-containing protein 5B (ARID5B) and binds the promoter regions of target genes by forming a complex with demethylated ARID5B. PHF2 interacts with CCAAT/enhancer-binding protein alpha (CEBPA), one of the major regulators of adipogenesis, and promotes adipogenesis by demethylating H3K9me2 in the promoter region of CEBPA target gene. These results indicate that PHF2 promotes adipogenesis by coactivation of CEBPA and that PHF2 is a novel histone methylation-modifying enzyme that modulates adipogenesis.
As mentioned earlier, some studies have shown that adipogenic differentiation is associated with CCRCC. One study showed that gene expression patterns and IHC results in CCRCC tissues were associated with adipogenesis. They also found that CCRCC cells undergo adipogenic transdifferentiation, based on the observation that the clear cell morphology of CCRCC cells disappeared in standard cell culture media and was redeveloped in adipogenic media. This study concluded that adipogenesis was a type of epithelial mesenchymal transition. Another study has shown that adipose differentiation-related protein (ADFP) expression is increased at both mRNA and protein levels of CCRCC compared to non-neoplastic renal tissues and other types of renal cell carcinoma. ADFP was originally known as a protein associated with lipid metabolism, highly expressed in adipocytes and variably expressed in other cells. Higher ADFP levels in CCRCC were associated with a lower grade, lower stage and better prognosis, while higher ADFP levels were an independent good prognostic factor in multivariate analysis.

In previous studies, CCRCC with low nuclear grade showed a typical clear cell morphology, but CCRCC with a higher nuclear grade showed reduced clear cell morphology and other morphological features that are relatively frequently observed in metastatic renal cell carcinoma, such as eosinophilic cytoplasm and rhabdoid feature. These results suggest that clear cell morphology due to adipogenesis in CCRCC is associated with low nuclear grade and good prognosis. In this regard, our findings suggest that adipogenic differentiation by histone modification is a new tumorigenic mechanism that reflects the clear cell morphology in CCRCC. In addition, the loss of PHF2 expression seems to be correlated with tumor progression and a poor prognosis of CCRCC due to the loss of adipogenic differentiation.

The role of PHF2 in tumorigenesis is not yet well understood, and only a few studies have examined the association between human cancer and PHF2 expression. The expression of PHF2 in breast cancer was not correlated with patient prognosis. In esophageal squamous cell carcinoma, PHF2 was overexpressed in cancer cells compared to the non-neoplastic epithelium, and high cytoplasmic expression of PHF2 tended to be associated with decreased overall survival of the patients, but there was no statistical significance. In head and neck squamous cell carcinoma, loss of heterozygosity was observed in the chromosomal 9q22 locus located in the PHF2 gene in relation to dysplastic lesions, but it was not related to the clinicopathological index or patient survival rate.

A recent study showed that PHF2 can modulate p53 by demethylating H3K9me2, and down-regulation of PHF2 expression occurs in colon and stomach cancer. In the study, they suggested that PHF2 acts as a tumor suppressor by regulating the function of p53. Even p53-positive colon cancer cells have a malignant phenotype associated with the suppression of p21, a downstream molecule of p53. They also suggested that PHF2 expression levels could help predict patient outcomes in tumors expressing functional p53. These results provide another perspective on the role of PHF2 in CCRCC, although the role of p53 in the progression of CCRCC remains controversial.

Our study showed the clinicopathological significance of PHF2 expression in CCRCC. Low expression of PHF2 is associated with aggressiveness and poor prognosis of CCRCC, and this effect of PHF2 may be related to the role of PHF2 in adipogenesis or the regulation of p53. More functional studies will help clarify the role of PHF2 in the development and progression of CCRCC.

Conflicts of Interest
No potential conflict of interest relevant to this article was reported.

REFERENCES


Yes-Associated Protein Expression Is Correlated to the Differentiation of Prostate Adenocarcinoma

Myung-Giun Noh · Sung Sun Kim · Eu Chang Hwang · Dong Deuk Kwon · Chan Choi

Background: Yes-associated protein (YAP) in the Hippo signaling pathway is a growth control pathway that regulates cell proliferation and stem cell functions. Abnormal regulation of YAP was reported in human cancers including liver, lung, breast, skin, colon, and ovarian cancer. However, the function of YAP is not known in prostate adenocarcinoma. The purpose of this study was to investigate the role of YAP in tumorigenesis, differentiation, and prognosis of prostate adenocarcinoma. Methods: The nuclear and cytoplasmic expression of YAP was examined in 188 cases of prostate adenocarcinoma using immunohistochemistry. YAP expression levels were evaluated in the nucleus and cytoplasm of the prostate adenocarcinoma and the adjacent normal prostate tissue. The presence of immunopositive tumor cells was evaluated and interpreted in comparison with the patients’ clinicopathologic data. Results: YAP expression levels were not significantly different between normal epithelial cells and prostate adenocarcinoma. However, YAP expression level was significantly higher in carcinomas with a high Gleason grades (8–10) than in carcinomas with a low Gleason grades (6–7) (p < .01). There was no statistical correlation between YAP expression and stage, age, prostate-specific antigen level, and tumor volume. Biochemical recurrence (BCR)-free survival was significantly lower in patients with high YAP expressing cancers (p = .02). However high YAP expression was not an independent prognostic factor for BCR in the Cox proportional hazards model. Conclusions: The results suggested that YAP is not associated with prostate adenocarcinoma development, but it may be associated with the differentiation of the adenocarcinoma. YAP was not associated with BCR.

Key Words: Yes-associated protein (YAP); Prostate adenocarcinoma; Immunohistochemistry; Gleason score

Prostate cancer (PC) remains a leading cause of cancer-related death in North-American men and is becoming an increasingly common cancer in South Korea. Pathological stage and Gleason grade are important predictors of prognosis in patients with primary prostate adenocarcinoma who undergo a radical prostatectomy. The Gleason grading system is a well-established system for prostate adenocarcinoma that correlates with the differentiation of the prostate gland, pathological stage, disease progression, and recurrence. However, prostate adenocarcinoma is a remarkably heterogeneous disease. Distinguishing tumors associated with a poor outcome at the time of radical prostatectomy is problematic. The molecular mechanisms of prostate carcinogenesis remain poorly understood. The Hippo pathway, a vital growth regulator of cell proliferation and apoptosis, was first identified by mosaic screens in Drosophila melanogaster. Information about the Hippo pathway in Drosophila is likely applicable directly to mammalian systems, as it has been shown that mammalian homologues are capable of rescuing Drosophila mutants defective in the Hippo signaling pathway. Yes-associated protein (YAP) is a transcriptional coactivator of the Hippo pathway and is a highly conserved component of this pathway in mammalian systems. In humans, amplification of the chromosomal region containing the YAP gene (11q22) has been reported in several tumor types. Recent genetic mouse models and studies with cancer patient demonstrated the critical roles of Hippo-YAP signaling in cancer development. For examples, immunohistochemistry studies have shown that an elevated expression/nuclear localization of YAP or transcriptional coactivator with a PDZ-binding domain (TAZ) correlates with malignant features in lung cancer. In datasets of breast cancer patients, elevated expression of gene signatures for YAP/TAZ activity correlates with high histological grade, enrichment of stem cell signatures, metastasis proclivity, and poor outcome. High expression of YAP activity has been found to be prognostic for bad outcome in four datasets of colorectal cancer patients and correlated with cetuximab resistance.
studies on human hepatocellular carcinoma samples showed that elevated expression of YAP or TAZ correlates with poor tumor differentiation and is prognostic of bad outcome.\textsuperscript{13} By immunohistochemistry on human pancreatic tissue samples, YAP and TAZ were found to be almost absent from normal acini, but moderately expressed and nuclearly localized in PanINs and in a subset of primary pancreatic ductal adenocarcinoma, whereas strong nuclear staining of YAP was found in metastases derived from pancreatic adenocarcinoma.\textsuperscript{14}

Although the net effect of deregulated YAP and TAZ activities in many tissues is similar, their activities appear to be controlled by different regulatory mechanisms in different tissues.\textsuperscript{15} Previous studies demonstrated the biological significance of the Hippo-YAP signaling pathway in prostate adenocarcinoma, but large-scale studies have failed to identify YAP amplification and mutations in castration resistant PCs.\textsuperscript{16} Zhang et al.\textsuperscript{3} showed that the Hippo effector YAP regulates cell motility, invasion, and castration-resistant growth of prostate adenocarcinoma in rats. Hu et al.\textsuperscript{17} showed that YAP expression in PC is inversely correlated with Gleason score. However, the clinical significance of YAP amplification in human prostate adenocarcinoma has largely remained unknown. This study investigates the role of YAP in the development, differentiation, and prognosis of prostate adenocarcinoma.

**MATERIALS AND METHODS**

**Patients and tumor samples**

Prostate acinar adenocarcinoma specimens were obtained from 188 patients who had undergone radical prostatectomy at Chonnam National University Hwasun Hospital from 2005 to 2012. The availability of adequate tissue material was the only inclusion criterion. Diagnostic criteria of prostate acinar adenocarcinoma were in agreement with the World Health Organization classification. Clinicopathologic data were collected from the medical records. All patients were advised to have prostate-specific antigen (PSA) follow-up every 3 months in the first year postoperation and at least biannually thereafter. Biochemical recurrence (BCR) was defined as two consecutive PSA measurements ≥0.2 ng/mL within an interval of more than 3 months. PSA progression-free survival time was defined as the time from radical prostatectomy to the first follow-up date showing PSA ≥0.2 ng/mL or until the last follow-up. This study was approved by the Institutional Review Board of Chonnam National University Hwasun Hospital (CNUHH-2017-022).

**Immunohistochemistry for YAP**

One representative slide of the prostate adenocarcinoma was selected for immunohistochemical staining. Tumor samples obtained during surgical treatment were fixed in formalin and embedded in paraffin for histologic studies. The slides of the tumor surgical specimens were stained with hematoxylin and eosin and were reviewed and representative tissue blocks were selected. YAP (1A12) mouse monoclonal antibody (Cell Signaling Technology, Danvers, MA, USA) at a dilution of 1:200 was used for immunohistochemical analysis. Immunostaining was performed using the avidin-biotin complex method. Briefly, representative paraffin blocks were cut consecutively at 4-μm thickness, and immunohistochemical staining was carried out using a BOND-MAX Automated IHC/ISH Stainer (Leica Biosystems, Wetzlar, Germany).

Sections were deparaffinized in xylene and treated with 0.3% hydrogen peroxide in methanol for 20 minutes to block any endogenous peroxidase activity. Citrate buffer was used for antigen retrieval. Nonspecific binding was limited by using protein blocking buffer for 10 minutes. The sections were washed in phosphate-buffered saline and then incubated with the primary antibody for 20 minutes at room temperature. The samples were then incubated in secondary antibody (biotinylated) for 10 minutes, followed by incubation with streptavidin-horseradish peroxidase for 10 minutes, and exposed to diaminobenzidine, which was used as a chromogen. All labeled streptavidin-biotin-horseradish peroxidase system chemicals were obtained from Dako Cytomation Corp. (Carpinteria, CA, USA). Counterstaining was performed with Mayer’s hematoxylin. Negative controls were treated similarly with the exception of incubation with the primary antibody (nonspecific staining control).

**Assessment of protein expression**

All immunostained slides were evaluated twice by two independent investigators blinded to the clinical details. Five views of the highest expression site were examined per slide, and 100 cells were observed per view at 400× magnification. In each slide, adjacent noncancerous basal cells of the prostate acinar tissue were available as internal positive controls and scattered lymphocytes were used as internal negative controls. YAP expression was graded according to the distribution, intensity, and percentage of positive cells as described previously.\textsuperscript{8} Both the tumor cells and noncancerous acinar cells were graded (Fig. 1). For the cytoplasmic distribution, weak cytoplasmic reactivity was considered as low expression regardless of the extent. Strong cytoplasmic reactivity with less than 50% positive cells was graded as low
expression; otherwise, it was graded as high expression if there were greater than 50% positive cells. For the nuclear distribution, samples with nuclear staining in less than 10% of cells were graded as low YAP expression, and samples with nuclear staining in more than 10% of cells were graded as high YAP high expression. YAP expression levels in prostate tissues were divided into high YAP expression and low/negative YAP expression. Two pathologists reviewed cases of inconclusive samples together and reached an agreement as to the YAP expression level.

**Gene Expression Omnibus data**

Microarray data of prostate adenocarcinoma deposited in the National Center for Biotechnology Information (NCBI) Gene Expression Omnibus (GEO) were analyzed.

**Statistical analysis**

Statistical analyses were performed with the statistical computing environment R (ver. 3.3.2, R Core Team) and the software program IBM SPSS (ver. 21.0 for Windows, IBM Corp., Armonk, NY, USA). Correlation with tumor and adjacent non-tumor tissue in prostate samples was assessed by the Spearman’s rank correlation coefficient test. Association between the Gleason grade and clinical factors on radical prostatectomy specimens with YAP expression was calculated using chi-square test. Statistically significant trends associated with an increasing Gleason grade and YAP expression were analyzed by Cochran Armitage trend test. Multivariate logistic regression analysis was used to develop a trend score for tumor differentiation based on patient characteristics (age, initial PSA tumor volume, YAP expression, and pathologic stage). Kaplan-Meier survival curves were used to

**Fig. 1.** Scoring according to the expression of Yes-associated protein (YAP) in prostate adenocarcinoma. (A) In prostate adenocarcinoma, both nuclei and cytoplasm are strongly positive for YAP, scored as high. (B) Negative expression is noted for YAP in prostate adenocarcinoma cells, the nuclear stain over one tenth is scored as high. (C) YAP expression in normal prostate glands. Basal cells are strongly positive which were used as a positive control. Luminal cells did not show YAP expression. Stromal cells reveal mild expression of YAP.
assess the prognostic significance of YAP in predicting BCR. A multivariate analysis was performed using the Cox regression model to study the effects of different variables on BCR. All p-values were based on two-tailed statistical analyses and p < .05 was considered to be statistically significant. In GEO data, the Mann-Whitney and paired Wilcoxon tests were utilized when parameters were not normally distributed, while independent-samples and paired-sample t tests were used when parameters were normally distributed. The Kolmogorov-Smirnov test was employed to demonstrate deviation from the normal distribution.

RESULTS

YAP expression in 188 cases of prostate adenocarcinoma and their paired adjacent nontumor tissues was investigated by immunohistochemistry. In the cytoplasm of normal prostate luminal cells, 45 cases (23.9%) were completely negative, 125 cases (66.5%) showed low expression in cytoplasm, and 18 cases (9.6%) showed high expression. In the nucleus, 140 cases (74.4%) were negative, 33 cases (17.6%) showed low expression, and 15 cases (8.0%) showed high expression (Table 1, Fig. 2A–D). In the nucleus of

<table>
<thead>
<tr>
<th>Type</th>
<th>Prostate adenocarcinoma</th>
<th>Normal prostate glands adjacent to adenocarcinoma</th>
</tr>
</thead>
<tbody>
<tr>
<td>Complete absence of reactivity</td>
<td>16/188 (8.5)</td>
<td>45/188 (23.9)</td>
</tr>
<tr>
<td>Absent</td>
<td>44/188 (23.4)</td>
<td>95/188 (50.5)</td>
</tr>
<tr>
<td>Low</td>
<td>84/188 (44.7)</td>
<td>33/188 (17.6)</td>
</tr>
<tr>
<td>High</td>
<td>10/188 (5.3)</td>
<td>7/188 (3.7)</td>
</tr>
<tr>
<td>Cytoplasm</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>9/188 (4.8)</td>
<td>0/188 (0)</td>
</tr>
<tr>
<td>Low</td>
<td>124/188 (66.0)</td>
<td>125/188 (66.5)</td>
</tr>
<tr>
<td>High</td>
<td>5/188 (2.7)</td>
<td>10/188 (5.3)</td>
</tr>
<tr>
<td>Nucleus and cytoplasm</td>
<td></td>
<td></td>
</tr>
<tr>
<td>High</td>
<td>34/188 (18.1)</td>
<td>8/188 (4.3)</td>
</tr>
</tbody>
</table>

Values are presented as number (%). YAP, Yes-associated protein.

Fig. 2. Expression of Yes-associated protein (YAP) in prostate adenocarcinoma and normal prostate glands. (A–D) Hematoxylin and eosin (H&E)–stained section of benign glands and immunohistochemical stain of YAP in normal prostate glands. Luminal cells of normal prostate gland show no expression to mild expression of YAP in the cytoplasm. All of the basal cells and stroma show positive YAP.

(Continued to the next page)
prostate adenocarcinoma, 60 cases (31.9%) were negative, 84 cases (44.7%) showed low expression and 44 cases (23.4%) were high expression. In the cytoplasm, 25 cases (13.3%) were negative, 124 cases (66.0%) showed low expression, and 39 cases (20.7%) showed high expression (Table 1, Fig. 2E–H). Basal cells and stroma were positive for YAP (Fig. 2).17

According to the YAP immunohistochemical scoring system used in this study, 49 cases of prostate adenocarcinoma tissues (26.1%) highly expressed YAP in both the nuclei and cytoplasm, whereas 139 cases of adenocarcinoma tissues (73.9%) were negative/low positive for YAP expression (Table 2). In contrast, 163 cases (86.7%) revealed low/negative expression of YAP and only 25 cases of these normal cells (13.3%) were highly positive for YAP expression (Table 2). The expression of YAP was not significantly different between tumor and adjacent normal tissues in the prostate adenocarcinoma samples (p > .05) (Table 2).

YAP mRNA expression level in prostate adenocarcinoma available from GEO profiles was evaluated.18-24 Only one study found YAP expression to be higher in prostate adenocarcinoma than in normal tissue.19 In two other studies, YAP expression was lower in prostate adenocarcinoma than in normal tissue.20,23 There was no significant tendency of YAP RNA expression profiles between prostate adenocarcinoma and normal tissue (Table 3).

Next, the relationship between YAP expression and the clinicopathologic factors were analyzed. For well-to-moderately differentiated adenocarcinoma (Gleason score of 6–7), 118 cases (81.9%) were completely negative or weakly stained (intensity score of “low”) for YAP (p < .01) (Table 4, Fig. 2E, F). In comparison, 23 cases of poorly differentiated adenocarcinoma (Gleason score of 8–10) exhibited strong staining for YAP (intensity score of “high”) (p < .05) (Table 4, Fig. 2G, H).

Fig. 2. (Continued from the previous page) (E) H&E-stained section of well-to-moderately differentiated prostate adenocarcinoma (Gleason score, 4 + 3 = 7). There are poorly formed glands and a few well-formed glands. (F) Expression of YAP in well-to-moderately differentiated prostate adenocarcinoma (Gleason score, 4 + 3 = 7). Tumor cells show either no or mild expression in the cytoplasm. (G) H&E-stained section of poorly differentiated prostate adenocarcinoma (Gleason score, 5 + 5 = 10). (H) Tumor cells display strong positivity in both nuclei and cytoplasm.
score of “high”) (p < .01) (Table 4, Fig. 2G, H). However, no significant difference in YAP expression level was observed according to the age of the patient, tumor volume, preoperative serum PSA, or tumor stage (Table 4). A multivariate logistic regression model showed YAP expression (odds ratio [OR], 9.41; 95% confidence interval [CI], 3.76 to 25.31), initial PSA (OR, 1.10; 95% CI, 1.06 to 1.15), and pathologic stage (OR, 3.58; 95% CI, 1.40 to 9.42) to be significantly correlated with tumor differentiation (Table 5).

The BCR-free survival was significantly lower in patients with high YAP expressing cancers than those with negative/low YAP expressing cancers (p = .02) (Fig. 3). In addition, univariate analysis showed that the Gleason grade (hazard ratio, 5.02; p < .01) and high YAP expression (hazard ratio, 1.919; p = .02) were associated with BCR. Furthermore, multivariate analysis using a Cox regression model demonstrated that the Gleason grade (p < .01) and T stage were independent prognostic factors for BCR. However, YAP expression was not a significantly independent prognostic factor for BCR in the Cox proportional hazards model (Table 6).

**Table 2.** Comparison of YAP expression between adenocarcinoma and normal prostate tissue adjacent to the adenocarcinoma

<table>
<thead>
<tr>
<th>Type</th>
<th>YAP negative/low expression</th>
<th>YAP high expression</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal prostate glands adjacent to adenocarcinoma</td>
<td>163/188 (86.7)</td>
<td>25/188 (13.3)</td>
<td>&gt;.05</td>
</tr>
<tr>
<td>Prostate adenocarcinoma</td>
<td>139/188 (73.9)</td>
<td>49/188 (26.1)</td>
<td></td>
</tr>
</tbody>
</table>

Values are presented as number (%).

YAP, Yes-associated protein.

**Table 3.** YAP mRNA expression in normal tissues and prostate adenocarcinoma from GEO profiles

<table>
<thead>
<tr>
<th>Study</th>
<th>Normal No. of cases</th>
<th>Prostate cancer No. of cases</th>
<th>p-value</th>
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</thead>
<tbody>
<tr>
<td>Varambally et al.18</td>
<td>3,797.54 ± 2,444.70</td>
<td>3,017.58 ± 2,076.74</td>
<td>.29</td>
</tr>
<tr>
<td>Chandran et al.19</td>
<td>44.48 ± 21.65</td>
<td>56.90 ± 36.73</td>
<td>.01</td>
</tr>
<tr>
<td>Yu et al.20</td>
<td>1,280.71 ± 445.43</td>
<td>878.24 ± 414.46</td>
<td>&lt;.01</td>
</tr>
<tr>
<td>Satake et al.21</td>
<td>1,984.20 ± 1,985.80</td>
<td>1,254.13 ± 1,500.77</td>
<td>.42</td>
</tr>
<tr>
<td>Planche et al.22</td>
<td>8.43 ± 1.54</td>
<td>8.05 ± 1.75</td>
<td>.11</td>
</tr>
<tr>
<td>Arredouani et al.23</td>
<td>1,139.21 ± 806.37</td>
<td>754.87 ± 496.63</td>
<td>.02</td>
</tr>
<tr>
<td>Nanni et al.24</td>
<td>190.02 ± 98.42</td>
<td>189.80 ± 64.38</td>
<td>.98</td>
</tr>
</tbody>
</table>

Values are presented as mean±standard deviation.

YAP, Yes-associated protein; GEO, Gene Expression Omnibus.

**Table 4.** Clinicopathologic features of the patients

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>No. of patients</th>
<th>Low YAP expression</th>
<th>High YAP expression</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yr)</td>
<td></td>
<td></td>
<td></td>
<td>.74</td>
</tr>
<tr>
<td>&lt;70</td>
<td>105</td>
<td>79 (75.2)</td>
<td>26 (24.8)</td>
<td></td>
</tr>
<tr>
<td>≥70</td>
<td>83</td>
<td>60 (72.3)</td>
<td>23 (27.7)</td>
<td></td>
</tr>
<tr>
<td>Initial PSA (ng/mL)</td>
<td></td>
<td></td>
<td></td>
<td>.44</td>
</tr>
<tr>
<td>≤4.0</td>
<td>7</td>
<td>5 (71.4)</td>
<td>2 (28.6)</td>
<td></td>
</tr>
<tr>
<td>4.1–10.0</td>
<td>103</td>
<td>77 (74.8)</td>
<td>26 (25.2)</td>
<td></td>
</tr>
<tr>
<td>10.1–20.0</td>
<td>44</td>
<td>29 (65.9)</td>
<td>15 (34.1)</td>
<td></td>
</tr>
<tr>
<td>&gt;20</td>
<td>34</td>
<td>28 (82.4)</td>
<td>6 (17.6)</td>
<td></td>
</tr>
<tr>
<td>Tumor volume (g)</td>
<td></td>
<td></td>
<td></td>
<td>.22</td>
</tr>
<tr>
<td>&lt;15</td>
<td>122</td>
<td>94 (77.0)</td>
<td>28 (23.0)</td>
<td></td>
</tr>
<tr>
<td>≥15</td>
<td>66</td>
<td>45 (68.2)</td>
<td>21 (31.8)</td>
<td></td>
</tr>
<tr>
<td>Tumor differentiation</td>
<td></td>
<td></td>
<td></td>
<td>&lt;.01&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>Moderate to well differentiation (Gleason score 6–7)</td>
<td>144</td>
<td>118 (81.9)</td>
<td>26 (18.1)</td>
<td></td>
</tr>
<tr>
<td>Poor differentiation (Gleason score 8–10)</td>
<td>44</td>
<td>21 (47.7)</td>
<td>23 (52.3)</td>
<td></td>
</tr>
<tr>
<td>Pathologic stage</td>
<td></td>
<td></td>
<td></td>
<td>.69</td>
</tr>
<tr>
<td>pT2</td>
<td>146</td>
<td>109 (74.7)</td>
<td>37 (25.3)</td>
<td></td>
</tr>
<tr>
<td>pT3</td>
<td>42</td>
<td>30 (71.4)</td>
<td>12 (28.6)</td>
<td></td>
</tr>
</tbody>
</table>

Values are presented as number (%).

YAP, Yes-associated protein; PSA, prostate-specific antigen.

<sup>a</sup>Cochran Armitage trend test, p < .01.
YAP regulates cell proliferation, stem behavior and regeneration.15 Promotion of cell stemness and proliferation are important factors for cancer development and regeneration.15 YAP can be used during the development of cancer and other phenotypes during reprogramming of mature and differentiated cells.15

As noted above, the expression of YAP via immunohistochemical staining showed no statistically significant difference between prostate adenocarcinoma and adjacent normal tissues. Similarly, in GEO data profiles, YAP mRNA was not statistically higher in prostate adenocarcinoma than in normal tissue. This suggested that YAP expression is not associated with tumorigenesis of prostate adenocarcinoma. YAP and/or TAZ knockdown in human colorectal cancer cell lines suppresses their growth and ability to trigger tumor formation after injection in mice.25 On the other hand, YAP has been reported to be functionally implicated to tumorigenesis in other human cancers. Liver-specific YAP overexpression in transgenic mice leads to hepatomegaly and development of human hepatocellular carcinoma.26 Similar results were obtained by delivering a phospho-mutant form of YAP in the mouse liver using transposon-mediated hydrodynamic transfection.27 YAP or TAZ knockdown strongly reduces subcutaneous tumor growth of human and mouse hepatocellular carcinoma cell lines.28 Functionally, pancreas-specific YAP knockout abrogates tumor progression from early pancreatic intraepithelial neoplasia lesions to pancreatic ductal adenocarcinoma in a mouse model of pancreatic cancer.29

Poorly differentiated prostate adenocarcinoma cells stained more strongly for YAP than in moderately to well-to-moderately differentiated prostate adenocarcinoma. YAP immunoreactivity was inversely associated with the differentiation of prostate adenocarcinoma. In addition, YAP expression was found to be a factor affecting tumor differentiation. This finding indicates that there is a meaningful inverse relationship between YAP expression and differentiation of prostate adenocarcinoma, which suggests that YAP may play a role in the differentiation of prostate adenocarcinoma.

Hu et al.17 showed that down-regulation of YAP expression in prostate adenocarcinoma correlates with an increase in Gleason score, and this study showed differences from the results. It can be explained for several factors. First, the expression of YAP showed intratumoral heterogeneity in prostate adenocarcinoma. In tumors with heterogeneous expression, many tissue cores are

Table 5. Significant risk factors for tumor differentiation in patients with prostate adenocarcinoma determined by multivariate logistic regression model

<table>
<thead>
<tr>
<th>Factor</th>
<th>Multivariate Odds ratio (95% CI) p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age</td>
<td>1.24 (0.54–2.89) .61</td>
</tr>
<tr>
<td>Initial PSA</td>
<td>1.10 (1.06–1.15) &lt;.01</td>
</tr>
<tr>
<td>Tumor volume</td>
<td>1.08 (0.44–2.64) .87</td>
</tr>
<tr>
<td>YAP expression</td>
<td>9.41 (3.76–23.51) &lt;.01</td>
</tr>
<tr>
<td>Pathologic stage</td>
<td>3.58 (1.40–9.42) &lt;.01</td>
</tr>
</tbody>
</table>

CI, confidence interval; PSA, prostate specific antigen; YAP, Yes-associated protein.

Fig. 3. Kaplan-Meier estimates of biochemical recurrence. Graph of the curve for progression-free survival in prostate adenocarcinoma patients, according to Yes-associated protein (YAP) expression (p = .02).

Table 6. Univariate and multivariate analyses for predictive factors in patients with prostate adenocarcinoma

<table>
<thead>
<tr>
<th>Factor</th>
<th>Univariate Hazard ratio (95% CI) p-value</th>
<th>Multivariate Hazard ratio (95% CI) p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gleason grade group</td>
<td>5.02 (2.97–8.50) &lt;.01</td>
<td>4.33 (0.56–1.87) &lt;.01</td>
</tr>
<tr>
<td>T stage</td>
<td>2.56 (1.49–4.39) &lt;.01</td>
<td>1.75 (2.39–7.85) .05</td>
</tr>
<tr>
<td>High YAP expression</td>
<td>1.92 (1.11–3.32) .02</td>
<td>1.03 (0.99–3.08) .93</td>
</tr>
</tbody>
</table>

CI, confidence interval; YAP, Yes-associated protein.

https://doi.org/10.4132/jptm.2017.05.04
required to get comprehensive information. Hu et al.\textsuperscript{17} used tissue microarray (TMA) slides with two cores of intratumoral and peritumoral tissue per slide. In this study, we used a representative slide instead of TMA slide. Second, the interpretation method of YAP expression was different. It was estimated according to the distribution and intensity of positive staining in this study according to Steinhardt et al.\textsuperscript{8} However Hu et al.\textsuperscript{17} interpreted it as three categories, only cytoplasmic, only nuclear, both nuclear and cytoplasmic. Those factors might affect the difference of the results of this study and Hu et al.\textsuperscript{17}

Enhanced YAP activity tends to promote stem cells and progenitor cells, while inhibiting differentiation.\textsuperscript{30} Immunohistochemical studies on human hepatocellular carcinoma samples showed that elevated expression of YAP or TAZ correlates with poor differentiation of cancer cells and poor outcomes.\textsuperscript{31} In datasets of patients with breast cancer, an elevated expression of gene signatures for YAP/TAZ activity correlate with high histological grades, enrichment of stem cell signatures, tendency to metastasize, and poor outcomes.\textsuperscript{10,32} Prostate stem cell antigen expression identified through immunohistochemical staining and in situ hybridization is significantly increased in human PCs with a high Gleason score.\textsuperscript{35} Considering that YAP is generally associated with cancer stem cells, we can further compare the expression of YAP and prostate stem cell antigens with differentiation stages of prostate adenocarcinoma. Once the relationship between PC stem cells and YAP and its mechanisms of action are revealed, it could be further explored as a new therapeutic target.

In this study, YAP immunoreactivity was not associated with the BCR of prostate adenocarcinoma. The univariate analysis was statistically significant, but not the multivariate analysis. This difference between the univariate and multivariate analyses might be attributed to the contribution of the Gleason score. Therefore, YAP immunoreactivity for prostate adenocarcinoma was not useful for predicting adverse outcomes. Expression of YAP in a variety of human cancers has been reported to be associated with a poor prognosis. High levels of YAP expression were found to be associated with poor outcomes in four datasets of colorectal cancer patients, and were positively correlated with cetuximab resistance.\textsuperscript{34} YAP mRNA and protein levels have been shown to be upregulated in gastric adenocarcinoma, and YAP protein expression and nuclear localization were associated with poor patient outcomes.\textsuperscript{35}

There are several limitations to this study. First, this study investigated the relationship between YAP expression and clinicopathologic factors in prostate adenocarcinoma based on the morphological expression of the cancer. No functional studies of YAP have been performed in relation to tumorigenesis of prostate adenocarcinoma. In addition, this study did not show a direct functional relationship between YAP expression and the differentiation pathway of prostate adenocarcinoma. The second limitation was that the tumor differentiation was interpreted in a two-tier system: well to moderate differentiation (Gleason score of 6–7) or poorly-differentiation (Gleason score of 8–10). The reason for this was that the number of patients subclassified by their Gleason score was largely uneven. The number of patients with Gleason score of 6 was 39, of 7 was 105, of 8 was 26, of 9 was 17, and of 10 was 1. If a larger cohort of patients was included in this study, the study might have had more specific results.

This study demonstrates that YAP is highly expressed in poorly differentiated prostate adenocarcinoma compared to well-differentiated or moderately differentiated prostate adenocarcinoma. However, YAP expression was not associated with BCR in the Cox proportional hazards model.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

REFERENCES

7. Lai ZC, Wei X, Shimizu T, et al. Control of cell proliferation and
Basaloid Squamous Cell Carcinoma of the Head and Neck: Subclassification into Basal, Ductal, and Mixed Subtypes Based on Comparison of Clinico-pathologic Features and Expression of p53, Cyclin D1, Epidermal Growth Factor Receptor, p16, and Human Papillomavirus

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Basaloid squamous cell carcinoma (BSCC) of the head and neck is a rare variant of squamous cell carcinoma (SCC) with distinct pathologic characteristics with multipotential differentiation.1 Histologic characteristics of BSCC include solid nesting, comedonecrosis, cribriform pattern, trabecular arrangement, ductal differentiation, and associated conventional SCC, most frequently seen in situ. The histogenesis of BSCC is not fully understood, and the cancer has been suggested to originate from a totipotent primitive cell in the basal cell layer of the surface epithelium or in the proximal duct of secretory glands.2 The incidence of ductal differentiation in BSCC has been reported to range from 27.5%–45%3,4 and was associated with significantly better survival than those without ductal differentiation in a study by Imamhasan et al.5 However, the clinical significance of the differentiation profiles of BSCC has not otherwise been studied. Most studies on BSCC cytokeratin (CK) immunoprofiles have focused on diagnostic value and the ability to distinguish BSCC from other carcinomas with basaloid features.5–12 CK14 was the most consistently expressed CK in BSCC,3,5,6,10–12 which is similar to the pattern seen in conventional SCC.4 Nonetheless, considering the histologic diversity of BSCC, CK immunoprofiles of BSCC are expected to be heterogeneous and not unique. The aim of this study was to examine expression patterns of basal and ductal immunomarkers, including variable CKs, in BSCC of the head and neck; to subclassify them according to basal or ductal immunomarker expression; and to compare their histology, other oncogene expression, human pap-
illomavirus (HPV) status, and clinical features.

MATERIALS AND METHODS

Immunohistochemistry and in situ hybridization

Twenty-six cases of head and neck BSCC from Asan Medical Center, Seoul, Korea, reported during a 14-year period were subject to immunohistochemical study for CK5/6 (1:200, Zymed, San Francisco, CA, USA), CK7 (1:400, Dako, Glostrup, Denmark), CK8/18 (1:200, Cell Marque, Rocklin, CA, USA), CK19 (1:100, Cell Marque), p63 (1:200, Dako), p53 (1:3,000, Glostrup, cyclin D1 (1:100, Neomarkers, Fremont, CA, USA), epidermal growth factor receptor (EGFR; 1:200, Invitrogen, Carlsbad, CA, USA), p16INK4 (1:10, Pharmingen, Franklin Lakes, NJ, USA) and in situ hybridization for HPV. Immunostaining was carried out on 4-μm sections using a Ventana autostainer and an ultra-view DAB detection kit (Ventana Medical Systems Inc., Tucson, AZ, USA), according to the manufacturer’s instructions. Cytoplasmic staining of CK5/6, CK7, CK8/18, and CK19 and nuclear staining of p63, p53, cyclin D1, and p16INK4 were regarded as positive if diffusely or heterogeneously staining was noted in ≥ 20% of the tumor cells. EGFR staining was regarded as positive if ≥ 10% of tumor cells showed membranous staining with strong or intermediate intensity. For HPV in situ hybridization, the INFORM HPV III Family 16 Probe (B) was used in conjunction with the ISH iView Blue Plus Detection Kit (Ventana Medical Systems Inc.). The INFORM HPV III Family 16 Probe (B) detects the following high-risk HPV genotypes: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, and 66. Light microscopy results were analyzed, and any blue nuclear dots in the tumor cells were regarded as positive staining.

Clinicopathologic analysis

Cases that expressed either one or two basal (CK5/6 and p63) or ductal (CK7 and CK8/18) markers were considered as basal or ductal subtypes, and expression of both basal and ductal markers was subclassified into a mixed subtype. The case histologies were re-evaluated for the presence of nuclear palisading, comedo-necrosis, cribriform pattern, ductal differentiation, mucin production, and desmoplasia. Clinical information, including survival period of the patients, was obtained via medical records. The average follow-up period for all patients was 79.5 months.

For descriptive statistics, all categorical data were compared with the chi-square test. Continuous data were expressed as mean values and were compared using ANOVA. A survival curve was estimated using the Kaplan-Meier method, and differences in survival between groups were compared using the log-rank test. p < .05 was considered statistically significant.

RESULTS

Expression of basal and ductal markers

Positive rates for basal (CK5/6; p63) and ductal markers (CK7; CK8/18) in 26 BSCC are summarized in Table 1. Overall positive rates for CK5/6, p63, CK7, and CK8/18 were 65.4%, 73.1%, 34.6%, and 46.2%, respectively. Twelve combination patterns were observed, and BSCCs were subdivided into basal (nine cases), ductal (five cases), and mixed (12 cases) subtypes (Fig. 1). Clinical features of the 26 cases based on subtype are summarized in Table 2. Advanced age and male gender were more frequently observed in the mixed subtype than in the basal and ductal subtypes. While 60% of laryngeal or hypopharyngeal BSCC was classified as the mixed subtype, 60% of oropharyngeal BSCC was determined to be the basal subtype. BSCC was the cause of death in 19 of the 26 patients. Overall survival periods were not significantly different among the three subtypes (Fig. 2).

Histologic parameters

The relationships between histologic parameters and subtypes are listed in Table 3. Nuclear palisading was observed in both basal and ductal subtypes, but not in the mixed subtype (p = .010). Comedo-necrosis was most commonly observed in the basal subtype, followed by ductal and mixed subtypes (p = .001). Ductal differentiation or mucin production was observed not only in the ductal subtype but also in the basal and mixed subtypes. Desmoplasia was most common in the basal subtype, followed by ductal and mixed subtypes (p = .021). Representative histo-
logic features are shown in Fig. 3.

Expression of CK19, p53, cyclin D1, EGFR, p16, and HPV

The expression rates of other tested markers in the three subgroups are compared in Table 4 and illustrated in Fig. 4. CK19, which normally exists in the basal layers of both squamous epithelium and various ductal structures, was positive in 42.3% of cases and was most commonly identified in the mixed subtype, followed by ductal and basal subtypes (p > .05). Overexpression of p53 was observed in 34.6% of cases and was more commonly observed in mixed than in basal or ductal subtypes (p > .05). Cyclin

DISCUSSION

BSCC is a type of squamous cell carcinoma that occurs in the supraglottic larynx, pyriform sinus, base of tongue, or esophagus. The main histologic components of BSCC are undifferentiated
cells, but there can be divergent differentiation, including ductal differentiation. The multipotency of BSCC has been described more often with an esophageal origin than in the head and neck.1,3,4

Several studies on the CK immunoprofiles of BSCC have been reported, based on diagnoses in the head and neck region, and most of them focused on the diagnostic value and distinguishing BSCC from other carcinomas with basaloid features.5,6,8-12 The most frequently expressed CK subtype in BSCC was CK14 (53%–100%),6,8,10,11 and CK5/6 (100%),7 followed by CK19 and CK8/18 (80%),9 and CK17 (68%).8 These have been described as useful markers for differentiating BSCC from other histologically similar tumors including basal cell carcinoma with squamous metaplasia, adenoid cystic carcinoma, and neuroendocrine carcinoma. The CK7 expression rate has varied depending on the report (0%–53%).8,10,11,13,14

Based on the variable histology and CK expression of BSCC, we hypothesized that this tumor is heterogeneous and not homogenous. Kobayashi et al.3 also concentrated on the histologic diversity of esophageal BSCC. They analyzed expression patterns of CK7, CK14, and smooth muscle actin in relationship to different components of BSCCs. However, our attempt to subclassify BSCC according to the expression of basal (CK5/6, p63) or ductal markers (CK7, CK8/18) is novel.

Imamhasan et al.4 studied 22 esophageal BSCCs in comparison with conventional SCC. BSCC frequently manifested with ductal differentiation (27.5%), and cases with ductal differentiation were associated with better outcomes than cases without ductal differentiation, although this result was not statistically significant. Ductal differentiation was observed in eight of 26 cases (30.8%)

Table 3. Histologic features of head and neck basaloid squamous cell carcinoma according to subtype

<table>
<thead>
<tr>
<th>Histologic finding</th>
<th>Basal (n=9)</th>
<th>Ductal (n=5)</th>
<th>Mixed (n=12)</th>
<th>Total (n=26)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Palisading</td>
<td>4 (44.4)</td>
<td>3 (60.0)</td>
<td>0</td>
<td>7 (26.9)</td>
</tr>
<tr>
<td>Comedonecrosis</td>
<td>9 (100)</td>
<td>3 (60.0)</td>
<td>3 (25.0)</td>
<td>15 (57.7)</td>
</tr>
<tr>
<td>Cribriform pattern</td>
<td>2 (22.2)</td>
<td>3 (25.0)</td>
<td>1 (8.3)</td>
<td>6 (23.1)</td>
</tr>
<tr>
<td>Ductal differentiation</td>
<td>4 (44.4)</td>
<td>2 (40.0)</td>
<td>1 (8.3)</td>
<td>7 (26.9)</td>
</tr>
<tr>
<td>Mucin production</td>
<td>2 (22.2)</td>
<td>3 (25.0)</td>
<td>1 (8.3)</td>
<td>6 (23.1)</td>
</tr>
<tr>
<td>Desmoplasia</td>
<td>4 (44.4)</td>
<td>2 (40.0)</td>
<td>0</td>
<td>6 (23.1)</td>
</tr>
</tbody>
</table>

Values are presented as number (%). *p = .010; **p = .001; ***p = .021.

Table 4. Other immunomarker expression in head and neck basaloid squamous cell carcinoma according to subtype

<table>
<thead>
<tr>
<th>Immunomarker</th>
<th>Basal (n=9)</th>
<th>Ductal (n=5)</th>
<th>Mixed (n=12)</th>
<th>Total (n=26)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CK19</td>
<td>2 (22.2)</td>
<td>2 (40.0)</td>
<td>7 (58.3)</td>
<td>11 (42.3)</td>
</tr>
<tr>
<td>p53</td>
<td>2 (22.2)</td>
<td>1 (20.0)</td>
<td>6 (50.0)</td>
<td>9 (34.6)</td>
</tr>
<tr>
<td>Cyclin D1</td>
<td>7 (77.8)</td>
<td>2 (40.0)</td>
<td>9 (75.0)</td>
<td>18 (69.2)</td>
</tr>
<tr>
<td>EGFR</td>
<td>6 (66.7)</td>
<td>1 (20.0)</td>
<td>9 (75.0)</td>
<td>16 (61.5)</td>
</tr>
<tr>
<td>p16</td>
<td>1</td>
<td>2</td>
<td>1</td>
<td>4 (15.4)</td>
</tr>
<tr>
<td>HPV</td>
<td>1</td>
<td>2</td>
<td>0</td>
<td>3 (11.5)</td>
</tr>
</tbody>
</table>

Values are presented as number (%).

CK, cytokeratin; EGFR, epidermal growth factor receptor; HPV, human papillomavirus.

Fig. 3. Representative histologic parameters of basaloid squamous cell carcinomas. (A) Nuclear palisading. (B) Comedo-necrosis. (C) Cribriform pattern. (D) Ductal differentiation. (E) Mucin production. (F) Desmoplasia.
in our study. Ductal differentiation or mucin production was not restricted to the ductal subtype but was associated with basal or mixed subtypes. Ductal differentiation and ductal phenotypes were not significantly associated with improved prognoses. There was no significant difference in overall survival among the three subtypes. Among other histologic parameters, peripheral nuclear palisading, comedo-necrosis, and desmoplasia were significantly less frequent in the mixed subtype than in the basal or ductal phenotype. However, the clinical significance of this result was not fully elucidated.

When expression rates for other immunomarkers were compared, CK19, which can be both a basal and ductal marker, was positive in 42.3% of cases, which was lower than the rate of CK5/6 (65.4%) or CK8/18 (46.2%) expression. The p53 expression rate in head and neck BSCC was 34.6%, which was slightly lower than reported rates of 40%–100%. CyclinD1 and EGFR were positive in 69.2% and 61.5%, respectively. There are currently no reported data on expression patterns of these proteins in head and neck BSCC. A few studies on esophageal BSCC showed 25% cyclinD1 positivity and 56%–100% EGFR positivity. The ductal subtype tended to exhibit lower levels of p53, cyclinD1, and EGFR expression than basal or mixed subtypes; however,

Fig. 4. Representative expression patterns of cytokeratin 19 (A), p53 (B), cyclinD1 (C), epidermal growth factor receptor (D), p16 (E), and human papillomavirus in situ hybridization (F).
based on the small number of cases in this study, the result was not statistically significant. A small number of p16- and/or HPV-positive cases were not restricted to one subtype. Three HPV-positive cases consisted of one basal subtype from the tonsil, one ductal subtype from the tonsil, and one ductal subtype from the nasal cavity. In the past, a relationship between HPV and basaloid morphology of head and neck squamous cell carcinoma has been suggested, but it is now accepted that not all BSCC cases are associated with HPV, and HPV status is more closely related to the location than histology of SCC. A recent study suggested a relationship between high-risk HPV and CK19 expression, but two of three HPV-positive cases in this study were negative for CK19.

In summary, BSCC consists of a heterogeneous group that contain tumors with basal, ductal, or mixed immunophenotypes, and this phenomenon suggests a complex histogenesis. Mixed subtypes showed fewer histological characteristics, and the ductal subtype showed less oncogenic protein expression (p53, cyclin D1, and EGFR), although the clinical implications of these findings have not been fully elucidated. The low expression rate of BSCC for CK19 contrasts with the high expression in conventional SCC and limits the diagnostic utility of CK19 mRNA detection in this entity.

Although this study suggested a heterogeneous pathogenesis of head and neck BSCC, there were some limitations to this study, including the retrospective design, small number of cases, and the fact that all patients came from a single institution. Extended or multi-institutional studies might produce more significant results. Furthermore, remarkable advances in investigative tools, such as genomic microarray technologies and next-generation sequencing, could help confirm the results.

**Conflicts of Interest**

No potential conflict of interest relevant to this article was reported.

**REFERENCES**


Morphological Features and Immunohistochemical Expression of p57Kip2 in Early Molar Pregnancies and Their Relations to the Progression to Persistent Trophoblastic Disease

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Background: Although the morphological features characteristic of products of conception specimens including molar pregnancies are well described, substantial histopathological similarities are observed between the different entities, especially in cases of early pregnancies. Furthermore, there are no current solid criteria that could predict cases with progression to persistent gestational trophoblastic disease. In this study, we aimed to determine the most specific histopathological and immunohistochemical features required for accurate diagnosis that can reliably predict the clinical behavior. Methods: Sixty-five cases of products of conception were reviewed clinically and pathologically, and any progression to persistent gestational trophoblastic disease (GTD), if present, was noted. Pathological assessment of the archival material included re-cut sections of 5 μm in thickness, routine staining with hematoxylin and eosin and immunohistochemical staining of p57Kip2. Results: Certain histopathological criteria were found to be significant in differentiation between complete hydatidiform mole (CHM) and partial hydatidiform mole including villous shape and outline, villous trophoblast hyperplasia, and atypia in extravillous trophoblasts. There were no significant differences in any morphological or immunohistochemical features between cases with or without subsequent development of GTD. Conclusions: Histopathological diagnosis of molar pregnancy remains problematic especially in early gestation. Their diagnosis should be stated after a constellation of specific histopathological criteria in order not to miss CHM. p57Kip2 immunohistochemistry is of great value in diagnosis of cases that had equivocal morphology by histopathological examination. However, there were no significant features to predict cases that subsequently developed persistent GTD.

Key Words: Hydatidiform mole; Complete hydatidiform mole, early; p57Kip2 immunohistochemistry

Hydatidiform mole (HM) is an abnormal gestation characterized by significant hydropic change and variable trophoblastic proliferation involving part or all of chorionic villi.1,2 HM is categorized into two separate entities, complete hydatidiform mole (CHM) and partial hydatidiform mole (PHM), according to morphology and cytogenetics. However, hydropic abortion (HA) could morphologically mimic HM.3,4

Characteristic morphologic features have been proposed for the diagnosis of product of conception (POC) specimens including CHM, PHM, and HA. However, substantial histopathological similarities are observed between the three entities, especially in cases of early detection resulting in interobserver and intraobserver variability in the diagnosis of POC specimens.5,6 Thus, the diagnosis of POC on the basis of histopathology alone remains a challenge for pathologists, even those experienced.7

Persistent gestational trophoblastic disease (PGTD) develops after CHM in 10% to 30% of cases,4 and after PHM in 0.5% to 5% whereas HA has no relation to PGTD.3 Thus, follow-up serum β-human chorionic gonadotropin (β-HCG) measurements aren’t essential in cases of HA, whereas these measurements form part of surveillance for PGTD in cases of HM. Therefore, accurate distinction of CHM from PHM on one hand and of HM from HA on the other hand is important for appropriate clinical management and has prognostic implications.3

Difficult cases may require molecular techniques using the differences in DNA content of different POC cases.8 However, such molecular diagnostic methods are technically difficult, relatively costly, and non-available in most of pathology laboratories.2 Detection of the expression of gene products such as p57Kip2 by the trophoblastic cells should be highlighted. The p57Kip2 gene (CDKN1C) is a strongly paternally imprinted gene expressed only by the paternal allele in most tissues and is involved in implan-
In normal placenta, nuclear p57Kip2 expression is seen in villous cytotrophoblast, extravillous trophoblast, villous stromal cells, and decidua. Incorporation of p57Kip2 can identify CHM (androgenetic diploidy) by the lack of its expression and can be helpful in distinguishing CHM from PHM and non molar HA, but can’t distinguish PHM (diandric monogynic triploidy) from nonmolar (biparental diploidy) specimens as both are positive. CHM mostly doesn’t contain maternal genome. Therefore, p57Kip2 staining is unexpressed or greatly reduced in the nuclei of their cytotrophoblasts and stromal cells. Products of conception containing maternal genetic material, PHM and nonmolar HA, show positive nuclear p57Kip2 staining in cytotrophoblast and villous stromal cells. p57Kip2 is also expressed in intermediate trophoblast islands and decidual cells, serving as positive internal control in all POC cases.

In this study, we aimed to define precise histopathological and immunohistochemical features to make a proper diagnosis of the different types of HMs especially in early gestational ages and to distinguish those from other mimics. Furthermore, specific features to predict prognosis and progression to PGTD were investigated.

MATERIALS AND METHODS

Sample collection
Archival materials of sixty five cases of POC specimens were retrieved from the department of Pathology, Faculty of Medicine, Mansoura University, between January 2013 and December 2014. The protocol was approved by the Ethical Committee of Mansoura University and informed consents were obtained from the patients.

Histopathological review
The paraffin blocks were re-cut in 5-μm-thick sections and stained with hematoxylin and eosin stain and were independently reviewed by two pathologists for evaluation of the main morphological findings of HM.

Immunohistochemistry
Five-micrometer-thick tissue section from each case was subject to immunohistochemical staining using monoclonal antibody against the p57Kip2 protein (NeoMarkers/Lab Vision Corporation, Fremont, CA, USA) with dilution 1:200 and the Envision system (DakoCytomation, Glostrup, Denmark). All the steps were performed according to the manufacturers’ instructions.

In all cases, p57Kip2 was assessed in the nuclei of the villous cytotrophoblasts, extravillous trophoblasts, and villous stromal cells. Specimens were interpreted as positive for p57Kip2 staining when there was distinct nuclear staining of villous cytotrophoblasts and stromal cells. The p57Kip2 stain was interpreted as negative when there was no distinct staining or limited nuclear staining (< 10%) of villous cytotrophoblasts and stromal cells. Staining of intermediate trophoblasts and/or maternal decidua was considered as the positive internal control for these specimens.

Statistical methods
Data were analyzed using the program SPSS ver. 20 (IBM Corp., Armonk, NY, USA) to obtain descriptive statistics. Statistical significance was determined at 95% level of confidence (i.e., differences will be considered significant if p < .05).

RESULTS

Examination of clinical data
The age of patients ranged from 16 to 50 years with a mean of 26 ± 8 years and a median of 24 years. All cases were in the first or early second trimester. Their gestational age ranged from 6 to 14 weeks with a mean of 9 ± 2 weeks and a median of 8 weeks.

Histopathological examination
Significant histopathological criteria for the diagnosis and distinguishing different entities are shown in Table 1. Examples of the morphological features are demonstrated in Fig. 1.

Immunohistochemistry
Specimens that showed distinct nuclear staining of cytotrophoblasts and villous stromal cells were interpreted as positive for p57Kip2 (Fig. 1H). On the other hand, specimens that showed no or limited nuclear staining (in < 10%) of cytotrophoblasts

Table 1. Significant histopathological criteria according to the final diagnosis of the studied cases using p57Kip2 immunohistochemistry

<table>
<thead>
<tr>
<th>Significant criteria differentiating between CHM and PHM</th>
<th>Significant criteria differentiating between molar and nonmolar pregnancy</th>
</tr>
</thead>
<tbody>
<tr>
<td>Villous shape and outline: p &lt; .001</td>
<td>Villous shape and outline: p &lt; .001</td>
</tr>
<tr>
<td>Villous trophoblast hyperplasia: p = .001</td>
<td>Cistern formation: p &lt; .001</td>
</tr>
<tr>
<td>Atypia at extravillous trophoblast: p &lt; .001</td>
<td>Trophoblastic inclusion: p = .001</td>
</tr>
<tr>
<td>Villous trophoblast hyperplasia: p &lt; .001</td>
<td></td>
</tr>
<tr>
<td>CHM, complete hydatidiform mole; PHM, partial hydatidiform mole.</td>
<td></td>
</tr>
</tbody>
</table>
and villous stromal cells with positive internal control were considered negative for p57Kip2 stain (Fig. 1). Accordingly, the studied cases were redistributed as shown in Table 2.

CHM originally constituted 39 cases. However, after reevaluation based on the histopathological criteria and p57Kip2 immunohistochemistry, seven cases were added to CHM (previously diagnosed as PHM). On the other hand, one case originally diagnosed as CHM was converted to PHM. The final typing of the molar cases was, therefore, 45 cases of CHM and 11 cases of PHM.

Follow-up

By following the clinical history of all studied cases, nine cases (14% of total/16% of molar cases) were found to progress into gestational trophoblastic disease (GTD) based on persistence of symptoms and serum $\beta$-HCG level (Table 3). These represented 18% of CHM and 9% of PHM cases. As shown in Table 4, histopathological criteria of these cases were examined and compared with the results of p57Kip2 immunohistochemistry to detect the most statistically significant histopathological feature that can predict the progression of molar disease into PGTD. According to the current study, none of the studied histopathological parameters could differentiate between cases with or without progression to GTD.

Table 2. Comparison of the diagnosis of the studied cases by histopathological examination and p57Kip2 immunohistochemistry ($n = 56$)

<table>
<thead>
<tr>
<th>Method of diagnosis</th>
<th>Case</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Haematoxylin and eosin stain</td>
<td>39 (60)</td>
<td>17 (26)</td>
</tr>
<tr>
<td>p57Kip2 Immunohistochemistry</td>
<td>45 (69)</td>
<td>11 (17)</td>
</tr>
</tbody>
</table>

Values are presented as number (%). CHM, complete hydatidiform mole; PHM, partial hydatidiform mole.
DISCUSSION

The present study was performed to investigate histopathological parameters commonly used during routine histopathological examination in the differential diagnosis of POC cases, especially those with early gestational age. Also, we aimed to study the value of using these histopathological criteria for diagnosis of HM specimens by comparing them with the results of p57Kip2 immunohistochemical staining.

In agreement with previous reports, our results showed that examination of the villous shape was statistically significant in differentiation between CHM and PHM cases on one hand and between molar and nonmolar cases on the other hand.16,17 Most of our studied CHM cases (62%) had a population of distended villi with irregular outline. This was significant in differentiation between POC specimens and in favor of early CHM.15,18 Our results were in contrast with the results of the study by Ishikawa et al.,16 who found that enlarged villi with regular round outline was a good marker for the diagnosis of CHM. This disagreement can be explained by the fact that the gestational age of our cases was in the first trimester, and at this early gestational age, villi of CHM don’t exhibit the well formed distended villi with regular outline and have more irregular outline.

Cistern formation is a major criterion in HM. Our study showed that cistern formation was found in CHM (96%) more common than in PHM cases (91%). It was significant in differentiation between molar and nonmolar pregnancy. On the other hand, it wasn’t significant in differentiation between the CHM and PHM. This is in contrast to other studies that found it significant and in favor of diagnosis of PHM.17,18 This may be because of the difference in number of studied CHM cases.

We found that the presence of trophoblastic inclusion in the
villus core was present in 82% of PHM and in 67% of CHM. This was statistically significant in differentiation between cases of molar and nonmolar pregnancy but was insignificant in differentiation between the CHM and PHM as described in previous reports. Trophoblastic hyperplasia is an essential requirement for the diagnosis of molar pregnancy. The results of this study demonstrated that 59% of CHM cases had circumferential villous trophoblastic hyperplasia, while 88% of PHM cases exhibited multifocal villous trophoblast hyperplasia. Polar trophoblast hyperplasia was in favor of nonmolar POC diagnosis. It was found to be statistically significant in differentiation between CHM and PHM on one hand and between molar and nonmolar POC on the other hand, in agreement with previous studies. On the other hand, our results disagreed with the study of Abdou et al. on 59 cases of POC specimens and found that trophoblast hyperplasia had no statistically significant value in that differentiation. This marked circumferential villous trophoblast hyperplasia which was found in CHM cases may be related to the absence of p57Kip2 expression by the villous cytotrophoblasts and villous stromal cells in CHM cases which lack the maternal genome. This results in loss of cell cycle control and hence increased trophoblast proliferation.

It is known that villi in CHM have a high level of stromal karyorrhexic debris. In this study, villous stromal karyorrhectic debris wasn’t observed in any case diagnosed as PHM or nonmolar POC. Although the villous stromal karyorrhectic debris was found in 36% of CHM cases, it wasn’t statistically significant in differentiation of the three entities; CHM vs PHM or molar vs nonmolar pregnancies. This was in agreement with the finding that stromal karyorrhexis couldn’t differentiate between POC cases. This was in disagreement with the results of a study including 113 specimens of POC before 15th week gestation and found that the rate of stromal karyorrhexis was significantly higher in early CHM than in PHM (p < .001). They proved and confirmed that the frequent karyorrhexis in the villous stroma is a useful histopathologic parameter in the differential diagnosis of CHM from PHM and HA in early gestational age. However, the stromal karyorrhexis couldn’t be used as a significant feature in cases showing diffuse hydropic change due to an absence of cellular components in the stroma.

The current study showed that atypia (in the form of increased nucleocytoplasmic ratio, hyperchromasia, and pleomorphism) in extravillous trophoblast was mainly of marked degree in 74% of CHM, and of mild degree in 71% of PHM cases. Marked atypia was in favor of CHM diagnosis, while mild atypia was in favor of PHM and HA diagnosis. Marked degree of atypia was statistically significant in differentiation between CHM and PHM specimens, while mild degree of atypia wasn’t significant in differentiation between molar and nonmolar POC. It may be a useful histopathologic feature regarding the diagnosis and classification of HM. This was consistent with the observation that trophoblast atypia could differentiate between CHM and PHM where diffuse marked atypia was found in most CHM and focal mild atypia in most PHM. In our study, seven cases diagnosed histopathologically as PHM were reclassified as CHM after p57Kip2 immunohistochemistry (showed no or scattered nuclear staining in villous CT and villous stromal cells). Six of these cases exhibited morphological features similar to PHM on histopathological examination in the form of two populations of villi; small and large distended villi with irregular outline and multifocal villous trophoblast hyperplasia. One case showed two villous populations, circumferential villous trophoblast hyperplasia and trophoblastic inclusion in the core of their villi without stromal karyorrhexis. In these cases, we can consider p57Kip2 as a gold standard for the diagnosis and classification of HM cases. This was similar to previous observations.

In a retrospective study done by Landolsi et al. on 220 specimens of HA, negative p57Kip2 expression was observed in 8 cases with a histopathological diagnosis of PHM, and in one case with a diagnosis of HA. Landolsi et al. tried to explain this negative p57Kip2 expression either due to mis-diagnosis of CHM or lack of staining due to loss of antigenicity. They proved the mis-diagnosis of CHM by genotyping analysis of their discordant nine cases and found absence of their maternal allele. In our study, seven cases diagnosed histopathologically as CHM were reclassified as PHM after p57Kip2 immunohistochemistry which showed positive nuclear staining in villous CT and villous stromal cells. This was also found in the results of previous studies in which one case out of 132 CHM cases was found to show positive p57Kip2 immunostaining in their CT and villous stromal cells. This was explained by false-positive immunohistochemistry or a mis-diagnosis of PHM or HA. Another possibility is the presence of twin gestation, one of them normal and the other CHM. Furthermore, rare CHMs are biparental in origin and contain both maternal and paternal chromosomal components. They explained it as a mis-diagnosis, as no adequate

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http://jpatholtm.org/
DNA material was available for genetic study to confirm. In such rare cases, we can’t consider p57Kip2 as a gold standard for the diagnosis and the matter will require further studies including molecular techniques using the differences in DNA content.

Concerning the follow-up data, nine cases out of the 65 total cases (14%) developed PGTD. They were diagnosed based on persistence of their symptoms and their serum β-HCG level which didn’t come down to the basal level. Eight cases of these were diagnosed as CHM (18% of total CHM). Similar results were found in a study performed by Van Cromvoirt et al.21 to identify cases the developed PGTD after CHM; 89 cases (20%) in inconsistent with the percentage reported by Chen treated successfully by methotrexate chemotherapy. Also, it was found that 2.5%–7.5% of PHM can progress to PGTD. We can attribute this difference between the two studies to the very limited number of the PHM cases studied in the current study. We reviewed these cases histopathologically and didn’t find any specific morphological or immunohistochemical features associated with their progression to PGTD. In that regard, the current study was in agreement with the histopathological and immunohistochemical study done by Petts et al.25 on 150 cases of molar pregnancy. To achieve this distinction, further studies are required using a larger number of cases and implementing further immunohistochemical markers that may have a benefit.

In conclusion, histopathological diagnosis of molar pregnancy remains problematic especially in early gestational age. The diagnosis requires a constellation of specific histopathological criteria in order not to miss the diagnosis of CHM. p57Kip2 immunohistochemistry is of great value in the diagnosis of cases that have equivocal morphology by histopathological examination. However, there are no significant features to predict cases that subsequently develop persistent trophoblastic disease. Further immunohistochemical markers should be studied to accurately distinguish PHM from CHM and also to predict patients’ outcome.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

REFERENCES

23. van Cromvoirt SM, Thomas CM, Quinn MA, McNally OM, Bekkers RL. Identification of patients with persistent trophoblastic disease after complete hydatidiform mole by using a normal 24-hour urine hCG regression curve. Gynecol Oncol 2014; 133: 542-5.
Loss of Progesterone Receptor Expression Is an Early Tumorigenesis Event Associated with Tumor Progression and Shorter Survival in Pancreatic Neuroendocrine Tumor Patients

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Pancreatic neuroendocrine tumors (PanNETs) are rare, amounting to only 3% of pancreatic neoplasms and 9% of all gastrointestinal neuroendocrine tumors in Korea.\(^1\) Although PanNET patients have better survival outcomes than pancreatic ductal adenocarcinoma patients, PanNETs are still malignant neoplasms with a 10-year survival rate of only 40%—50% after surgical resection.\(^2\) Surgical resection is the main curative treatment option for PanNET patients, although other therapies with somatostatin analogs, cytotoxic chemotherapies, and molecular targeted therapies have recently been used in metastatic or unresectable PanNET patients.\(^3\) A better understanding of the molecular mechanisms of PanNETs is important to better predict clinical outcomes and identify patients who may benefit from therapies targeting PanNETs. A previous whole-exome sequencing study revealed the genomic landscape of PanNETs including a higher mutational frequency in MEN1, ATRX (alpha thalassemia/mental retardation syndrome X-linked), and DAXX (death-domain associated protein) and a lower mutational frequency in several genes involving mammalian target of rapamycin pathways, including PTEN, TSC2, and PIK3CA.\(^7\) In that study also, less than half of PanNETs had inactivating mutations in either the ATRX or DAXX genes, in mutually exclusive ways.\(^8\) Alterations in the ATRX or DAXX proteins were closely associated with the alternative lengthening of telomeres (ALT) mechanism in PanNETs,\(^9\) and the loss of ATRX or DAXX expression and ALT activation were associated with poorer survival in primary PanNET patients.\(^10\)

The hormonal expression status in PanNETs has clinical implications. Pancreatic neuroendocrine tumors (PanNETs) are the second most common pancreatic neoplasms and there is no well-elicited biomarker to stratify their detection and prognosis. Previous studies have reported that progesterone receptor (PR) expression status was associated with poorer survival in PanNET patients. Methods: To validate previous studies, PR protein expression was assessed in 21 neuroendocrine microadenomas and 277 PanNETs and compared with clinicopathologic factors including patient survival. Results: PR expression was gradually decreased from normal islets (49/49 cases, 100%) to neuroendocrine microadenoma (14/21, 66.6%) to PanNETs (60/277, 21.3%; \(p<.001\)). PanNETs with loss of PR expression were associated with increased tumor size \((p<.001)\), World Health Organization grade \((p=.001)\), \(pT\) classification \((p<.001)\), perineural invasion \((p=.028)\), lymph node metastasis \((p=.004)\), activation of alternative lengthening of telomeres \((p=.005)\), other peptide hormonal expression \((p<.001)\) and ATRX/DAXX expression \((p=.015)\). PanNET patients with loss of PR expression (5-year survival rate, 64.1%) had significantly poorer recurrence-free survival outcomes than those with intact PR expression (90%) by univariate \((p=.012)\) but not multivariate analyses. Similarly, PanNET patients with PR expression loss (5-year survival rate, 76%) had significantly poorer overall survival by univariate \((p=.015)\) but not multivariate analyses. Conclusions: Loss of PR expression was noted in neuroendocrine microadenomas and was observed in the majority of PanNETs. This was associated with increased grade, tumor size, and advanced \(pT\) and \(pN\) classification; and was correlated with decreased patient survival time by univariate but not multivariate analyses. Loss of PR expression can provide additional information on shorter disease-free survival in PanNET patients.

Key Words: Pancreas; Neuroendocrine tumors; Receptors, progesterone; Survival
plications. Peptide hormonal expression is reported to have better prognostic implications for PanNET patients. Increased expression of insulin, GLP1, and other peptide hormones are associated with better overall survival in PanNET patients, whereas gastrin expression has been reported to be associated with worse survival. Among the steroid hormones, only progesterone receptors (PRs), but not estrogen receptors or androgen receptors, are expressed in normal islets. PR expression in normal pancreatic islets suggests a possible role for progesterone in pancreatic islet function. A study in mice with intact gonads demonstrated that progesterone treatment stimulated β-cell proliferation in pancreatic islets. However, those proliferation effects were lost in gonadectomized mice. Those observations suggested that progesterone effects require intact gonadal function for normal β-cell proliferation. In our present study, we determined the clinical and prognostic significances of PR expression in surgically resected PanNET patients using tissue microarray immunolabeling.

MATERIALS AND METHODS

Case selection

After approval (2015-0387) from the Institutional Review Board of Asan Medical Center, the records of 277 surgically resected primary PanNETs and 21 sporadic neuroendocrine microadenomas resected between January 1995 and December 2015 were retrieved from the Department of Pathology at Asan Medical Center. Primary PanNETs were defined as well-differentiated, nonfunctional neuroendocrine neoplasms with diameters ≥0.5 cm, while neuroendocrine microadenomas were defined as well-differentiated, nonfunctional NETs with diameters <0.5 cm. All PanNET cases were classified using the 2010 World Health Organization (WHO) classification scheme with mitotic activity and the Ki-67 labeling index. The Ki-67 labeling index was measured by manually assessing the tumor’s hottest spot in high-power fields after printing the captured image, as previously described. A minimum of 500 tumor cells were included in the manual count. Poorly differentiated neuroendocrine carcinomas, such as small-cell carcinomas and large-cell carcinomas, were excluded. Pathological data, such as tumor size, extension, lymph node and distant metastases, and perineural and lymphovascular tumor invasion, were extracted from the pathology reports. The clinical data reviewed included patient age, gender, symptoms, and survival outcomes. The expression profiles of specific peptide hormones, including insulin, glucagon, gastrin, serotonin, somatostatin, and glucagon-like peptide 1; other proteins, such as ATRX and DAXX; and the ALT status were used as previously reported.

Tissue microarray construction

Tissue microarrays were constructed from archived, formalin-fixed, paraffin-embedded tissue blocks with a manual tissue microarrayer (Uni TMA Co. Ltd., Seoul, Korea), as previously described. Briefly, three cores from the tumors and one core from the normal pancreatic parenchyma with a diameter of 2 mm were punched from donor blocks and placed in recipient blocks.

Immunohistochemical staining

Immunohistochemical labeling was performed at the immunohistochemical laboratory of the Department of Pathology, Asan Medical Center. In brief, 4-mm-thick tissue sections were deparaffinized and hydrated in xylene and serially diluted in ethanol. Endogenous peroxidase was blocked by incubation in 3% H2O2 for 10 minutes, and then heat-induced antigen retrieval was performed. Primary antibodies were used with a Benchmark autostainer (Ventana Medical Systems, Tucson, AZ, USA) in accordance with the manufacturer’s protocol. Sections were incubated at room temperature for 32 minutes in primary antibodies for PR (1:200, NCL-L-PGR-312, Novocastra, Newcastle upon Tyne, UK), synaptophysin (1:200, DiNona, Seoul, Korea), chromogranin (1:200, DAK-A3, DakoCytomation, Glostrup, Denmark), and Ki-67 (1:100, 7B11, Zymed, San Francisco, CA, USA). The sections were then labeled with an automated immunostaining system and processed with an iView DAB detection kit (Benchmark XT, Ventana Medical Systems). Immunostained sections were lightly counterstained with hematoxylin, dehydrated in ethanol, and cleared in xylene. Immunoreactivity was interpreted by light microscopic examination and independently evaluated by two pathologists, coauthors of this study (S.J.K. and S.M.H.), who were blind to the clinicopathologic information.

Evaluation of PR labeling

Immunohistochemical labeling of the PR protein was scored using a previously described histological scoring system, which takes into account the size of the stained area and the intensity of the labeling. To be included in the analyses, a tumor had to have sufficient numbers of PR-labeled cells to permit quantification (>100 PR-positive tumor cells). The labeled area was scored from 0 to 4 for having <5%, 5%–25%, 26%–50%, 51%–75%, or >75% PR-positive cells, respectively. The intensity scale ranged from 0 to 2 as follows: 0, no labeling of tumor cells; 1, weak labeling; and 2, intense labeling, as previously described.
The total histological score (H-score) was calculated by multiplying the area score by the intensity score. The resulting H-score ranged from 0 to 8. We considered cases with an H-score below 2 to have loss of PR expression and cases with an H-score greater than or equal to 2 to have intact PR expression. PR labeling in normal islets was used as an internal positive control. Representative images of PR expression in normal islet and in PanNETs are depicted in Fig. 1. Immunohistochemical staining was evaluated by two independent pathologists (S.J.K. and S.M.H.).

Statistical analyses
SPSS software ver. 20.0 (IBM Corp., Chicago, IL, USA) was used for statistical analyses. The overall and recurrence-free survival times were calculated from the date of diagnosis of PanNET to that of death from any cause and from the date of diagnosis of PanNET to that of recurrence, respectively. Both overall and recurrence-free survival rates were calculated using the Kaplan-Meier method, and the association between overall survival rate and clinicopathologic factors was compared using the log-rank
test. The correlations between PR expression and other prognostic factors were analyzed using the chi-square and Fisher exact tests. Possible prognostic factors associated with survival probability were calculated using the Cox’s proportional hazard regression model; p < .05 was considered statistically significant.

RESULTS

Patient characteristics

Patient characteristics are summarized in Table 1. In the total cohort, 134 patients (49%) were male and 143 (51%) were female. The mean age of the patients was 52.3 ± 12.7 years. According to the WHO classification, there were 85 G1, 95 G2, and 8 G3, respectively. The mean tumor size of the PanNETs and neuroendocrine microadenomas was 3.0 ± 2.2 cm and 0.3 ± 0.1 cm, respectively. In all, 126 cases were classified as pT1, while others had higher pT classifications (95 pT2, 52 pT3, and 4 pT4); 79 cases (28.5%) had lymphovascular invasion and 42 cases (15.2%) had perineural invasion. Metastasis to regional lymph nodes occurred in 39 cases (14.1%) and metastasis to distant organs at the surgical resection of PanNET was observed in 10 cases (3.6%). The median follow-up period was 38 ± 35 months (range, 1 to 188 months).

PR expression

All of the normal pancreatic islets in the patient samples expressed the PR protein in various proportions. PR protein expression was observed in 63.5 ± 9.8% of the endocrine cells in the islets. A representative image of PR expression in an islet is depicted in Fig. 1. This expression gradually decreased from normal islets (49/49, 100%) to neuroendocrine microadenoma (14/21, 66.6%) to PanNETs (60/277, 21.3%). The H-scores for PR in normal islets, neuroendocrine microadenomas, and PanNETs were 5.1 ± 2.3, 3.0 ± 3.3, and 1.3 ± 2.5, respectively (p < .001) (Figs. 1, 2). The mean H-score of the PR expression loss group was 0.1 ± 0.3, while that of the PR expression intact group was 5.8 ± 2.3. The majority of PanNETs showed a loss of PR expression (218/277, 78.7%) (Fig. 1).

Correlations between PR expression and clinicopathologic factors

The associations between PR expression and clinicopathologic factors are summarized in Table 1. Loss of PR expression was more commonly observed in PanNETs with larger tumors (p < .001), a higher WHO grade (p = .001), higher Ki-67 labeling index (p = .004), higher pT classification (p < .001), frequent peri-

Table 1. Clinicopathologic factors associated with PR expression in PanNETs

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>PR loss</th>
<th>Intact PR</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yr)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>≤ 60</td>
<td>149 (78.0)</td>
<td>42 (22)</td>
<td>.412</td>
</tr>
<tr>
<td>&gt; 60</td>
<td>69 (80.2)</td>
<td>17 (19.8)</td>
<td></td>
</tr>
<tr>
<td>Sex</td>
<td></td>
<td></td>
<td>.380</td>
</tr>
<tr>
<td>Male</td>
<td>107 (79.9)</td>
<td>27 (20.1)</td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>111 (77.6)</td>
<td>32 (22.4)</td>
<td></td>
</tr>
<tr>
<td>Tumor size (cm)</td>
<td></td>
<td></td>
<td>&lt; .001</td>
</tr>
<tr>
<td>≤ 3</td>
<td>126 (71.2)</td>
<td>51 (28.8)</td>
<td></td>
</tr>
<tr>
<td>&gt; 3</td>
<td>92 (86.4)</td>
<td>8 (13.6)</td>
<td></td>
</tr>
<tr>
<td>WHO grade</td>
<td></td>
<td></td>
<td>.001</td>
</tr>
<tr>
<td>Grade 1</td>
<td>65 (76.5)</td>
<td>20 (23.5)</td>
<td></td>
</tr>
<tr>
<td>Grade 2</td>
<td>83 (87.4)</td>
<td>12 (12.6)</td>
<td></td>
</tr>
<tr>
<td>Grade 3</td>
<td>8 (100.0)</td>
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</tr>
<tr>
<td>pT classification</td>
<td></td>
<td></td>
<td>&lt; .001</td>
</tr>
<tr>
<td>pT1</td>
<td>70 (82.5)</td>
<td>42 (17.5)</td>
<td></td>
</tr>
<tr>
<td>pT2–T4</td>
<td>148 (89.7)</td>
<td>17 (10.3)</td>
<td></td>
</tr>
<tr>
<td>Lymphovascular invasion</td>
<td></td>
<td></td>
<td>.077</td>
</tr>
<tr>
<td>Absent</td>
<td>151 (76.3)</td>
<td>47 (23.7)</td>
<td></td>
</tr>
<tr>
<td>Present</td>
<td>67 (84.8)</td>
<td>12 (15.2)</td>
<td></td>
</tr>
<tr>
<td>Perineural invasion</td>
<td></td>
<td></td>
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<tr>
<td>Absent</td>
<td>180 (76.6)</td>
<td>55 (23.4)</td>
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<tr>
<td>Present</td>
<td>38 (90.5)</td>
<td>4 (9.5)</td>
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<tr>
<td>Lymph node metastasis</td>
<td></td>
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<tr>
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<td>181 (76.1)</td>
<td>57 (23.9)</td>
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<tr>
<td>Present</td>
<td>37 (94.9)</td>
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<td>Distant metastasis</td>
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<td>57 (21.3)</td>
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<td>Present</td>
<td>8 (80.0)</td>
<td>2 (20.0)</td>
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<td>Ki-67 labeling index (%)</td>
<td></td>
<td></td>
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<tr>
<td>≤ 3</td>
<td>137 (73.3)</td>
<td>50 (26.7)</td>
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<tr>
<td>&gt; 3 and &lt; 20</td>
<td>57 (89.1)</td>
<td>7 (10.9)</td>
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<td>≥ 20</td>
<td>2 (100.0)</td>
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<tr>
<td>Hormone expression</td>
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<td></td>
<td>&lt; .001</td>
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<tr>
<td>Absent</td>
<td>91 (94.8)</td>
<td>5 (5.2)</td>
<td></td>
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<tr>
<td>Present</td>
<td>49 (68.1)</td>
<td>23 (31.9)</td>
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<td>ALT expression</td>
<td></td>
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<td>.005</td>
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<tr>
<td>Absent</td>
<td>166 (75.7)</td>
<td>54 (24.5)</td>
<td></td>
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<tr>
<td>Present</td>
<td>52 (91.2)</td>
<td>5 (8.8)</td>
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<tr>
<td>ATRX/DAXX expression</td>
<td></td>
<td></td>
<td>.015</td>
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<tr>
<td>Absent</td>
<td>45 (90.0)</td>
<td>5 (10.0)</td>
<td></td>
</tr>
<tr>
<td>Present</td>
<td>166 (75.5)</td>
<td>54 (24.5)</td>
<td></td>
</tr>
</tbody>
</table>

Values are presented as number (%).

PR, progesterone receptor; PanNET, pancreatic neuroendocrine tumor; WHO, World Health Organization; ALT, alternative lengthening of telomeres.
Kim SJ, et al. Higher WHO grade (p < .001), higher pT classification (p = .015), and frequent lymphovascular (p = .002), perineural (p < .001), and regional lymph node metastasis (p < .001).

Multivariate analyses of clinicopathologic factors

The Cox proportional hazard model was employed with other significant clinicopathologic factors to determine the prognostic significance of PR expression as well as other clinicopathologic factors in PanNET patients. Only WHO grade (p = .001) and lymphovascular invasion (p = .020) were independently prognostic, but loss of PR expression was not a prognostic factor in our model (p = .117) (Table 2).
DISCUSSION

PR immunohistochemistry has been used to identify ovarian-type stroma for the diagnosis of mucinous cystic neoplasms of the pancreas in the field of pancreas pathology. A recent study demonstrated that loss of or decreases in PR expression in ovarian type stroma of invasive carcinoma with mucinous cystic neoplasm and mucinous cystic neoplasm with high-grade dysplasia were associated with decreased volumes of ovarian-type stroma in mucinous cystic neoplasms. These results suggest the possible utility of PR immunolabeling as a surrogate marker for invasion in the diagnosis of mucinous cystic neoplasms.

In addition to the use of PR expression for diagnosing mucinous cystic neoplasms, this expression has also been reported in all normal islets of Langerhans, and previous studies have detected PR in 40%–75% of islets. In our current study, PR expression was noted in 64% of normal islets, a similar proportion to that found in previous studies.

Neuroendocrine microadenomas are precursors and initiating lesions for PanNETs. Only a few previous studies have conducted biomarker evaluations for neuroendocrine microadenomas. Decreased menin expression and increased cytokeratin 19 expression have been reported in neuroendocrine microadenomas. However, controversy exists over the loss of ATRX or DAXX expression in these tumors. Confirming previous findings, we observed PR expression in 67% of neuroendocrine microadenomas, and our observations suggest that PR expression loss can also be included as an early event in neuroendocrine tumorigenesis.

PR expression was present in only 21% of the PanNETs examined in our current study, and the majority of PanNETs showed loss of PR expression. Previous studies have found wide ranges of PR expression, from 46% to 76%. Plausible explanations for the lower PR expression in our study include different ethnic backgrounds (Western vs Korean), antibody clones, and the cut-off point used for positive expression. While previous studies examined European or American populations, the present study was performed on a Korean population. This difference in ethnic background could have affected PR expression. In addition, previous studies considered PR expression loss when the proportion of nuclear PR labeling was < 1% or < 5%, whereas we used the H-score, multiplying the intensity and proportion of PR expression. Different cutoff points for the evaluation of PR expression could explain the lower levels detected in our study.

Loss of PR expression was associated with larger tumor size, higher WHO grade, higher pT classification, and frequent lymphovascular and perineural invasion, and regional lymph node metastasis. Our current observations are thus concordant with the results of previous studies. Arnason et al. previously examined 40 PanNET cases and observed that PanNETs with strong PR expression were associated with fewer nodal or distant metastases of PanNETs. Viale et al. studied 96 PanNETs and reported that less

Table 2. Univariate and multivariate analyses of PR expression in PanNETs

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Univariate analyses</th>
<th>Multivariate analyses</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>5-Year survival rate (%)</td>
<td>Hazard ratio</td>
<td>95% confidence interval</td>
</tr>
<tr>
<td>PR expression</td>
<td>Loss</td>
<td>64.1</td>
<td>.012</td>
</tr>
<tr>
<td></td>
<td>Intact</td>
<td>90</td>
<td></td>
</tr>
<tr>
<td>Age (yr)</td>
<td>≤ 60</td>
<td>70.3</td>
<td>.015</td>
</tr>
<tr>
<td></td>
<td>&gt; 60</td>
<td>62.1</td>
<td></td>
</tr>
<tr>
<td>Sex</td>
<td>Male</td>
<td>59.1</td>
<td>.609</td>
</tr>
<tr>
<td></td>
<td>Female</td>
<td>56.9</td>
<td></td>
</tr>
<tr>
<td>Tumor size (cm)</td>
<td>≤ 3</td>
<td>74.6</td>
<td>.008</td>
</tr>
<tr>
<td></td>
<td>&gt; 3</td>
<td>55.7</td>
<td></td>
</tr>
<tr>
<td>WHO grade</td>
<td>Grade 1</td>
<td>76.9</td>
<td>&lt;.001</td>
</tr>
<tr>
<td></td>
<td>Grade 2</td>
<td>65.8</td>
<td>1.24</td>
</tr>
<tr>
<td></td>
<td>Grade 3</td>
<td>0</td>
<td>10.74</td>
</tr>
<tr>
<td>pT classification</td>
<td>pT1</td>
<td>76.7</td>
<td>.015</td>
</tr>
<tr>
<td></td>
<td>pT2–T4</td>
<td>61.8</td>
<td></td>
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<tr>
<td>Lymphovascular invasion</td>
<td>Absent</td>
<td>71.5</td>
<td>.002</td>
</tr>
<tr>
<td></td>
<td>Present</td>
<td>54</td>
<td></td>
</tr>
<tr>
<td>Perineural invasion</td>
<td>Absent</td>
<td>71.5</td>
<td>&lt;.001</td>
</tr>
<tr>
<td></td>
<td>Present</td>
<td>41</td>
<td></td>
</tr>
<tr>
<td>Lymph node metastasis</td>
<td>Absent</td>
<td>73</td>
<td>&lt;.001</td>
</tr>
<tr>
<td></td>
<td>Present</td>
<td>27.8</td>
<td></td>
</tr>
</tbody>
</table>

PR, progesterone receptor; PanNET, pancreatic neuroendocrine tumor; WHO, World Health Organization.

https://doi.org/10.4132/jptm.2017.03.19 http://jpatholtm.org/
PR immunoreactivity was more commonly associated with malignant behaviors, including metastasis, invasion of surrounding tissues, or larger vessel involvement. Estrella et al. reviewed 160 PanNET cases and found that loss of PR expression was associated with larger tumor size and advanced American Joint Committee on Cancer tumor staging but was not correlated with age, gender, or WHO grade.

In our present study series, we observed that PanNET patients with PR expression loss had significantly poorer overall and disease-free survival outcomes by univariate but not multivariate analyses. Thus, PR status can provide additional survival information for PanNET patients but cannot be used as a prognostic indicator. Previous studies have also evaluated PanNETs with PR expression and patient survival. Arnason et al. reported that pancreas and small intestinal NET patients with intact PR expression had significantly better disease-free survival (median, 155 months) than those with decreased PR expression (median, 38 months). However, they further found that this was only marginally significant when they restricted their examination to only PanNET patients. Estrella et al. observed no significant differences in overall survival based on PR expression status only. However, when they compared the survival of PanNET patients after combining PTEN and PR expression status, dual PR- and PTEN-negative PanNET patients showed shorter metastasis-free survival than either single PR- or PTEN-positive patients or dual PR- and PTEN-positive patients. The biological roles of PR in normal islets and PanNETs have not been completely elucidated. One previous study demonstrated that the administration of progesterone to PR knock-out mice with intact gonads induced β-cell proliferation, which suggests anti-proliferation activity for PR. PRs exist as two protein isoforms, PRA and PRB. Recently, Yazdani et al. demonstrated that PanNET tumorigenesis occurred via activation of PRB after its binding to progesterone, which was induced by the activation of transcription factors FOS and Jun and followed by overexpression of CCND1. They also demonstrated that PRA in the progesterone signaling pathway inhibited PanNET tumorigenesis by suppressing the PRB promoter.

In summary, we performed an immunohistochemical study of PR in 21 surgically resected neuroendocrine microadenomas and 277 PanNETs. Our key findings were loss of PR expression in neuroendocrine microadenomas and in the majority of PanNETs and associated with increased WHO grade, tumor size, and advanced pT and pN classification. The loss of PR expression also correlated with decreased patient survival time according to univariate, but not multivariate, analysis. In conclusion, the loss of PR expression can provide additional information on shorter disease-free survival outcomes in PanNET patients.

**Conflicts of Interest**

No potential conflict of interest relevant to this article was reported.

**Acknowledgments**

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**REFERENCES**

27. de Wilde RF, Heaphy CM, Maitra A, et al. Loss of ATRX or DAXX expression and concomitant acquisition of the alternative lengthening of telomeres phenotype are late events in a small subset of MEN1 syndrome pancreatic neuroendocrine tumors. Mod Pathol 2012; 25: 1033-9.
HER2 Status and Its Heterogeneity in Gastric Carcinoma of Vietnamese Patient

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Quoc Dat Ngo
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Sao Trung Nguyen · Anh Tu Thai1
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Background: Human epidermal growth factor receptor 2 (HER2) is related to the pathogenesis and poor outcome of numerous types of carcinomas, including gastric carcinoma. Gastric cancer patients with HER2 positivity have become potential candidates for targeted therapy with trastuzumab. Methods: We investigated 208 gastric cancer specimens using immunohistochemistry (IHC), fluorescence in situ hybridization and dual in situ hybridization (ISH). We also investigated the concordance between IHC and ISH. The correlation between HER2 status and various clinicopathological findings was also investigated. Results: In total, 15.9% (33/208) and 24.5% (51/208) of gastric cancers showed HER2 gene amplification and protein overexpression, respectively. A high level of concordance between ISH and IHC analyses (91.3%, κ=0.76) was found. A significant correlation between HER2 status and intestinal-type (p<.05) and differentiated carcinomas (p<.05) was also noted. The HER2 heterogeneity was high in gastric cancers; we found 68.8% phenotypic heterogeneity and 57.6% genotypic heterogeneity. Heterogeneity in HER2 protein expression and gene amplification showed a close association with diffuse histologic type and IHC 2+. Conclusions: HER2 protein overexpression and gene amplification were detected in 24.5% and 15.9% of gastric cancer specimens, respectively. Intestinal-type showed a higher level of HER2 protein overexpression and gene amplification than diffuse type. HER2 status also showed a significant relationship with well- and moderately-differentiated carcinomas. The ratio of phenotypic and genotypic heterogeneity of HER2 was high in gastric carcinomas and was associated with HER2 IHC 2+ and diffuse histologic type.

Key Words: Protein HER2; Stomach neoplasms; Protein overexpression; Immunohistochemistry; Gene amplification; Fluorescence in situ hybridization; Dual-color silver in situ hybridization

According to global cancer statistics from 2012 (GLOBOCAN 2012), gastric cancer remains the fourth most frequent cause of cancer-related mortality in developing countries. In Vietnam, the incidence of gastric cancer has been increasing in recent years and poor outcome was due to delayed diagnosis and advanced stage.

Conventional chemotherapy has not been very beneficial for gastric cancer. Targeted therapeutics, especially human epidermal growth factor receptor 2 (HER2) inhibitors, have recently shown more advantages in treating late-stage gastric cancer. The Trastuzumab for Gastric Cancer trial has proven the efficacy of humanized monoclonal antibody against HER2 protein, so-called trastuzumab, in the treatment of advanced gastric cancer or gastroesophageal junction cancer with HER2 positivity. HER2 is a proto-oncogene encoded by ERBB2 on chromosome 17, also a member of the HER family. The intracellular domain of HER2, a membrane-bound receptor tyrosine kinase, is responsible for extracellular signal transmission to initiate cellular signaling pathways such as mitogen-activated protein kinase, phosphoinositide 3-kinase, phospholipase C, and protein kinase C, leading to cell proliferation, cell survival, angiogenesis, migration, and invasion. HER2-positive status (protein overexpression and/or gene amplification) has been identified in several carcinomas, especially in breast cancer, where it has a frequency of 20%–25%, suggesting its prognostic and predictive value in this carcinoma. On the other hand, in gastric cancer, the reported level of HER2 overexpression and amplification varies widely from 7%–27%, and the prognostic value of HER2 status is not commonly accepted.

This study aimed to detect HER2 status including protein expression and gene amplification, as well as investigate HER2 heterogeneity and the concordance between two methods (immunohistochemistry [IHC] and in situ hybridization). We also
investigated the relationship between HER2 status and clinicopathological findings.

MATERIALS AND METHODS

Patients and tissue specimens
For this study, we retrospectively enrolled 208 gastric cancer patients who underwent curative gastrectomy at the University Medical Center, Ho Chi Minh City, Vietnam, from January 2012 to June 2015. No patients had received preoperative radiation or chemotherapy. All surgical specimens were promptly fixed in 10% buffered formalin, no longer than 30 minutes after resection, and for a fixing time of 8–48 hours. Clinicopathological features were recorded including age, sex, histologic type by Lauren, tumor differentiation, depth of invasion, lymph node status, and lymphovascular invasion.

IHC staining
We performed IHC staining for HER2 on the automated slide stainer (BenchMark XT), using monoclonal antibody 4B5 (Ventana Medical System, Tucson, AZ, USA) following the manufacturer’s procedures. We incubated 5-µm slices after antigen retrieval (10 mM citrate buffer at 98°C for 40 minutes) with 4B5 for 30 minutes. Subsequently 3,3’-diaminobenzidine was added for visualization. The sections were stained with hematoxylin for counterstaining and then cover-slipped. Negative controls (omission of primary antibody) and positive controls (gastric cancer tissue with known IHC HER2 3+) were included in each staining run.

Evaluation of IHC results
Slides were scored by a pathologist following the scoring system of Hofmann et al.6 The generally accepted cut-off definition for HER2 membranous positivity is 10% of the tumor cells. A score of 0 was defined as no reactivity or membrane staining in <10% of the tumor cells. Faint membrane staining was defined as a score of 1+. Weak-to-moderate complete membrane staining was defined as a score of 2+, and moderate-to-strong complete membrane staining was defined as a score of 3+. Scores of 2+ and 3+ were defined as HER2 protein positive/over-expression. Phenotypic heterogeneity was defined as the proportion of tumor cells showing membranous staining in more than 10%, but less than 50% of tumor cells.

In situ hybridization
We performed in-situ hybridization (ISH) for 208 cases of gastric cancer using dual-color silver in situ hybridization (SISH) (28 cases randomly selected) and fluorescence in situ hybridization (FISH) (the remaining 180 cases).

Fluorescence in situ hybridization
HER2 gene amplification was detected by FISH using a Pathvision HER2 DNA probe kit (Vysis Inc., Downers Grove, IL, USA) according to the manufacturer’s instructions. We heated hybridization buffer, DNA probe, and purified water to 75°C for 5 minutes. Slides were dipped in a denaturing bath (70% formamide 2× saline-sodium citrate [SSC]) for 5 minutes at 75°C, then dehydrated in increasing ethanol concentrations, and then covered with the probe mixture. We incubated all slides for 30 minutes at 42°C, then washed with 0.4× SSC/0.3% NP-40 for 2 minutes, air-dried in darkness, counterstained with 4’,6-diamidino-2-phenylindole (DAPI), and lastly covered with a cover-slip. The signals were visualized under fluorescence microscope (BX51, Olympus, Tokyo, Japan) and the images were captured and saved with Spectral Imaging software (Applied Spectral Imaging Inc., Carlsbad, CA, USA). HER2 gene expression is represented by discrete red signals or signal clusters and Chr17 as green signals.

Dual-color SISH
We performed automated SISH on the Ventana Benchmark XT platform (Ventana Medical Systems Inc.) following the manufacturer’s protocols and using the INFORM HER2 DNA and Chromosome 17 probes. HER2 is identified by a dinitrophenyl (DPN) labeled probe and visualized utilizing ultraView SISH DNP Detection Kit. The Chr17 centromere is labeled with a digoxigenin (DIG) probe and detected using ultraView Red ISH DIG Detection Kit (Ventana Medical Systems Inc.). The signals were visualized under light microscopy (BX50, Olympus) in which HER2 expression appeared as discrete black signals (SISH) and Chr17 as red signals. The mean HER2/CEP17 ratio was calculated by counting signals in 20 tumor cells under high-power field (×600).

Evaluation of FISH and dual SISH results
HER2 gene status was based on the ratio of HER2 gene copies and Chr17 copies in nuclei of tumor cells. HER2 gene status was defined as non-amplified if the HER2/Chr17 ratio was under 2.0 or amplified if the HER2/Chr17 ratio was 2.0 or more. If the HER2/CEP17 ratio was 1.8 or more but less than 2.0, 20 more tumor cells were investigated to determine the HER2 gene status. HER2 amplification was further sub-classified into low-level amplification (HER2/CEP17 ratio 2–5) and high-level amplification.
(HER2/CEP17 ratio ≥ 5).

Polysomy of chromosome 17 was set as more than 3 CEP17 signals per nuclei. Genotypic heterogeneity was defined by the existence of more than 5% but less than 50% of tumor cells with a HER2/CEP17 ratio higher than 2.0.

Statistical analysis

We used SPSS ver. 15.0 statistical software program (SPSS Inc., Chicago, IL, USA) for statistical analyses. The chi-square tests (Kappa chi-square test or chi-square test) were performed to analyze correlations between HER2 status and clinicopathological features. p-values below .05 indicate statistically significant results.

RESULTS

Clinicopathological characteristics of patients

Patient age ranged widely from 30 to 81 years (median, 58.2 years). There was a higher number of male patients (n = 139) than female patients (n = 69), with a male/female ratio of 2:1. Clinicopathological features are noted in Table 1.

Overexpression of HER2

HER2 protein expression in 208 gastric tissue samples was investigated by IHC staining (Fig. 1). Of the 208 gastric carcinoma tissue samples, 148 (71.2%) were scored as 0, 9 (4.3%) scored as 1+, 23 (11.1%) scored as 2+, and 28 (13.4%) scored as 3+. In total, 24.5% were positive (51/208).

HER2 amplification

The amplification rate of the HER2 gene was 15.9% (33/208) in all gastric carcinoma tissue samples, with 29 cases detected by FISH and four cases detected by dual SISH (Fig. 2). Of amplified cases, 27 cases (81.8%) showed low-level amplification (ratio HER2/CEP17, 2 to < 5) and only six cases (18.2%) showed a high ratio HER2/CEP17 (> 5). Polysomy of chromosome 17 was not found in any of the tissue samples.

The correlation between IHC and ISH in the surgical specimens is shown in Table 2. A concordance between IHC and ISH results was seen in 190/208 (157 IHC 0/1+ not amplified, 6 IHC 2+ amplified, and 27 IHC 3+ amplified) with a rate of 91.3% (κ = 0.76). Six out of 23 IHC 2+ cases (26.1%) showed amplification and one out of 28 IHC 3+ cases (3.6%), which showed signet ring cell carcinoma, was not-amplified.

Correlation between clinicopathological findings and HER2 status

The clinicopathological differences were shown in gastric carcinoma with or without HER2 protein expression or HER2 gene amplification. The HER2 protein expression and HER2 gene amplification rates were 43.6% (34/78) and 29.5% (23/78) in intestinal type gastric carcinomas (p < .05). HER2 status was also related to good differentiation (p < .05). However, no significant correlation was found between HER2 status and other clinicopathological features including sex and age of the patients, depth of invasion, lymph node status, and lymphovascular invasion (Table 3).

Intratumor heterogeneity of HER2

HER2 showed phenotypic heterogeneity (IHC expression) and genotypic heterogeneity (gene amplification) (Fig. 3). Phenotypic heterogeneity was observed in 35 of 51 IHC cases (68.8%), in which 12 cases of 28 IHC 3+ cases (42.9%) and all 23 IHC 2+ cases (100%) demonstrated heterogeneity. Phenotypic heterogeneity of HER2 expression was higher in the diffuse type than in the intestinal type (75% vs 67.6%). Genotypic heterogeneity was observed in 19 of 33 ISH cases (57.6%). Genotypic heterogeneity was higher in diffuse-type than in intestinal-type cancers (66.7% vs 52.2%).

Table 1. Clinical and pathologic features of 208 cases of gastric cancer

<table>
<thead>
<tr>
<th>Parameter</th>
<th>No. (%)</th>
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<td>Sex</td>
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<td>139 (66.8)</td>
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<tr>
<td>Female</td>
<td>69 (33.2)</td>
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<tr>
<td>Age</td>
<td></td>
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<tr>
<td>Range</td>
<td>30–82</td>
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<tr>
<td>Mean ± standard deviation</td>
<td>58.2 ± 11.6</td>
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<td>Histologic type by Lauren</td>
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<tr>
<td>Intestinal type</td>
<td>78 (37.5)</td>
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<tr>
<td>Diffuse type</td>
<td>109 (52.4)</td>
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<td>Mixed</td>
<td>21 (10.1)</td>
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<td>77 (37.0)</td>
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<td>12 (5.8)</td>
</tr>
<tr>
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<td>36 (17.3)</td>
</tr>
<tr>
<td>T3</td>
<td>10 (4.8)</td>
</tr>
<tr>
<td>T4</td>
<td>150 (72.1)</td>
</tr>
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<td>Lymph node status (pN stage)</td>
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</tr>
<tr>
<td>N0</td>
<td>87 (41.8)</td>
</tr>
<tr>
<td>N1, 2, 3</td>
<td>121 (58.2)</td>
</tr>
<tr>
<td>Lymphovascular invasion</td>
<td>57 (27.4)</td>
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</table>
HER2 Status in Gastric Carcinoma

Fig. 1. The human epidermal growth factor receptor 2 (HER2) protein expression detected by immunohistochemistry (IHC). HER2 IHC scores of 1+ (A), 2+ (B), and 3+ (C).

Fig. 2. The human epidermal growth factor receptor 2 (HER2) gene status detected by fluorescence in situ hybridization (FISH) and dual-color silver in situ hybridization (SISH). Representative photomicrographs of FISH (A, C) and dual-color SISH (B, D). Non-amplified cases detected by FISH (A) and dual-color SISH (B). Amplified cases detected by FISH (C) and dual-color SISH (D).

Table 2. Correlation between HER2 protein expression and HER2 gene amplification

<table>
<thead>
<tr>
<th>HER2</th>
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<tr>
<td></td>
<td>3+ (n=28)</td>
<td>2+ (n=23)</td>
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<tr>
<td>Amplified</td>
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<tr>
<td>Not-amplified</td>
<td>1</td>
<td>17</td>
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HER2, human epidermal growth factor receptor 2; ISH, in situ hybridization; FISH, fluorescence in situ hybridization.
DISCUSSION

We investigated HER2 protein overexpression and gene amplification in 208 surgical tissue samples of gastric carcinoma. HER2 protein positivity and HER2 gene amplification were observed in 24.5% and 15.9% of cases, respectively. While HER2 protein positivity in gastric carcinoma was reported from 11 to 24%,6,8,11-13 HER2 gene amplification varied from 7% to 27%.7-10 This variation was attributed to an inconsistent HER2 IHC scoring system, different methodologies including antibody selection and/or random selection of patients. Moreover, the interpretation of IHC or ISH can be influenced by subjective factors such as the prompt fixing time after removal, the volume of formalin, and the duration of fixation. In this study, all gastric specimens were fixed no later than 30 minutes after removal in 10% buffered formalin for 8–48 hours.

In our study, we used the HER2 scoring system proposed by Hofmann et al.,6 showing a high level of concordance (91.3%) between IHC and ISH. A number of studies investigated the relationship between IHC and FISH in gastric cancer specimens.

![Fig. 3. Intratumoral heterogeneity of human epidermal growth factor receptor 2 (HER2). Heterogeneity of HER2 protein expression (A) and heterogeneity of HER2 gene amplification (B).](image-url)
and also reported a high level of concordance that varied from 86.9 to 93.5%. Our study also showed high concordance between IHC and ISH. We did not compare the concordance between dual SISH and FISH in this study. However, some studies have shown a high level of concordance between dual SISH and FISH, which ranged from 94.5% to 98.3%.3,9

In this study, we recognized that one of 28 HER2 IHC 3+ cases showed non-amplification, and it was signet ring cell carcinoma. HER2 status may be found in this rare histologic type of gastric cancer with a very low rate of positivity. The HER2 protein expression in signet ring cell carcinoma may be misinterpreted due to strong nuclear/cytoplasmic staining, which was observed in the IHC staining method. Therefore, ISH confirmation should be considered in signet ring cell carcinoma.

The higher positive rates of HER2 in intestinal-type compared to other types of gastric carcinoma have been widely reported and our results were similar.8,11,13-16 It was also reported that HER2 overexpression is related to well-and-moderately differentiated gastric cancer but not with tumor stage or the age and sex of gastric cancer patients.13,17,18 On the other hand, the correlation of positive HER2 status with other clinicopathological factors, including age, sex, and TNM stage, remains controversial. It has been reported that a higher frequency of HER2 positivity is significantly correlated with older age, male gender, upper location of tumor, lymphatic or vascular invasion, higher T stage, lymph node metastasis, distant metastasis, and higher TNM stage.8,11,13,19-21 In our study, we demonstrated that there was no significant relationship between HER2 status and clinicopathological variables such as sex and age of the patients, depth of invasion, lymph node status and lymphovascular invasion.

Genotypic heterogeneity was defined by College of American Pathologists (CAP) in 2009 for breast cancers, in which the tumor showed at least 5% but fewer than 50% of nuclei with a HER2/CEP17 ratio ≥ 2.0.22 The 2013 American Society of Clinical Oncology/CAP HER2 guideline update2 recommends using an IHC HER2 test to define areas of potential amplification. In gastric cancer, there are no guidelines for tumor heterogeneity assessment. Yang et al.16 recognized 79.3% of cases with heterogeneous HER2 protein expression and 44.0% of cases with HER2 genetic heterogeneity. Our results also showed that frequency of phenotypic heterogeneity was higher than that of genotypic heterogeneity (68.8% vs 57.6%).

Kim et al.11 found that heterogeneity was more prevalent in IHC 2+ cases than IHC 3+ cases (90.9% vs 40.9%). Nishida et al.23 also reported that phenotypic heterogeneity was recognized in 63.5% of IHC 2+ cases, and in only 28.3% in IHC 3+ cases. Our results are consistent with results from these studies, in which phenotypic heterogeneity and genotypic heterogeneity showed a higher rate in IHC 2+ cases than IHC 3+ cases.

Lee et al.24 recognized that diffuse or mixed histological subtypes have a higher rate of heterogeneous HER2 expression. In our study, we also found that the diffuse type showed a higher rate of heterogeneity for HER2 expression or amplification than did the intestinal type. These results may be important for pathologists to consider when predicting HER2 heterogeneity in IHC and ISH results during routine histological examination.

In conclusion, our study demonstrates that HER2 protein expression and gene amplification were recognized in 24.5% and 15.9% of gastric cancer patients, respectively. Intestinal-type disease showed a higher rate of HER2 protein expression and gene amplification than diffuse type. HER2 status also showed a significant relationship with well and moderately differentiated carcinomas. The ratio of phenotypic and genotypic heterogeneity of HER2 was high in gastric carcinoma and related to HER2 IHC 2+ and diffuse histologic type.

Conflicts of Interest
No potential conflict of interest relevant to this article was reported.

Acknowledgments
We would like to give special thanks to Hoffmann La Roche Company and Roche Diagnostics Vietnam for HER2 IHC testing and dual-color SISH support and The Ho Chi Minh City Oncology Hospital, Vietnam, for HER2 FISH testing support.

REFERENCES
Prognostic Significance of a Micropapillary Pattern in Pure Mucinous Carcinoma of the Breast: Comparative Analysis with Micropapillary Carcinoma

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Jung Yeon Kim · Guhyun Kang
Geumhee Gwak1 · Inseok Park1

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Background: Mucinous carcinoma of the breast is an indolent tumor with a favorable prognosis; however, micropapillary features tend to lead to aggressive behavior. Thus, mucinous carcinoma and micropapillary carcinoma exhibit contrasting biologic behaviors. Here, we review invasive mucinous carcinoma with a focus on micropapillary features and correlations with clinicopathological factors.

Methods: A total of 64 patients with invasive breast cancer with mucinous or micropapillary features were enrolled in the study. Of 36 pure mucinous carcinomas, 17 (47.2%) had micropapillary features and were termed mucinous carcinoma with micropapillary features (MUMPC), and 19 (52.8%) had no micropapillary features and were termed mucinous carcinoma without micropapillary features. MUMPC were compared with 15 invasive micropapillary carcinomas (IMPC) and 13 invasive ductal and micropapillary carcinomas (IDMPC).

Results: The clinico-pathological factors of pure mucinous carcinoma and MUMPC were not significantly different. In contrast to IMPC and IDMPC, MUMPC had a low nuclear grade, lower mitotic rate, higher expression of hormone receptors, negative human epidermal growth factor receptor 2 (HER2) status, lower Ki-67 proliferating index, and less frequent lymph node metastasis (p < 0.05). According to univariate analyses, progesterone receptor, HER2, T-stage, and lymph node metastasis were significant risk factors for overall survival; however, only T-stage remained significant in a multivariate analysis (p < 0.05).

Conclusions: In contrast to IMPC and IDMPC, the micropapillary pattern in mucinous carcinoma does not contribute to aggressive behavior. However, further analysis of a larger series of patients is required to clarify the prognostic significance of micropapillary patterns in mucinous carcinoma of the breast.

Key Words: Breast; Micropapillary; Mucinous; Prognosis

Mucinous carcinoma of the breast, believed to be an indolent tumor, affects older females. Pure mucinous carcinoma with a > 90% mucinous component has a better prognosis than that of mixed mucinous carcinoma. This carcinoma tends to follow an indolent course with infrequent lymphatic or hematogenic dissemination and to have a favorable prognosis.1-4 Invasive micropapillary carcinoma (IMPC) was first reported by Siriaunkgul and Tavassoli5 in 1993 as a rare subtype of breast carcinoma. IMPC is characterized by tumor cells arranged in tubules with a small or obliterated lumen, which extensively penetrate the lymphatic or vascular space, leading to a high frequency of lymph node metastasis.6-14 In a previous study, axillary lymph node metastases were found in all of 27 patients at the initial diagnosis. Twelve of these patients were followed up, and six died at a mean of 22 months after the initial treatment.12 It is important for pathologists to identify micropapillary formations, as they indicate the potential for aggressive tumor behavior and influence the choice of therapy. Thus, the micropapillary and pure mucinous subtypes of invasive carcinoma are opposites in terms of their biological behavior.15 These two patterns sometimes coexist in the same tumor. Tumor cells in mucinous carcinoma can exhibit different patterns, such as cords, trabeculae, cribriform structures, and solid lobules. Previous studies have suggested that the micropapillary pattern in cases of mucinous carcinoma is associated with aggressive tumor behavior.16-18 The pathogenetic association of this mucinous micropapillary carcinoma (MUMPC) with pure mucinous carcinoma is unclear.

The aim of this study was to characterize the clinicopathological parameters of MUMPC and to compare them with those of pure mucinous carcinoma, IMPC, and invasive ductal and micropapillary carcinoma (IDMPC). Moreover, the prognostic significance of micropapillary features in breast carcinoma was
determined.

**MATERIALS AND METHODS**

We analyzed 64 patients diagnosed with MUMPC, IMPC, IDMPC, or mucinous carcinoma without micropapillary features (MUC). These 64 cases were diagnosed and treated for breast carcinoma at Inje University Sanggye Paik Hospital between 1997 and 2012. Only cases with tumors containing a ≥ 90% mucin component were defined as mucinous carcinoma. Morphologically, micropapillae were defined as clusters of tumor cells separated from surrounding stroma by clear spaces and that exhibited reverse polarity, also known as an “inside-out” growth pattern, whereby the apical pole of the cells faces the stroma rather than the luminal surface.**\(^1\)** MUMPC was defined as a tumor in which micropapillary features constituted >50% of the tumor epithelial components (Fig. 1). Patient age, tumor stage by TNM classification, lymph node metastasis, nuclear grade, mitotic rate, estrogen receptor (ER) status, progesterone receptor (PR) status, human epidermal growth factor receptor 2 (HER2) status, Ki-67 proliferating index, molecular subtype, and survival were recorded. The following antibodies were used for immunohistochemistry (IHC): ER (SP1, Dako, Carpinteria, CA, USA), PR (SP2, Dako), HER2 (HercepTest, Dako), Ki-67 (MIB-1, Dako), cytokeratin (CK) 5/6 (D5/16, Dako), and epidermal growth factor receptor (EGFR; E30, Dako). Cases were classified as ER+, PR+, and HER2-positive or negative, and the latest American Society of Clinical Oncology guidelines for the detection and scoring of ER, PR, and HER2 were followed.**\(^2\)** Cases with a HER2 IHC grade of 2+ subsequently underwent evaluation of HER2 amplification by fluorescence in situ hybridization (PathVysion, Abbott, Des Plaines, IL, USA) and were designated as positive or negative. A high Ki-67 proliferation index was noted in eight cases of MUC (42.1%) and 10 cases of MUMPC (58.8%). Among the MUC and MUMPC cases, 11 (57.9%) and seven cases (41.2%) were luminal A, respectively, and eight (42.1%) and 10 cases (58.8%) were luminal B. Neither the HER2 nor basal-like subtype was detected in MUC or MUMPC. Among the MUC cases, 11 cases (57.5%) were defined as follow: luminal A (ER+ and/or PR+, HER2+, Ki-67 low), luminal B (ER+ and/or PR+, HER2+/ ER+ and/or PR+, HER2+, Ki-67 high), HER2-positive (ER+, PR+, HER2+), and basal-like (ER+, PR+, HER2+, EGFR+ and/or CK5/6).**\(^3\)** Patients underwent either mastectomy or breast conserving surgery with lymph node dissection. Adjuvant chemotherapy or hormonal therapy was routinely considered. All patients underwent mammography, physical examination, chest X-ray, and breast ultrasonography during follow-up of 6 to 96 months. This study was approved by the Institutional Review Board of our hospital (SGPAIN 2016-04-002).

The following clinical and pathological factors were assessed: age at diagnosis, nuclear grade (a one-level increase on a 1–3 scale), mitotic rate (a one-level increase on a 1–3 scale), ER expression (positive vs negative), PR expression (positive vs negative), HER2 amplification (positive vs negative) and the Ki-67 proliferation index (high vs low). Molecular tumor subtypes were categorized as luminal A, luminal B, basal-like, or HER2 positive. The clinicopathological features of MUC, MUMPC, IMPC, and IDMPC cases were compared. Pearson’s chi-square test, Student’s t test, and ANOVA were used to evaluate discrete and quantitative variables. Among the clinicopathological variables, the potential risk factors for overall survival (OS) were examined using univariate and multivariate logistic regression analyses. Statistical analyses were performing using IBM SPSS Statistics ver. 20 software (IBM Corp., Armonk, NY, USA). Statistical significance was determined using the likelihood ratio test and accepted at values of *p* < .05.

**RESULTS**

The 64 cases comprised 19 (29.7%) with MUC, 17 (26.6%) with MUMPC, 15 (23.4%) with IMPC, and 13 (20.3%) with IDMPC. All cases were female and aged 29–79 years (mean, 51.8 ± 12.8 years).

Both MUC and MUMPC showed low nuclear grades and low mitotic rates. In both groups, all tumor cells were positive for ER, irrespective of the presence of micropapillary features. All but one of the MUC cases revealed strong PR expression but no HER2 amplification. All but one of the MUMPC cases were positive for PR and negative for HER2 amplification. A high Ki-67 proliferation index was noted in eight cases of MUC (42.1%) and 10 cases of MUMPC (58.8%). Among the MUC and MUMPC cases, 11 (57.9%) and seven cases (41.2%) were luminal A, respectively, and eight (42.1%) and 10 cases (58.8%) were luminal B. Neither the HER2 nor basal-like subtype was detected in MUC or MUMPC. Among the MUC cases, 11 cases (57.5%) were...
were T1, seven cases (36.8%) were T2, and a case (5.3%) was T3. Among the MUMPC, six cases (35.3%) were T1, seven cases (41.2%) were T2, and four cases (23.5%) were T3. Metastasis to the axillary lymph node was detected in two cases of MUC (10.5%) and four cases of MUMPC (23.5%). The surrounding breast parenchyma showed changes suggestive of ductal carcinoma in situ in 13 cases (36.1%), comprising eight cases of MUC and five of MUMPC (Table 1).

Compared with IMPC and IDMPC, nuclear grade and mitotic rate were lower in MUMPC (p < .01). MUMPC showed lower frequency of ER and PR negativity (p = .04) and HER2 positivity (p = .01). The Ki-67 proliferation index of the MUMPC cases was lower than that of the IMPC and IDMPC cases (p < .01). Upon comparison of MUC and MUMPC, IMPC and IDMPC revealed less frequent luminal A and more frequent HER2 and basal-like subtypes (p < .01). Among the MUMPC cases, six cases (35.3%) were T1, seven cases (41.2%) were T2, and four cases (23.5%) were T3. Among the IMPC and IDMPC cases, six cases (40.0%) and four cases (30.8%) were T1, six cases (40.0%) and seven cases (53.8%) were T2, and three cases (20.0%) and two cases (15.4%) were T3, respectively. Four of the MUMPC cases (23.5%) exhibited lymph node involvement, while metastasis to the axillary lymph node was detected in 12 (80.0%) and nine (69.2%) cases of IMPC and IDMPC, respectively (p < .01) (Table 1). The surrounding breast parenchyma showed changes suggested of ductal carcinoma in situ in 19 cases (67.9%), comprising eight cases of IMPC and 11 of IDMPC.

Mean follow-up duration was 83 ± 51 months. Overall, PR and HER2 statuses, T-stage, and lymph node metastasis were correlated with death (Table 2). Univariate analysis using a logistic regression model revealed that OS was significantly associated with PR and HER2 statuses, T-stage, and lymph node metastasis (Table 3). However, on multivariate analysis, only T-stage was significantly associated with OS (Table 3).

### DISCUSSION

Pure mucinous carcinoma of the breast has a good prognosis with a low rate of regional lymph node metastasis and excellent survival, in contrast to invasive ductal carcinoma of no special type.

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**Table 1. Comparisons of clinicopathological parameters among MUC, MUMPC, IMPC, and IDMPC**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>MUC (n = 19)</th>
<th>MUMPC (n = 17)</th>
<th>IMPC (n = 15)</th>
<th>IDMPC (n = 13)</th>
<th>p-value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, mean (yr)</td>
<td>52.7 ± 14.0</td>
<td>53.9 ± 11.8</td>
<td>45.9 ± 11.7</td>
<td>54.3 ± 12.6</td>
<td>.22b</td>
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<td>6 (31.6)</td>
<td>1 (5.9)</td>
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<td>0</td>
</tr>
<tr>
<td></td>
<td>Intermediate</td>
<td>11 (57.9)</td>
<td>14 (82.4)</td>
<td>4 (26.7)</td>
<td>4 (30.8)</td>
</tr>
<tr>
<td></td>
<td>High</td>
<td>2 (10.5)</td>
<td>2 (11.8)</td>
<td>11 (73.3)</td>
<td>9 (69.2)</td>
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<tr>
<td>Mitosis</td>
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<td>16 (84.2)</td>
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<td>2 (15.4)</td>
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<td>4 (23.5)</td>
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<td>10 (66.7)</td>
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<td>17 (100)</td>
<td>10 (66.7)</td>
<td>9 (69.2)</td>
</tr>
<tr>
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<td>1 (5.9)</td>
<td>7 (46.7)</td>
<td>5 (38.5)</td>
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<tr>
<td></td>
<td>Positive</td>
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<td>16 (94.1)</td>
<td>8 (53.3)</td>
<td>8 (61.5)</td>
</tr>
<tr>
<td>HER2</td>
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<td>18 (94.7)</td>
<td>16 (94.1)</td>
<td>9 (60.0)</td>
<td>8 (61.5)</td>
</tr>
<tr>
<td></td>
<td>Positive</td>
<td>1 (5.3)</td>
<td>1 (5.9)</td>
<td>6 (40.0)</td>
<td>5 (38.5)</td>
</tr>
<tr>
<td>Ki-67</td>
<td>Low</td>
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<td>7 (41.2)</td>
<td>1 (6.7)</td>
<td>1 (7.7)</td>
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<tr>
<td></td>
<td>High</td>
<td>8 (42.1)</td>
<td>10 (58.8)</td>
<td>14 (93.3)</td>
<td>12 (92.3)</td>
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<tr>
<td>MS</td>
<td>LA</td>
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<td>6 (35.3)</td>
<td>1 (6.7)</td>
<td>1 (7.7)</td>
</tr>
<tr>
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<td>LB</td>
<td>8 (42.1)</td>
<td>11 (64.7)</td>
<td>9 (60.0)</td>
<td>8 (61.5)</td>
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<tr>
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<td>6 (40.0)</td>
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<td>2</td>
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<td>7 (41.2)</td>
<td>6 (40.0)</td>
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<td>4 (23.5)</td>
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<td>Presence</td>
<td>2 (10.5)</td>
<td>4 (23.5)</td>
<td>12 (80.0)</td>
<td>9 (69.2)</td>
</tr>
</tbody>
</table>

Values are presented as number (%).

MUC, mucinous carcinoma without micropapillary feature; MUMPC, mucinous carcinoma with micropapillary feature; IMPC, invasive micropapillary carcinoma; IDMPC, mixed invasive ductal and micropapillary carcinoma; NG, nuclear grade; ER, estrogen receptor; PR, progesterone receptor; HER2, human epidermal growth factor receptor 2; MS, molecular subtype; LA, luminal A; LB, luminal B; LNM, lymph node metastasis.

*Chi-square test; ANOVA test.
A micropapillary architecture in breast carcinoma has been identified in pure mucinous carcinoma, although its prognostic significance is unclear. In this study, 17 of 36 cases of mucinous carcinoma (47.2%) had micropapillary features. As reported in a previous study, these micropapillary subtypes of mucinous carcinoma impact patient survival via their propensity for nodal metastases, depending on the amount of mucin within the tumor. The study also demonstrated that even tumors classified as MUMPC can lead to IMPC-type metastasis. Prior studies have reported that mucinous carcinomas result in nonmucinous or ductal metastasis. However, the results are conflicting. In the current study, four of 17 MUMPC cases (23.5%) had lymph node metastases of the ductal, micropapillary, or mucinous type. MUC and MUMPC were not significantly influenced by the presence of micropapillary features. The micropapillary pattern in mucinous carcinoma indicates a possible histogenetic association with IMPC. Indeed, the ability of IMPC to undergo at least partial mucinous differentiation has been reported.

We separately reviewed 45 cases of breast carcinoma with micropapillary features, of which 17 cases were MUMPC, 15 IMPC, and 13 IDMPC, and found that nuclear grade; mitotic rate; ER, PR, HER2, and Ki-67 expression; and lymph node status differed significantly among the subtypes (p < .05). MUMPC and IMPC have been reported to have similar high nuclear grades, with 70%–80% having florid mitotic activity. This suggests that MUMPC and IMPC are components of the same spectrum; they show similar nuclear grades and vary only in their mucin content. In contrast, compared with patients with IMPC, those with MUMPC have a better prognosis irrespective of tumor stage. In the present study, MUMPC cases had lower and intermediate nuclear grades and a lower mitotic rate than those of IMPC cases. These results indicate that MUMPC exhibits a level of aggressiveness intermediate to those of pure mucinous carcinoma and IMPC.

Previous studies have reported high HER2 expression and low hormone receptor expression in IMPC of the breast. IMPC has a high propensity for lymph node metastasis and more frequent involvement of the lymph nodes compared with invasive ductal carcinoma. In this study, four cases of MUMPC (23.5%) had lymph node metastasis; however, synchronous lymph node metastasis was detected in 12 cases of IMPC (80.0%) and nine

### Table 2. Clinicopathological parameters related to death

<table>
<thead>
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<th>Parameter</th>
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<th>Dead</th>
<th>p-value*</th>
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<td>53.0 ± 16.0</td>
<td>.75</td>
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<td>MUMPC</td>
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</table>

MUC, mucinous carcinoma without micropapillary feature; MUMPC, mucinous carcinoma with micropapillary feature; IMPC, invasive micropapillary carcinoma; IDMPC, mixed invasive ductal and micropapillary carcinoma; NG, nuclear grade; ER, estrogen receptor; PR, progesterone receptor; HER2, human epidermal growth factor receptor 2; MS, molecular subtype; LA, luminal A; LB, luminal B; LNM, lymph node metastasis.

*Chi-square test; t test (2-tailed).
cases of IDMPC (69.2%). In IMPC, histologic grade, lymphatic vessel density, and lymphocyte infiltration might influence lymph node metastasis.9,16,33,35,36 In the present study, MUMPC tumors showed high rates of hormone receptor expression: ER and PR were expressed in 17 (100%) and 16 (94.1%) of 17 cases, respectively; one case (5.9%) was positive for HER2 amplification. A high rate of hormone receptor expression and a low rate of HER2 amplification suggested good prognosis. Hsu and Shaw37 reported no HER2 amplification in mucinous breast cancer, suggesting that HER2 is rarely involved in its tumorigenesis; low HER2 amplification might also contribute to a better prognosis of this cancer.

Most cases of IMPC are associated with nodal metastases and a poor prognosis.12,13,34,38,39 There were typically multiple metastases, with 51% of cases having three or more positive lymph nodes.13,35 The average number of metastatic lymph nodes was shown to be 10.7.39 In addition, lymphatic and vascular invasion has been reported in 33%–67% of cases.9,12 In this study, 12 of 15 IMPC cases (80.0%) exhibited synchronous axillary lymph node metastasis. Of these 12 cases, multiple lymph node metastases (> 4) were noted in five cases (41.7%). The mechanism underlying the high incidence of lymph node metastasis in IMPC is unclear, and the intrinsic subtype is considered an important prognostic factor.40,41

The prognosis of luminal A type breast cancer is markedly superior to that of HER2-positive and triple-negative breast cancers. A previous study reported that MUMPC revealed more prevalent luminal B or HER2 subtypes than MUC.42 However, we could not identify the clinical significance of breast carcinoma molecular subtype since there was no HER2 positivity in either MUC or MUMPC cases in this study.

In conclusion, our findings suggest heterogeneous biological behavior among tumors with a micropapillary architecture. Morphologically, MUMPC shares features with both mucinous carcinoma and IMPC, which could result in its intermediate clinical behavior. Although IMPC and IDMPC are aggressive tumors, the presence of micropapillary features in MUMPC was not associated with poorer prognosis. It can be postulated that MUMPC has a more favorable prognosis than that of IMPC, and that the micropapillary pattern in mucinous carcinoma does not contribute to its aggressive behavior. However, further analyses involving a larger cohort are required to clarify the pathogenetic relationships among these tumor types. Furthermore, the classical tumor stage was the strongest predictor of prognosis.

**Conflicts of Interest**
No potential conflict of interest relevant to this article was reported.

**Acknowledgments**
This work was supported by the 2012 Inje University research grant for Kyeongmee Park.

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**Table 3.** Univariate and multivariate analyses of the associations between clinicopathological parameters and overall survival using logistic regression model

<table>
<thead>
<tr>
<th>Category</th>
<th>Univariate RR (95% CI)</th>
<th>p-value</th>
<th>Multivariate RR (95% CI)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>MUMPC</td>
<td>1.0</td>
<td>Reference</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>IMPC vs MUMPC</td>
<td>1.2 (0.2–6.9)</td>
<td>.87</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>IDMPC vs MUMPC</td>
<td>1.4 (0.2–8.4)</td>
<td>.71</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>NG</td>
<td>2.5 (0.7–8.7)</td>
<td>.15</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Mitosis</td>
<td>1.9 (0.8–4.1)</td>
<td>.13</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>ER</td>
<td>0.3 (0.1–1.2)</td>
<td>.09</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>PR</td>
<td>0.2 (0.1–0.7)</td>
<td>.02</td>
<td>0.7 (0.1–6.6)</td>
<td>.78</td>
</tr>
<tr>
<td>HER2</td>
<td>7.3 (1.6–33.0)</td>
<td>.01</td>
<td>1.1 (0.1–11)</td>
<td>.92</td>
</tr>
<tr>
<td>Ki-67</td>
<td>4.2 (0.5–36.0)</td>
<td>.19</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>MS</td>
<td>1.0</td>
<td>Reference</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>LA</td>
<td>0.0</td>
<td>.10</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>LB vs LA</td>
<td>4.8 (0.6–41.0)</td>
<td>.15</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>HER2 vs LA</td>
<td>1.2 (0.1–13.0)</td>
<td>.88</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Basal-like vs LA</td>
<td>15 (3.2–72.0)</td>
<td>.001</td>
<td>14 (2.1–88)</td>
<td>.006</td>
</tr>
<tr>
<td>T-stage</td>
<td>15 (1.8–130.0)</td>
<td>.01</td>
<td>9 (0.6–149)</td>
<td>.11</td>
</tr>
</tbody>
</table>

RR, relative risk; 95% CI, 95% confidence interval; MUMPC, mucinous carcinoma with micropapillary feature; IMPC, invasive micropapillary carcinoma; IDMPC, mixed invasive ductal and micropapillary carcinoma; NG, nuclear grade; ER, estrogen receptor; PR, progesterone receptor; HER2, human epidermal growth factor receptor 2; MS, molecular subtype; LA, luminal A; LB, luminal B; LNM, lymph node metastasis.
REFERENCES


The Use of the Bethesda System for Reporting Thyroid Cytopathology in Korea: A Nationwide Multicenter Survey by the Korean Society of Endocrine Pathologists

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Key Words: Thyroid; Biopsy, Fine-needle; Bethesda system; Cytopathology; Korea

Background: The Bethesda System for Reporting Thyroid Cytopathology (TBSRTC) has standardized the reporting of thyroid cytology specimens. The objective of the current study was to evaluate the nationwide usage of TBSRTC and assess the malignancy rates in each category of TBSRTC in Korea. Methods: Questionnaire surveys were used for data collection on the fine needle aspiration (FNA) of thyroid nodules at 74 institutes in 2012. The incidences and follow-up malignancy rates of each category diagnosed from January to December, 2011, in each institute were also collected and analyzed. Results: Sixty out of 74 institutes answering the surveys reported the results of thyroid FNA in accordance with TBSRTC. The average malignancy rates for resected cases in 15 institutes were as follows: nondiagnostic, 45.6%; benign, 16.5%; atypical of undetermined significance, 68.8%; suspicious for follicular neoplasm (SFN), 30.2%; suspicious for malignancy, 97.5%; malignancy, 99.7%. Conclusions: More than 80% of Korean institutes were using TBSRTC as of 2012. All malignancy rates other than the SFN and malignancy categories were higher than those reported by other countries. Therefore, the guidelines for treating patients with thyroid nodules in Korea should be revisited based on the malignancy rates reported in this study.
The Bethesda System for Reporting Thyroid Cytopathology (TBSRTC) was developed to provide uniform terminology and diagnostic criteria for reporting thyroid fine needle aspiration (FNA) and to relate these cytopathologic diagnoses to clinical management. TBSRTC describes six categories for the diagnosis and reporting of thyroid FNAs, each with an assigned "risk of malignancy" and associated recommendations for clinical management. Since its implementation, many studies have demonstrated that TBSRTC has improved the quality of reporting by decreasing the number of ambiguous reports, increasing the positive predictive value of malignancy in thyroid glands that are operated, and decreasing the rates of surgery for benign thyroid nodules.2-5

TBSRTC terminology was incorporated into the 2009 guidelines of the Korean Thyroid Association (KTA) for management of patients with thyroid nodules and thyroid cancer. Several studies in Korea have demonstrated that TBSRTC well stratifies the malignancy risk by diagnostic categories and provides clinicians with useful information on the management of thyroid nodules.7-10 However, a wide variation in the incidence and malignant risk of each diagnostic category (DC; from here on, we will use DC as the abbreviation of diagnostic category), especially of the atypia of undetermined significance/follicular lesion of undetermined significance (AUS/FLUS) categories, has been reported.11,12 Moreover, the malignant risk of the categories has been reported to be higher than the implied risk of malignancy in TBSRTC.7,9,13-15

In the current study, we aimed to evaluate the extent of implementation of TBSRTC for reporting thyroid FNAs and its impact on managing patients with thyroid nodules by assessing the incidence and malignancy rate in each category of TBSRTC in Korea.

**MATERIALS AND METHODS**

**Design and data collection**

A questionnaire designed by the Endocrine Pathology Study Group of the Korean Society of Pathologists to gather data concerning the extent of implementation of TBSRTC and its impact on clinical practice was mailed to 211 pathology laboratories in the year 2012. Participants were asked questions related to the year of implementation of TBSRTC, number of thyroid aspiration cases in 2010 and 2011, usage of liquid-based cytology, types of liquid-based cytology used, and usage of core needle biopsy. Subsequently, a questionnaire was mailed to 41 institutions that had agreed to provide the statistics of thyroid FNA including the incidence, operation rate and malignancy rate of each diagnostic categories of TBSRTC. All the participants’ answers were collated. Among them, sixteen institutes reported the incidence of each category (unsatisfactory, benign, AUS, follicular neoplasm [FN], suspicious for malignancy [SM], and malignancy) in TBSRTC diagnosed from January to December, 2011. In total, 42,132 cases were analyzed. Follow-up malignancy rates of each category in 15 institutes were also collected and analyzed. Malignancy rate of each category was estimated based on surgically resected specimens. This survey was approved by the Institutional Review Board of Seoul National University Bundang Hospital, waiving the requirement for informed consent (IRB No. B-1212/182-304).

**Statistics**

The statistical significance of the data was assessed using SPSS ver. 21.0 for Windows (IBM Corp., Armonk, NY, USA). The sensitivity, specificity, positive predictive value, and negative predictive value were calculated in two different settings considering thyroid FNA as a “screening test.” First, we calculated these parameters in malignant thyroid tumors. Surgical specimens which had been cytologically classified in the DC II of TBSRTC and diagnosed histologically as benign were considered to be true-negative samples. Surgical specimens whose cytologic diagnoses were in the remaining diagnostic categories (DC V, DC VI) and histologically proved to be malignant neoplasms were considered to be true-positives. The false-positive category included cases that had been diagnosed as SM or malignant (DC V, DC VI), but later confirmed histologically as benign (normal, non-neoplastic benign lesion, or benign neoplasm). The false-negative cases included those diagnosed as benign on FNA but confirmed as malignant upon surgical excision. In addition, those parameters were calculated for thyroid neoplasms including benign (such as follicular adenoma) and malignant. For this calculation, follow-up surgical specimens diagnosed as non-neoplastic benign disease or normal, which were interpreted as the DC II of TBSRTC, were considered to be true-negative samples, and the remaining diagnostic categories (DC IV, DC V, and DC VI), which proved to be either benign or malignant neoplasms by histologic examination of the surgical specimens, were considered to be true-positive. The false-positive category included cases that were diagnosed as FN, SM, and malignant but later confirmed histologically as benign non-neoplastic or normal. The false-negative cases included those diagnosed as DC II on FNA but confirmed as a true neoplasm upon surgical excision. The DC I and DC III (AUS/FLUS) categories were excluded from the statistical analysis because these diagnoses usually led to a repeat FNA rather than to surgical excision; moreover, these cases could not be categorized as either positive or negative.
RESULTS

General survey questions

Questionnaire surveys were sent out to 77 institutes of pathology with controlled quality authorized by the Korean Cytology Conference. Three institutes were excluded because they were sending out their cytology specimens to consultants for the diagnosis, and answers from 74 institutes were included in the assessment. The survey results are summarized in Table 1.

Among the 74 institutes, 60 used TBSRTC for their diagnostic categorization of thyroid FNA at the time of survey. The starting year of using this classification varied from 2008 to 2012. The usage increased particularly from 2010 to 2012, 3 to 5 years after the conference in Bethesda. In Korea, the corresponding guidelines of the KTA for management of patients with thyroid nodules and thyroid cancer were introduced in 2009, which might attribute in the subsequent sharp rise in the usage of TBSRTC.

Forty-one out of the 74 institutes used conventional method for FNA, whereas 11 institutes used liquid-based method; the remaining 22 adopted both. Out of the 33 institutes using liquid-based preparation, 10 had started doing so in 2010. Regarding the preparation method for liquid-based FNA, two representative commercial methods, Thin-prep and Sure-path, were predominant: almost every institute chose one or the other method. Core needle biopsy was being done in 21 institutes (27% of the 77). Among them, four institutes (5% of the 77) used both core needle biopsy and FNA as the primary investigation method of thyroid nodules. In thirty-eight institutes (51.4%), FNA was done by radiologists, followed by clinicians of internal medicine, surgeons, pathologists, and thyroid-specialized clinicians.

The survey indicated that the majority of the FNA specimens in Korea (60/74, 81%) were being diagnosed as per TBSRTC as of 2007. Moreover, the preparation method also shifted from conventional to liquid-based, the latter adopted in about 43% (33/77) including the institutes performing both.

Incidence and malignancy rate of each DC of TBSRTC in Korea

Sixteen out of 74 institutes answering the survey reported the results of thyroid FNA by TBSRTC. The mean distribution of six categories from 42,132 cases of 16 institutes was as follows: 11.1% nondiagnostic, 62.3% benign, 9.7% AUS, 0.9% FN, 6.7% SM, and 9.1% malignancy (Table 2). The mean operation rates of each category were as follows: 7.5% nondiagnostic, 2.4% benign, 20.2% AUS, 43.7% FN, 59.9% SM, and 69.2% malignancy (Table 3).

The average of malignancy rates for resected cases of 15 institutes are shown in Table 4: nondiagnostic, 45.6%; benign, 16.5%; AUS, 68.7%; FN, 30.2%; SM, 97.5%; and malignancy, 99.7%. Relatively high malignancy rates were noted in nondiagnostic and benign categories, which might have resulted from false-negative results of clinically suspicious nodules. Overall, the false-positive rate of the SM and malignancy category combined was 1.23%.

Follow-up histologic diagnosis of each categories of TBSRTC

The final histological diagnoses upon resection are listed in Table 5. Each DC is sub-classified according to the recommendation of TBSRTC and follow-up histologic diagnoses are summarized by the sub-classification of each DC.

Of 4,599 cases, 247 were operated in the non-DC (DC I) and
128 cases (51.8%) were revealed as malignant nodules. The malignant cases mainly consisted of papillary thyroid carcinoma (121 cases), followed by five cases of follicular carcinoma, one case of poorly differentiated carcinoma, and one case of lymphoma. The benign category (DC II) had 422 cases of follow-up operation out of 21,399 cases. Among them, 320 (75.8%) were revealed as benign nodules and 102 cases (24.2%) were malignant: 89 papillary thyroid carcinomas, 12 follicular carcinomas, and one medullary carcinoma. Out of the 3,708 AUS-categorized cases, 722 had follow-up surgical resection (Table 3). The 514 cases (71.2%) with malignant histologic diagnoses consisted of 471 cases of papillary carcinoma, 25 medullary carcinomas, two poorly differentiated carcinomas, and one case of lymphoma. The remaining 208 (28.8%) cases were proved to be benign lesions. The distribution of each diagnostic category of TBSRTC from 16 institutions is shown in Table 2.

<table>
<thead>
<tr>
<th>Diagnostic category</th>
<th>Total</th>
<th>Range (%)</th>
<th>Mean ± SD (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>UNS/ND</td>
<td>1,040</td>
<td>10.4–21.2</td>
<td>11.1 ± 5.3</td>
</tr>
<tr>
<td>Benign</td>
<td>3,397</td>
<td>9.7–19.8</td>
<td>9.7 ± 4.6</td>
</tr>
<tr>
<td>AUS</td>
<td>493</td>
<td>0.9–19.8</td>
<td>9.9 ± 4.6</td>
</tr>
<tr>
<td>SFM</td>
<td>1,178</td>
<td>2.0–16.6</td>
<td>6.7 ± 4.1</td>
</tr>
<tr>
<td>Malignant</td>
<td>521</td>
<td>0.8–17.1</td>
<td>9.1 ± 5.5</td>
</tr>
<tr>
<td>Total</td>
<td>7,084</td>
<td>17.8–59.7</td>
<td>32.8 ± 10.8</td>
</tr>
</tbody>
</table>

Values are presented as number (%).

TBSRTC of Thyroid Nodule in Korea

Diagnostic values for TBSRTC in Korea

Through the data obtained by the survey summarized above, sensitivity, specificity, positive predictive value, and negative predictive value were analyzed in two ways. First, we included only those in the benign category (DC II) as negative and malignant thyroiditis (Table 5). A total of 1,133 cases of DC V were histologically confirmed as benign nodules in 32 cases (2.8%) and malignancy in 1,101 cases (97.2%): 1,094 cases of papillary carcinoma and seven cases of medullary carcinoma (Table 5). Lastly, DC VI (malignancy) had 2,439 resected cases out of 3,343 FNAs. Overall, the final histologic diagnoses were well correlated with preoperative FNA diagnoses. Out of 2,439 cases, 2,419 (99.6%) with a preoperative diagnosis of papillary carcinoma on FNA sub-classification were proved to be papillary carcinoma upon resection. Only 20 out of 2,439 resected cases were non-papillary thyroid carcinoma lesions including one follicular carcinoma, three medullary carcinomas, two poorly differentiated carcinomas, one undifferentiated carcinoma, one metastatic carcinoma, and 12 various benign lesions (Table 5).

Table 2. The distribution of each diagnostic category of TBSRTC from 16 institutions

<table>
<thead>
<tr>
<th>Diagnostic category</th>
<th>Total</th>
<th>Range (%)</th>
<th>Mean ± SD (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Unsatisfactory (UNS)</td>
<td>1,040</td>
<td>10.4–21.2</td>
<td>11.1 ± 5.3</td>
</tr>
<tr>
<td>Non-diagnostic (ND)</td>
<td>493</td>
<td>0.9–19.8</td>
<td>9.9 ± 4.6</td>
</tr>
<tr>
<td>Atypia of undetermined significance (AUS)</td>
<td>1,178</td>
<td>2.0–16.6</td>
<td>6.7 ± 4.1</td>
</tr>
<tr>
<td>Follicular neoplasm (FNM)</td>
<td>521</td>
<td>0.8–17.1</td>
<td>9.1 ± 5.5</td>
</tr>
<tr>
<td>Total</td>
<td>7,084</td>
<td>17.8–59.7</td>
<td>32.8 ± 10.8</td>
</tr>
</tbody>
</table>

Values are presented as percentage (%).

TBSRTC, The Bethesda System for Reporting Thyroid Cytopathology; SD, standard deviation; UNS, unsatisfactory; ND, non-diagnostic; AUS, atypia of undetermined significance; FN, follicular neoplasm; SFM, suspicious for follicular neoplasm; SFM, suspicious for malignancy.
categories (DC V and VI) as positive. In this analysis, the sensitivity, specificity, positive predictive value and negative predictive value for malignant thyroid tumors were 97.19%, 87.91%, 98.76%, and 75.83%, respectively. Second, we included FN (DC IV) cases as positive and calculated each parameter for the neoplasm including both benign and malignant. In this case, the sensitivity, specificity, positive predictive value, and negative predictive value for neoplasm of thyroid were 97.14%, 72.89%,

Table 4. The malignancy rate by each diagnostic category of TBSRTC

<table>
<thead>
<tr>
<th>Diagnostic category</th>
<th>Range (%)</th>
<th>Mean ± SD (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>UNS/ND</td>
<td>0–100</td>
<td>46.5 ± 33.5</td>
</tr>
<tr>
<td>Benign</td>
<td>53/94</td>
<td>(94.3) ± (3.5)</td>
</tr>
<tr>
<td>FN/SFN</td>
<td>4/15</td>
<td>(0.6) ± (0.5)</td>
</tr>
<tr>
<td>SPM</td>
<td>1/1</td>
<td>(0.1) ± (0.1)</td>
</tr>
<tr>
<td>Malignant</td>
<td>271/272</td>
<td>94.3 ± 2.1</td>
</tr>
</tbody>
</table>

Values are presented as number (mal/op) (%).

TBSRTC, The Bethesda System for Reporting Thyroid Cytopathology; SD, standard deviation; UNS, unsatisfactory; ND, non-diagnostic; AUS, atypia of undetermined significance; FN, follicular neoplasm; SFN, suspicious for follicular neoplasm; SPM, suspicious for malignancy.

Table 5. Follow up surgical diagnosis by each DC of TBSRTC

<table>
<thead>
<tr>
<th>DC</th>
<th>No. of FNA</th>
<th>No. of operation</th>
<th>Surgical diagnosis</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Cyst</td>
<td>796</td>
<td>PTC, FTC, HTC, MTC, PDC, UC, SCC, MC, Lym, FA, HA, NH, LT, GT, O</td>
</tr>
<tr>
<td></td>
<td>Acellular</td>
<td>3,803</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Benign</td>
<td>4,599</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Total</td>
<td>19,179</td>
<td></td>
</tr>
<tr>
<td>II</td>
<td>Be</td>
<td>1,148</td>
<td></td>
</tr>
<tr>
<td></td>
<td>LT</td>
<td>49</td>
<td></td>
</tr>
<tr>
<td></td>
<td>O</td>
<td>1,023</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Total</td>
<td>21,399</td>
<td></td>
</tr>
<tr>
<td>III</td>
<td>AUS</td>
<td>3,708</td>
<td></td>
</tr>
<tr>
<td></td>
<td>IV FN</td>
<td>283</td>
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</tr>
<tr>
<td></td>
<td>Total</td>
<td>21,714</td>
<td></td>
</tr>
<tr>
<td>V</td>
<td>PTC</td>
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</tr>
<tr>
<td></td>
<td>MTC</td>
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<td></td>
</tr>
<tr>
<td></td>
<td>MC</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Lym</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td></td>
<td>O</td>
<td>12</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Total</td>
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<tr>
<td>VI</td>
<td>PTC</td>
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<td></td>
<td>PDC</td>
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<td>MTC</td>
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</tr>
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</tr>
<tr>
<td></td>
<td>SCC</td>
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</tr>
<tr>
<td></td>
<td>MC</td>
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<td></td>
</tr>
<tr>
<td></td>
<td>Lym</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td></td>
<td>O</td>
<td>7</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Total</td>
<td>3,343</td>
<td></td>
</tr>
</tbody>
</table>

DC, diagnostic category; TBSRTC, The Bethesda System for Reporting Thyroid Cytopathology; PTC, papillary thyroid carcinoma; FTC, follicular thyroid carcinoma; HTC, Hurthle cell thyroid carcinoma; MTC, medullary thyroid carcinoma; UC, undifferentiated carcinoma; SCC, squamous cell carcinoma; MC, metastatic carcinoma; Lym, lymphoma; FA, follicular adenoma; HA, Hurthle cell adenoma; NH, nodular hyperplasia; LT, lymphocytic thyroiditis; GT, granulomatous thyroiditis; O, others; Be, benign; AUS, atypia of undetermined significance; FN, follicular neoplasm; PDC, poorly differentiated carcinoma.
96.78%, and 75.83%, respectively.

**DISCUSSION**

TBSRTC was proposed in October 2007 in Bethesda, Maryland, and published in 2009;1 TBSRTC was then immediately introduced in the 2009 guidelines of the KTA.6,16 Despite the information about TBSRTC having been globally disseminated, the status of its usage in clinical practice in Korea has been unknown. This questionnaire provides a cross-sectional view, based on the collective data from participating pathology laboratories, of the degree of implementation of TBSRTC in Korea and evaluates its impact on daily practice by assessing the incidence and malignant risk of each DC at each pathology laboratory using TBSRTC.

Overall, the data shows that TBSRTC is well implemented in Korea. As of July 2012 (when the questionnaire was mailed), 81% (60 out of 74) of the participating pathology laboratories were already using TBSRTC, which is higher than the rate given in published data from the College of American Pathologists.17 In addition, the data demonstrates interesting technical points. Fifty-seven percent of the laboratories are using conventional alcohol-fixed direct smears as a preparatory method for thyroid FNA; in contrast, 14.9% of the laboratories are using liquid-based cytology and 29.7% are using both. Overall, 44.6% of the laboratories are using liquid based cytology. Twenty-one out of 74 pathology laboratories (27%) received thyroid core needle biopsies; among them, four institutes used both core needle biopsy and FNA as the primary diagnostic test for thyroid nodules. Although core needle biopsy of thyroid nodules is emerging as an effective alternative method, the primary use of core needle biopsy in a thyroid nodule is quite unusual. For example, current American Thyroid Association thyroid nodule management guidelines and the Society of Radiologists in Ultrasound Consensus Statement have not included core biopsy as an evaluation tool of thyroid nodule, and some other studies suggest that after several non-diagnostic FNA, surgical excision should be considered appropriate.18,19

More importantly, our data showed some differences in the incidence and malignant risk of certain categories between TBSRTC1 or published studies from other countries7,12 and the current results. As shown in Table 2, lower incidence of DC IV (0.9 ± 0.8%) and higher incidence of DC III (9.7 ± 4.6%), DC V (6.7 ± 4.1%), and DC VI (9.1 ± 5.5%) were demonstrated. Moreover, the malignancy rates of DC I, II, III, and V (mean of malignancy rates: 46.5 ± 33.5, 16.5 ± 13.1, 68.7 ± 15.8, and 97.5 ± 4.5, respectively) in our study were higher than the risk of malignancy indicated by the original TBSRTC (1%–4%, DC I; 0%–3%, DC II; 5%–15%, DC III, and 60%–75%, DC V)1 and other previous studies.11,18-24 However, these findings are consistently shown in previous reports from Korea.80 These can be explained by several reasons. First, in Korea, the prevalence of papillary thyroid carcinoma is more than 95%, whereas that of follicular carcinoma is very low, at 3.2%,6 which has resulted in the lower incidence of DC IV and higher incidence of DC III, V, and VI. In addition, higher prevalence of papillary thyroid carcinoma might influence higher malignant rates of DC II, DC III, and DC V. Second, most institutes submitting their details of TBSRTC are university-based tertiary centers. In tertiary referral institutes, the patients who had been diagnosed with suspicious or malignant nodules at primary care centers were more likely to receive an operation despite a DC III or DC V diagnosis of FNA, especially those whose imaging studies strongly suggest malignancy. In the present study, malignancy rates were calculated based on the resected cases. The malignancy rate assessed either in the total fine needle aspiration cytology cases or in the resected cases cannot be said to represent the exact malignancy rate of each DC of TBSRTC. The former can miss unrecorded malignant tumors, and the latter can be exaggerated by selection bias, which may occur in this type of retrospective study. Therefore, our malignancy rates cannot be directly compared with the malignant risk recommended in TBSRTC. Lastly, because Korean citizens tend to consider false-positive results more significantly than false-negative results, cytopathologists in Korea may have a tendency to under-diagnose FNA of thyroid nodules.20

There are some limitations in the data derived from this questionnaire, especially those evaluating the incidence and malignancy rate of each DC in TBSRTC, as mentioned above. First, 14 out of 16 institutes submitting the incidence and malignancy rate of TBSRTC are university-based centers. Therefore, the results of this survey may not be truly representative of all kinds of practices; from community to academic. Second, all data are self-reported and not verified. Third, the data might not reflect the current status in certain aspects because the questionnaire was mailed in July 2012. The implementation of TBSRTC is more widespread in Korea at present, and liquid-based cytology and core needle biopsy are likely to have been adopted more widely. The usage of core needle biopsy in thyroid nodules has been actively studied in Korea.25

In conclusion, overall, TBSRTC has been well adopted in Korea, with more than 80% of institutes using TBSRTC as of 2012. However, ongoing education is still necessary to reduce
the variation of incidence and malignant rates of TBSRTC. Moreover, it would be better to modify reference values of malignancy rate of each category of TBSRTC and revisit the guidelines for treating patients with thyroid nodules in Korea based on the results of this study. Finally, it is highly recommended that each institute review their report of thyroid aspiration and have their own incidence and malignant rates for each DC.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

Acknowledgments

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REFERENCES

Metaplastic Carcinoma with Chondroid Differentiation Arising in Microglandular Adenosis

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Microglandular adenosis (MGA) of the breast is a rare benign proliferative lesion that is often difficult to differentiate clinicopathologically from invasive carcinoma.1,2 Breast carcinomas arising in MGA have occasionally been reported and metaplastic carcinoma is an extremely rare subtype of breast carcinoma–associated MGA.3-9 In Korea, Choi and Bae9 previously reported the first two cases of invasive carcinoma arising in MGA. Invasive carcinomas arising in MGA usually show a transition from typical MGA through atypical MGA to carcinoma. Herein, we report an unusual case of metaplastic carcinoma with chondroid differentiation associated with typical MGA. To the best of our knowledge, this is the first case of metaplastic carcinoma arising in MGA reported in Korea.

CASE REPORT

A 60-year-old female presented with a palpable breast mass in the left breast. Ultrasound-guided core needle biopsy revealed invasive carcinoma. Preoperative breast magnetic resonance imaging showed an irregular solid mass approximately 1.5 cm in size in the upper outer quadrant of the left breast, which showed good contrast enhancement. A conserving operation was performed. Grossly, a grey to white colored, hard mass with ill-defined borders measuring 1.5 cm × 1.2 cm × 1 cm was observed. Microscopically, the tumor was composed of an invasive carcinoma area and MGA area (Fig. 1A). Invasive carcinoma of no special type, grade 2 with an abrupt transition to chondroid matrix without an intervening spindle cell sarcomatoid component was observed (Fig. 1B). The chondroid matrix comprised 70% of the invasive area. Focal chondroid differentiation in the chondroid matrix area was found, thus the patient was diagnosed with metaplastic carcinoma with chondroid differentiation. In the MGA area, small round glands lined by a single layer of cuboidal epithelial cells were diffused throughout the fibrous or adipose breast tissues (Fig. 1C). Some of the glands were filled with a colloid-like secretion. No obvious architectural complexity or cytological atypia was present. MGA showed direct transition to metaplastic carcinoma without an intervening atypical MGA or ductal carcinoma in situ area. The glands in MGA were surrounded by a basement membrane, which was clearly demonstrated by reticulin staining. However, the basement membrane was disrupted around the invasive nests of the metaplastic carcinoma (Fig. 1D). The immunohistochemical profile of metaplastic carcinoma was nearly identical to that of MGA.
instances, the epithelial cells were positive for cytokeratin (CK) 7 and S-100 protein, but negative for estrogen receptor (ER), progesterone receptor (PR), HER2/neu, and epidermal growth factor receptor (EGFR) (Fig. 2A, B). Other basal-like markers such as CK 5/6 and nestin were negative. No myoepithelial cells were demonstrated on immunohistochemical stainings for both smooth muscle myosin heavy chain and p63 (Fig. 2C). Ki-67 and p53 were positive in MGA and metaplastic carcinoma but tended to show a more intense staining in metaplastic carcinoma (Fig. 2D). The percentage of p53 and Ki-67 positive cells was 5% and 3% in MGA and 90% and 45% in metaplastic carcinoma, respectively. This case was finally diagnosed as metaplastic carcinoma with chondroid differentiation arising in MGA.

No metastasis was found in the sentinel lymph nodes. After surgery, the patient received adjuvant chemoradiotherapy. After 14 months of follow-up, no evidence of recurrence was observed.

DISCUSSION

MGA of the breast is a rare, benign glandular proliferative lesion that mimics invasive carcinoma clinicopathologically.\(^1,2\) Carcinoma arising in MGA has previously been reported.\(^3,8\) The incidence of invasive carcinoma ranges from 23%–64% in patients with MGA;\(^3,8\) the high upper limit may be due to referral bias at institutions.\(^5\)

Invasive breast carcinomas arising in MGA have various histological features.\(^1\) Invasive carcinoma of no special type is the most common type of carcinoma arising in MGA. Specialized variants of carcinoma, including carcinoma with secretory differentiation, carcinoma with squamous differentiation, acinic cell carcinoma, and adenoid cystic carcinoma, have also been reported.\(^3,8\) Metaplastic carcinoma of the breast represents 0.2%–5% of all invasive carcinomas.\(^6\) Metaplastic carcinoma with mesenchymal differentiation is an extremely rare subtype of breast carcinoma arising in MGA.\(^8\) Structural transitions from MGA to atypical MGA and intraductal carcinoma and to invasive carcinoma are usually observed in invasive carcinomas arising in MGA.\(^3,8\)

In the present case, invasive carcinoma showed abrupt transition to chondroid matrix without an intervening spindle cell component. An intervening atypical MGA or ductal carcinoma in situ component between MGA and invasive carcinoma was not observed. Histologically, this case was consistent with metaplastic carcinoma with chondroid differentiation associated

Fig. 1. Metaplastic carcinoma with mesenchymal differentiation arising in microglandular adenosis (MGA). (A) MGA on the right shows direct transition to invasive carcinoma on the left. (B) Invasive carcinoma primarily composed of cord-like cells scattered in the chondromyxoid matrix with focal chondroid differentiation (inset). (C) Typical glands in MGA are lined by uniform cuboidal cells regularly spaced around a lumen containing a colloid-like secretion. (D) Basement membranes highlighted by the reticulin stain are preserved in the MGA area (inset) but disrupted in the invasive carcinoma area.
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with typical MGA.

The main histological difference between MGA and MGA-associated invasive carcinoma is the presence of a basement membrane. The glands in MGA are surrounded by a basement membrane that can be highlighted using reticulin staining, periodic acid-Schiff staining, and immunohistochemical stains for type IV collagen and laminin. However, the basement membrane in MGA-associated carcinoma is disrupted around invasive nests.

The immunohistochemical profile of invasive carcinoma arising in MGA is similar to that of MGA. No myoepithelial cells are found in MGA and related lesions. The epithelial cells are usually positive for CK 7 and S-100 protein, but negative for ER, PR, and HER2/neu. In the present case, all the epithelial cells in MGA and invasive carcinoma were strongly immunoreactive for S-100 protein and negative for ER, PR, and HER2/neu, which was consistent with a triple-negative immunoprofile. Although the positivity for basal-like markers such as EGFR, CK 5/6, and nestin has been described in MGA and associated lesions, epithelial cells in the present case lacked these markers. Several studies have reported a trend toward increased positivity for the p53 and proliferation marker Ki-67 together with increased severity of the lesions. An increase in p53 labelling index is observed from MGA to invasive carcinoma.

The treatment of carcinomas arising in MGA follows the same general guidelines for breast carcinomas. The prognosis of invasive carcinoma arising in MGA varies in the literature. Some authors reported that patients with invasive carcinoma arising in MGA have a relatively favorable prognosis. Another study, however, showed that the outcome for patients with invasive carcinoma arising in MGA ranged from favorable to unfavorable. In our patient, no axillary lymph node metastasis was found. Metaplastic carcinoma, the subtype of invasive carcinoma

![Fig. 2. Immunohistochemical stains. (A) S-100 protein is strongly positive in microglandular adenosis (MGA) and invasive carcinoma. Entrapped normal mammary gland is negative for S-100 protein (arrow). (B) Estrogen receptor (ER) is positive in entrapped normal mammary glands (arrow); however, MGA and invasive carcinoma are negative for ER. (C) No immunoreactivity for p63 is observed in the MGA area or invasive carcinoma area. Normal mammary glands are stained positively in myoepithelial cells (arrow). (D) An increase in p53 labelling index is observed from MGA to invasive carcinoma.](http://jpatholm.org/)
found in the present case, has lower response rates to conventional adjuvant chemotherapy and a worse clinical outcome than other forms of triple-negative breast cancers. Because metaplastic carcinoma arising in MGA is extremely rare, the prognosis of the lesion requires further characterization with more cases and longer follow-up.

Conflicts of Interest
No potential conflict of interest relevant to this article was reported.

REFERENCES
Mammary Carcinoma Arising in Microglandular Adenosis: A Report of Five Cases

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Key Words: Microglandular adenosis; Breast; Carcinoma

Microglandular adenosis (MGA) is a rare benign breast lesion, and it often mimics invasive carcinoma due to the lack of myoepithelium and the infiltrative growth pattern in the stroma.1 The unique immunohistochemical characteristic of MGA is the positivity for S-100 protein which is not generally positive in normal breast ductal and glandular cells.

Carcinomas arising in MGA (MGACA) are reported to comprise 27% of all MGA cases.2 They are well known for their various histologic manifestations, showing apparent tumor heterogeneity as in the mixed carcinomas.3-5 The other proliferative entity related to MGA, whose atypia is insufficient to be classified as carcinoma, is classified as atypical MGA (AMGA). Whether AMGA is a precursor of the MGACA is still under discussion and the evidence from molecular and genetic analyses for this topic are being accumulated.

This case report presents five cases of MGACA focusing on some of the interesting histologic features and immunohistochemical findings to further clarify the clinicopathologic characteristics of this rare entity.

Mammary carcinoma arising in microglandular adenosis (MGA) is extremely rare, and MGA is regarded as a non-obligate precursor of triple-negative breast cancer. We report five cases of carcinoma arising in MGA of the breast. All cases showed a spectrum of proliferative lesions ranging from MGA to atypical MGA, ductal carcinoma in situ or invasive carcinoma. Immunohistochemically, all cases were triple-negative and expression of S-100 protein gradually decreased as the lesions progressed from MGA to atypical MGA and carcinoma. Three cases showed acinic cell differentiation with reactivity to α1-antitrypsin, and one case was metaplastic carcinoma. During clinical follow-up, one patient developed local recurrence. Carcinoma arising in MGA is a rare but distinct subset of triple-negative breast cancer with characteristic histologic and immunohistochemical findings.

CASE REPORT

By searching the electronic database, we found five cases of MGACA (including an in situ lesion). The clinical characteristics of these five are summarized in Table 1. The median age of the patients at the time of the diagnosis was 47 years (range, 40 to 60 years). All patients underwent breast conserving surgery. Two patients had lymph node metastasis on pathologic examination and four received adjuvant chemo-radiation therapy. During follow-up, one patient (case 1) developed recurrence in the ipsilateral breast and axillary lymph nodes 1 year after the surgery. The rest of the patients had no evidence of disease and are being regularly followed up. This study was approved by the Institutional Review Board of Seoul National University Bundang Hospital (protocol # B-1609-364-701), and informed consent was waived.

The pathologic findings and the results of the immunohistochemical studies are shown in the Table 2. One patient (case 5) was diagnosed as ductal carcinoma in situ (DCIS) and the rest of the cases were all invasive carcinomas. The lesions in these five cases displayed the histologic spectrum ranging from MGA to...
AMGA and finally to MGACA (Fig. 1). The MGA areas had the typical structure: groups of small round glands infiltrating the stroma of the breast and adipose tissue. Glands were single-layered and lined by an intact basement membrane, which can be highlighted by periodic acid–Schiff (PAS) and reticulin stains. However, myoepithelium was absent, as revealed by the absence of immunohistochemical reactivity for calponin. In the lumens of the glands, eosinophilic secretions were common. In areas diagnosed as AMGA, glands became irregular in shape and cytologically, the nuclear pleomorphism or atypism increased as the amount of eosinophilic secretions decreased. Furthermore, the proliferating atypical cells filled the lumens completely to form carcinoma in situ lesions and finally grew to break the basement membrane to invade the stroma which can be visualized as interrupted and discontinued strands of fibers on PAS and reticulin stains.

Table 1. Clinical characteristics of the five cases of MGACA

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Case 1</th>
<th>Case 2</th>
<th>Case 3</th>
<th>Case 4</th>
<th>Case 5</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yr)</td>
<td>40</td>
<td>38</td>
<td>47</td>
<td>46</td>
<td>60</td>
</tr>
<tr>
<td>Location</td>
<td>Left</td>
<td>Left</td>
<td>Left</td>
<td>Left</td>
<td>Left</td>
</tr>
<tr>
<td>T stage (size of tumor, cm)</td>
<td>T1c (2.0)</td>
<td>T2 (2.3)</td>
<td>T1c (1.2)</td>
<td>T2 (2.5)</td>
<td>Tis (4.5)</td>
</tr>
<tr>
<td>N stage</td>
<td>N1a</td>
<td>N0</td>
<td>N1a</td>
<td>N0</td>
<td>N0</td>
</tr>
<tr>
<td>Type of surgery</td>
<td>BCS</td>
<td>BCS</td>
<td>BCS</td>
<td>BCS</td>
<td>BCS</td>
</tr>
<tr>
<td>Adjuvant CRTx</td>
<td>Received</td>
<td>Received</td>
<td>Received</td>
<td>Received</td>
<td>Not received</td>
</tr>
<tr>
<td>Disease status</td>
<td>Local recurrence</td>
<td>NED</td>
<td>NED</td>
<td>NED</td>
<td>NED</td>
</tr>
</tbody>
</table>

MGACA, carcinoma arising in microglandular adenosis; BCS, breast-conserving surgery; CRTx, chemo-radiation therapy; NED, no evidence of disease.

Table 2. Pathologic and immunohistochemical features of the five cases of MGACA

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Case 1</th>
<th>Case 2</th>
<th>Case 3</th>
<th>Case 4</th>
<th>Case 5</th>
</tr>
</thead>
<tbody>
<tr>
<td>Histologic subtype</td>
<td>Invasive carcinoma, NST</td>
<td>Invasive carcinoma, NST</td>
<td>Invasive carcinoma, NST</td>
<td>Metaplastic carcinoma in situ</td>
<td>Ductal carcinoma</td>
</tr>
<tr>
<td>Acinic cell differentiation</td>
<td>Absent</td>
<td>Present</td>
<td>Present</td>
<td>Absent</td>
<td>Present</td>
</tr>
<tr>
<td>Estrogen receptor</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Progesterone receptor</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>HER-2</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>S-100 protein</td>
<td>Decreased</td>
<td>Decreased</td>
<td>Decreased</td>
<td>Decreased</td>
<td>Intact</td>
</tr>
<tr>
<td>Cytokeratin 5/6</td>
<td>Positive</td>
<td>Positive</td>
<td>Positive</td>
<td>Negative</td>
<td>Positive</td>
</tr>
<tr>
<td>p53</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
<td>Positive</td>
<td>Negative</td>
</tr>
<tr>
<td>Ki-67 index (%)</td>
<td>50</td>
<td>20</td>
<td>30</td>
<td>80</td>
<td>5</td>
</tr>
<tr>
<td>α1-Antitrypsin</td>
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<td>Positive</td>
<td>Positive</td>
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<td>Chymotrypsin</td>
<td>Negative</td>
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</table>

MGACA, carcinoma arising in microglandular adenosis; NST, no special type; HER-2, human epidermal growth factor receptor 2.

Even though this histological spectrum occurred in all four cases of the invasive cancers, the invasive components of some cases had unique morphologic features to be sub-classified separately. Cases 2 and 3 had some characteristic features in the glands, which were similar to those of the pancreas or of the salivary gland. These specific glands tend to make acinar structures with abundant eosinophilic cytoplasmic granules. Case 5, the DCIS lesion, also showed acinic cell differentiation. The acinic cell differentiation in these three cases was confirmed by the immunohistochemical staining for α1-antitrypsin and chymotrypsin (Table 2, Fig. 2). Case 4 was diagnosed as metaplastic carcinoma, specifically a matrix-producing carcinoma showing an abrupt transition from the typical invasive carcinoma to spindle cell carcinoma with chondromyxoid matrix.

All tumor cells of MGACA in our cases were negative to estrogen receptor, progesterone receptor, and human epidermal growth factor receptor 2, that is, triple-negative. For the immunohistochemical study with S-100 protein, which is the most well-known immunohistochemical marker for MGA lesions, we used a polyclonal antibody which is mainly reactive to S-100b subunit. The reactivity gradually decreased as the lesions progressed from MGA to AMGA and finally to MGACA, showing near negativity in some areas of invasive carcinoma (Fig. 3). On the other hand, Ki-67 index increased from MGA to AMGA and finally to MGACA, as expected. p53 reactivity was negative in four cases except for case 4. For cytokeratin 5/6, four cases including one DCIS case demonstrated at least focal positivity (Table 2).
**DISCUSSION**

The MGA differs substantially from other adenosis lesions in many aspects including the immunohistochemical properties. The myoepithelial linings, which disappear in MGA, resulted in negative reactivity to the myoepithelial markers such as calponin and p63. In our case, calponin was all negative in the MGA lesions as well as in the associated carcinoma. The PAS and reticulin staining instead revealed the intact and regular linings of the basement membranes of MGA. These staining patterns therefore could be used to distinguish the invasive portion from the DCIS or other AMGA lesions. The invasive cancers showed the irregular and discontinuous stromal fibers around tumor cell nests.

Through the immunohistochemical studies, we could detect some phenotypic changes from MGA to MGACA. Ki-67 index increased in a stepwise manner as the diseases progressed. Especially in case 4, p53 overexpression gradually increased during transition from AMGA to MGACA. In contrast, the positivity
of S-100 protein, the most unique immunohistochemical property of MGA, diminished gradually as the disease progressed to MGACA.

However, some studies reported positive results with S-100 protein in carcinoma portions as strong as in the pure MGA lesions.\(^5\,^6\) This discordance may have resulted from the use of different members of S-100 protein, which is known to have at least 21 types currently.\(^7\) In a study of MGACA, Koenig et al.\(^3\) used two separate antibodies of S-100 protein, S-100a and S-100b for immunohistochemical characterization of MGACA, and reported that S-100b protein showed decreased reactivity, which is similar to our result. They suggested that this finding might be derived from the different localizations of the S-100a and S-100b, which are ductal cytoplasm and myoepithelium, respectively.\(^7\) However, as MGA shows immunoreactivity to S-100b, it is plausible that epithelial cells of MGA pose myoepithelial cell features and this characteristic may be lost during the progression from MGA to MGACA.

The various histologic and immunohistochemical features of MGACA in our cases have some points of interest. Among the various molecular subtypes of the triple-negative breast cancer,\(^8\) the metaplastic carcinoma case resembles mainly the mesenchymal subtype because it had chondromyxoid and spindle cell areas and was highly proliferative (Ki-67 index, 80%). Several MGA-CAs of our cases demonstrated acinic cell differentiation. Interestingly, two invasive cancers with acinic cell differentiation were all positive for \(\alpha_1\)-antitrypsin and totally negative for chymotrypsin. The DCIS case with acinic cell differentiation showed weak positivity for \(\alpha_1\)-antitrypsin. These findings are reminiscent of acinic cell carcinoma of the salivary gland rather than that of the pancreas, which is usually positive for chymotrypsin and \(\alpha_1\)-antichymotrypsin and almost always negative for \(\alpha_1\)-antitrypsin. It is also consistent with the fact that the traditional acinic cell carcinomas of the breast look similar to that of the salivary gland. Large proportions of the acinic cell carcinomas of the breast reported in articles share many aspects in common with our cases both histologically and immunohistochemically; this suggests that most acinic cell carcinomas originated in breast may arise from MGA lesions.

The molecular characteristics of MGA and MGACA are currently being investigated\(^9\,-\,^12\) and one study reported some shared genetic mutations between MGA and MGACA in four out of 12 cases.\(^12\) The overlapped sequences included recurrent gains on 1q, 2q, and 8q and losses in 14q. They suggested that MGA...
might be a non-obligate precursor to a specific subgroup of high grade triple-negative breast carcinomas.\(^\text{11}\)

As the histologic spectrum of the MGA associated lesions (from the MGA to AMGA and MGACA) becomes a frequent finding, some researchers suggested a new term, “microglandular adenoma” instead of “microglandular adenosis” for its potential to become a cancer.\(^\text{9}\) According to their opinions, MGA seems to be more similar to the adenoma of the gastrointestinal tract rather than other adenosis lesions of the breast in terms of the possibility to progress to malignancies.

In conclusion, MGACA is one of the rare breast cancers and has many unique features that are worth classifying. Further studies will be needed to find the association between the genetic backgrounds and the characteristics of the lesions.

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**Fig. 2.** Acinic cell differentiation in case 3. Tumor cells have eosinophilic granular cytoplasm (A) and show immunoreactivity to \(\alpha\)-1-antitrypsin (B).

**Fig. 3.** S-100 protein and p53 expression in case 4. (A, B) Atypical microglandular adenosis (AMGA) shows diffuse strong positivity to S-100 protein (A), while carcinoma arising in microglandular adenosis (MGACA) shows decreased expression to S-100 protein (B). (C, D) p53 staining is evident in MGACA (D), but not in AMGA (C).
Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

REFERENCES

Perivascular Epithelioid Cell Tumor in the Stomach

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In the early 1990s, Pea et al. reported that renal angiomyolipomas demonstrated positive immunoreactivity for human melanoma black (HMB)-45, which recognize the premelanosomal protein encoded by the PMEL gene. Subsequently, the same group asserted that cells of renal angiomyolipomas and clear cell tumors of the lung demonstrated similar morphology and had identical immunohistochemical profiles. Bonetti et al. proposed the term perivascular epithelioid cell to describe the aforementioned tumor cells. Zamboni et al. used the term PEComa to describe tumor containing perivascular epithelioid cells with immunoreactivity for melanocytic markers when they detected a clear cell "sugar" tumor arising in the pancreas. In 2002, the World Health Organization classified PEComa as a distinct disease entity, a mesenchymal neoplasm composed of distinctive cells that partly show an association with blood vessel walls and usually express melanocytic and smooth muscle markers.

PEComa predominantly arises in the kidney, lung, falciform ligament, ligamentum teres, uterus, or gastrointestinal tract. PEComas located in the gastrointestinal tract predominantly affect the colon and small intestine and only a limited number of cases of gastric PEComa have been reported in the English literature. Here, we present two cases of gastric PEComa encountered in our hospital.

Perivascular epithelioid cell tumors or PEComas can arise in any location in the body. However, a limited number of cases of gastric PEComa have been reported. We present two cases of gastric PEComa. The first case involved a 62-year-old woman who presented with a 4.2 cm gastric subepithelial mass in the prepyloric antrum, and the second case involved a 67-year-old man with a 5.0 cm mass slightly below the gastroesophageal junction. Microscopic examination revealed that both tumors were composed of perivascular epithelioid cells that were immunoreactive for melanocytic and smooth muscle markers. Prior to surgery, the clinical impression of both tumors was gastrointestinal stromal tumor (GIST), and the second case was erroneously diagnosed as GIST even after microscopic examination. Although gastric PEComa is a very rare neoplasm, it should be considered in the differential diagnosis of gastric submucosal lesions.

Key Words: Perivascular epithelioid cell neoplasms; Stomach neoplasms; HMB-45 protein, human; MART-1 antigen; Gastrointestinal stromal tumors

CASE REPORT

The first patient was a 62-year-old woman. She underwent a routine check-up due to a minor injury to her foot. Elevated liver enzyme levels were noted, and subsequent abdomen ultrasonography revealed a submucosal mass in the stomach. She had neither signs of tuberous sclerosis complex nor any family history of it. An abdomen computed tomography image showed a 4.2-cm subepithelial mass with broad base in the prepyloric antrum (Fig. 1A). Endoscopic ultrasound revealed a hypoechoic mass involving the proper muscle layer, and laparoscopic wedge resection of stomach was performed. Lymph node enlargement or peritoneal metastasis was not detected. The patient was discharged after the surgery without any complication.

On gross examination, the subepithelial mass measured 4.2×3.2×2.0 cm and demonstrated fleshy cut surface with a well-defined border (Fig. 1B). Microscopically, the tumor displayed a nested pattern of epithelioid tumor cells with abundant granular eosinophilic cytoplasm and round to oval nuclei. A radial arrangement of tumor cells was found around the vascular lumens. The tumor cells showed mild pleomorphism, but coagulation necrosis was not observed. The mitotic count was 1 in 50 high power fields (HPF). Lymphovascular or perineural invasion was...
not found. The tumor was focally positive for human melanoma black 45 (HMB-45), smooth muscle actin (SMA), and desmin (Fig. 1D), and negative for Melan A, vimentin, CD117, CD34, S100, pancytokeratin, and transcription factor E3 (TFE3). The Ki-67 labeling index was 1%.

The second case was detected when we evaluated the expression of melanocytic and smooth muscle markers on our 343 gastrointestinal stromal tumor (GIST) cases. The patient was a 67-year-old man who was transferred to our hospital for further evaluation of low hemoglobin levels. Endoscopic examination revealed a 5.0-cm ulcerofungating mass with spontaneous bleeding below the gastroesophageal junction, and a subtotal gastrectomy was performed. The subepithelial mass measured 5.0 × 4.7 × 1.6 cm (Fig. 2A). Microscopically, the lesion was composed of radially arranged epithelioid cells, involving mucosa to subserosa (Fig. 2B, C). The nuclei showed marked pleomorphism and the mitotic counts were high (45/50 HPF). Direct sequencing of the KIT and PDGFRA genes revealed no mutations. The tumor showed focal positivity for SMA (Fig. 2D) and faint positivity for CD117. Thus, the tumor was initially diagnosed as epithelioid GIST. However, during the re-evaluation of GIST cases with melanocytic markers, this case was positive for Melan-A as well as SMA, and therefore, the diagnosis was revised to PEComa. The patient did not receive any further treatment, and he did not show any evidence of recurrence for 7 years.

**DISCUSSION**

PEComa involving the gastrointestinal tract is rare, and the most common site within the gastrointestinal tracts is the colon. Only six cases of gastric PEComa have been reported in English literature, and two cases among them were included in the case series of gastrointestinal PEComas. Thus, detailed information regarding these tumors is insufficient (Table 1).

Microscopically, PEComa shows perivascular epithelioid cells arranged radially around the vascular lumens. Spindle cells are usually located at the periphery compared with epithelioid cells. Perivascular epithelioid cells contain a large amount of lipids, and thus, are morphologically similar to lipocyte or lipoblast. Perivascular epithelioid cells have clear to slightly eosinophilic cytoplasm compared with smooth muscle cells. The aforementioned cells demonstrated small, centrally placed, round to oval...
nuclei and small nucleoli,\textsuperscript{16} as in our cases. Upon immunohistochemical analysis, PEComas are positive for melanocytic markers such as HMB-45, Melan A, and microphthalmia-associated transcription factor. Among them, HMB-45 is considered to be the most sensitive marker.\textsuperscript{8} In our two cases, the first case was positive for HMB-45, and the second was positive

![Image](99x484 to 297x653)

![Image](298x484 to 497x653)

**Fig. 2.** Case 2. (A) The tumor was a subepithelial lesion involving gastroesophageal junction. (B, C) Microscopically, the tumor was composed of perivascular epithelioid cells with heavy lymphoid cell infiltration. (D) Immunohistochemistry reveals positivity for Melan-A (left) and smooth muscle actin (right).

**Table 1.** Summary of gastric PEComa cases

<table>
<thead>
<tr>
<th>Case</th>
<th>Reference</th>
<th>Age (yr)</th>
<th>Sex</th>
<th>Size (cm)</th>
<th>Infiltrative border</th>
<th>Nuclear grade</th>
<th>Mitotic activity</th>
<th>Necrosis</th>
<th>Cellularity</th>
<th>Recurrence or metastasis</th>
<th>IHC</th>
<th>HMB-45</th>
<th>Melan-A</th>
<th>SMA</th>
<th>Desmin</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Mitteldorf et al.\textsuperscript{12}</td>
<td>71</td>
<td>F</td>
<td>3.0</td>
<td>-</td>
<td>Moderate</td>
<td>Low (1/50 HPF)</td>
<td>-</td>
<td>NR</td>
<td>NED for 19 mo</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Waters et al.\textsuperscript{13}</td>
<td>42</td>
<td>M</td>
<td>10.0</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>Metastasis to liver</td>
<td>NR</td>
<td>+</td>
<td>NR</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Yamada et al.\textsuperscript{14}</td>
<td>39</td>
<td>M</td>
<td>7.3</td>
<td>+</td>
<td>High</td>
<td>High (&gt;2/50 HPF)</td>
<td>+</td>
<td>NR</td>
<td>NED for 6 mo</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Kumar et al.\textsuperscript{15}</td>
<td>48</td>
<td>F</td>
<td>11.5</td>
<td>NR</td>
<td>High</td>
<td>High (2/10 HPF)</td>
<td>+</td>
<td>NR</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>NR</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>Case 1</td>
<td>62</td>
<td>F</td>
<td>4.2</td>
<td>-</td>
<td>Low</td>
<td>Low (1/50 HPF)</td>
<td>-</td>
<td>Low</td>
<td>NED for 8 mo</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>Case 2</td>
<td>67</td>
<td>M</td>
<td>5.0</td>
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<td>High</td>
<td>High (45/50 HPF)</td>
<td>-</td>
<td>High</td>
<td>NED for 7 yr</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td></td>
</tr>
</tbody>
</table>

Case 2–4: malignant gastric PEComas.

IHC, immunohistochemistry; HMB-45, human melanoma black 45; SMA, smooth muscle actin; F, female; HPF, high power fields; NR, not recorded; NED, no evidence of disease; M, male.
for Melan-A. The majority of PEComas are positive for smooth muscle markers, including SMA, desmin, and calponin, and among them, SMA is considered to be the most sensitive, whilst desmin is less sensitive. Both cases in our study demonstrated positivity for SMA, and the first case showed positivity for desmin. The first case was negative for CD117 and TFE3; these markers are known to show positivity in a small proportion of PEComas.

Clinically and microscopically, PEComa can be misdiagnosed as epithelioid GIST. Therefore, we re-analyzed 343 cases diagnosed as GIST between 2005 and 2011 in our hospital, including 20 c-kit negative cases. Of the 343 cases, we found that only a single case showed positivity for Melan-A, and we believe that this case should have been classified as PEComa. Malignant melanoma or clear cell sarcoma should also be considered in the differential diagnosis; however, our second case did not show immunopositivity for S100, which is characteristic for malignant melanoma or clear cell sarcoma.

A consensus regarding the prognostic factors of PEComa has not yet to be established because of its rarity. Folpe et al. concluded that malignant behavior, including recurrence or metastasis of PEComa involving soft tissue and gynecologic organs, is associated with a large size (> 5 cm), frequent mitosis (> 1/50HPF), necrosis, infiltrative growth pattern, high cellularity, and high nuclear grade. Hornick and Fletcher ascertained that malignant behavior was associated with nuclear atypia and pleomorphism. In addition, Doyle et al. revealed that the aggressive behavior of PEComa of the gastrointestinal tract is associated with a large size (> 6 cm), mitotic count (> 2/10HPF), marked cellular atypia, and diffuse pleomorphism.

Among the previously reported four cases of gastric PEComas, three cases displayed histologically malignant features. In our first case, the tumor size was 4.2 cm and the mitotic count was 1 in 50 HPF. Microscopically, the tumor showed mild pleomorphism. Since there was no evidence of malignant features, the surgeon decided to follow-up without additional treatment. In our second case, the tumor measured 5.0 cm and the mitotic count was 45 in 50 HPF. Microscopically, the tumor showed marked pleomorphism and high cellularity, and it should have been categorized as a high risk PEComa. However, it was erroneously diagnosed as epithelioid GIST and the patient underwent curative resection only without adjuvant therapy. Fortunately, there was no adverse event during the 7 years of follow-up.

In conclusion, gastric PEComa is a rare neoplasm, but some gastric PEComas show malignant behavior. Therefore, pathologists should include PEComa in the differential diagnosis for gastric subepithelial tumors with epithelioid cell features. Since PEComas may display faint CD117 immunoreactivity, CD117 immunoreactivity alone cannot justify the exclusion of PEComa during the diagnostic process. Its tendency to manifest immunopositivity for melanocytic and smooth muscle markers should warrant pathologists to make a diagnosis with correlation of the patient’s clinical information.

Conflicts of Interest
No potential conflict of interest relevant to this article was reported.

REFERENCES

10. Im S, Yoo C, Jung HJ, Choi HJ, Yoo J, Kang CS. Primary perivascular epithelioid cell tumor in the rectum: a case report and review of


A Rare Case of Intramural Müllerian Adenosarcoma Arising from Adenomyosis of the Uterus

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Müllerian adenosarcomas are usually polyoid masses in the endometrium of post-menopausal women. Occasionally, these tumors arise in the cervix, vagina, broad and round ligaments, ovaries and rarely in extragenital sites; these cases are generally associated with endometriosis. We experienced a rare case of extraendometrial, intramural adenosarcoma arising in a patient with adenomyosis. A 40-year-old woman presented with sudden-onset suprapubic pain. The imaging findings suggested leiomyoma with cystic degeneration in the uterine fundus. An ill-defined ovoid tumor with hemorrhagic degeneration, measuring 7.5 cm in diameter, was detected. The microscopic findings showed glandular cells without atypia and a sarcomatous component with pleomorphism and high mitotic rates. There was no evidence of endometrial origin. To recognize that adenosarcoma can, although rarely, arise from adenomyosis is important to avoid overstaging and inappropriate treatment.

Key Words: Uterus; Müllerian adenosarcoma; Adenomyosis

Müllerian adenosarcoma is a relatively uncommon variant of Müllerian mixed tumor with distinctive clinical and histologic characteristics. The term Müllerian adenosarcoma was first used in 1974 to describe an unusual variant of Müllerian mixed tumor of the uterus, characterized by a mixture of benign or occasionally mildly atypical glandular components and malignant, usually low-grade, stromal components. The morphologic features of these neoplasms distinguish them from other biphasic (epithelial and mesenchymal) tumors, both benign and malignant. Müllerian adenosarcoma is generally of low malignant potential, except when accompanied by sarcomatous overgrowth and myoinvasion. Patients with Müllerian adenosarcoma lacking these two histopathologic features have an excellent prognosis; these tumors may recur locally but only rarely metastasize.

Müllerian adenosarcomas generally arise from the eutopic endometrium in the uterine corpus and grow as polyloid masses in post-menopausal women who present with abnormal vaginal bleeding. These neoplasms, however, may also arise in ectopic foci of endometriosis, such as the uterine cervix, vagina, broad and round ligaments, and ovaries or even at extragenital sites in rare cases. For example, among 41 previously reported Müllerian adenosarcomas, 29 (71%) were extrauterine, with the sites including the ovaries, cervix, peritoneum, pelvis, gastrointestinal tract, bladder, perirectal tissue, rectovaginal septum, pouch of Douglas, and liver. Some studies have reported that adenosarcomas arising in extrauterine and extragenital area may be associated with more aggressive clinical behavior, probably due to an increasing tendency for peritoneal spread.

Extraendometrial uterine adenosarcomas, that is, tumors originating from adenomyosis and not from the eutopic endometrium, are rare, with no specific staging guidelines for these tumors provided in the new International Federation of Gynecology and Obstetrics (FIGO) staging system of uterine sarcomas. In fact, only a few prior cases of primary adenosarcoma arising from adenomyosis have been reported in the English literature. The present report describes the clinical and pathologic findings of a rare case of extraendometrial, intramural adenosarcoma arising from adenomyosis without eutopic endometrial involvement in a 40-year-old woman and briefly reviews the literature describing the clinical and pathologic characteristics of adenosarcoma arising from adenomyosis.

Because this is a descriptive case report and literature review, institutional review board approval was waived and personal information of the patient was de-identified.
CASE REPORT

A 40-year-old woman (gravida 2, para 2) presented in July 2015 with sudden onset suprapubic pain and initial low back pain. Gynecologic examination detected a round soft fist-sized mass in the right fundic area accompanied by bloody vaginal discharge. Laboratory examination of serum tumor markers showed that the concentration of carbohydrate antigen (CA) 125 was markedly elevated (5,000 IU/mL; normal, <16 IU/mL), while the concentration of CA19-9 was slightly elevated (39 IU/mL; normal, <37 IU/mL). In contrast, the serum concentrations of β-human chorionic gonadotropin (1.34 mIU/mL) and α-fetoprotein (0.90 ng/mL) were within normal limits.

Pelvic ultrasonography showed an enlarged uterus with a 7 cm solid and cystic mass (Fig. 1A). T2-weighted magnetic resonance imaging (MRI) demonstrated an enlarged uterus with a mass, measuring 71 × 59 × 72 mm in size, arising from the uterine fundus (Fig. 1B). An exophytic multilocular cystic lesion, measuring 43 × 39 × 31 mm, was also present in the cranial portion of the mass. These findings suggested peripheral intramural leiomyoma with cystic degeneration and cranial rupture.

The patient underwent laparoscopically-assisted total vaginal hysterectomy without nodal dissection and without oophorectomy. Laparoscopic examination revealed a hemorrhagic friable mass arising from the uterine fundus. The tumor was attached to the bowel and omentum. There was no evidence of peritoneal endometriosis or other peritoneal lesions.

Gross examination showed that the uterus measured 14 × 7.5 × 5.5 cm in size and weighed 54 g. The uterine fundus contained an ill-defined, rubbery ovoid tumor, measuring 7.5 cm in diameter, with hemorrhagic degeneration and rupture (Fig. 2A). The remaining endometrium and cervix were unremarkable. The cut surface of the lesion was tan-brown in color, multicystic and solid (Fig. 2B).

Microscopic examination revealed a biphasic tumor composed of both dilated glandular elements and abundant, hypercellular stromal elements (Fig. 3A, B). The tumor revealed expansile growth within the myometrium with extensive myometrial invasion and focal infiltration with expansile margins into the subsersosa (Fig. 3C).

The glands were lined by benign endometrioid cells that were columnar or cuboidal in shape, with focal secretion (Fig. 3D). Some of these glands showed mild hyperplasia with rare mitosis (0–1/10 high power fields [HPFs]) of the lining; however, the glandular epithelium showed no indications of malignancy.

The benign endometrioid glands were surrounded by hypercellular spindle cell proliferation, with cells growing in a fascicular pattern. The stromal cells showed mild to moderate focal atypia with occasional mitotic figures (Fig. 3D, E). The mitotic count was 5/10 HPFs, which was higher than that of the glandular cells. Diffuse hemorrhagic necrosis was present at the center of the mass.

Immunohistochemical analysis, performed with appropriate controls, showed that both the glandular and stromal cells were positive for estrogen and progesterone receptors (Fig. 3F) and focally positive for TP53 in the tumor tissue (Fig. 3G). The stromal cells were immunohistochemically positive for CD10 (Fig. 3H) and smooth muscle actin (Fig. 3I), and their Ki-67 proliferation index was mildly increased (Fig. 3J).

These microscopic findings suggested that the tumor was a mixed epithelial-stromal tumor consisting of glandular cells without atypia (benign endometrial hyperplasia) and sarcomatous stromal cells with mild pleomorphism and a high mitotic rate.

Fig. 1. Radiologic findings. (A) Pelvic ultrasonography, showing an enlarged uterus with a 7-cm solid and cystic mass. (B) T2-weighted magnetic resonance imaging, showing an enlarged uterus with a mass, measuring 71 × 59 × 72 mm, arising from the uterine fundus.
Taken together, these histologic findings indicated a low-grade Müllerian adenosarcoma arising from adenomyosis. The patient underwent follow-up evaluation 2 weeks after surgery. Further clinical workup included a whole body bone scan and positron emission tomography/computed tomography (PET-CT) scan. There was no evidence of regional or distant metastasis. One month later, the patient underwent an additional bilateral salpingooopherectomy, omental biopsy, and cul de sac washing cytology. There was no residual adenosarcoma in the specimen. To date, regular follow-up evaluation has shown no evidence (low-grade endometrial stromal sarcoma). There was no evidence of sarcomatous overgrowth, a heterologous (i.e., muscle, cartilage, bone) or sex-cord element, or lymphovascular invasion.

The endometrium was unremarkable and the background myometrium showed focal involvement of adenomyosis. There was no evidence of serosal endometriosis or other tumors intrinsic or extrinsic to the uterus. Although the specimen was submitted in a ruptured state, there was no distinct evidence of serosal involvement of the tumor. The adenosarcoma was limited to the myometrium and was not detected on the serosal surface.

**Fig. 2.** Macroscopic findings of the hysterectomy specimen. (A) View showing an ill-defined ovoid tumor, 7.5 cm in diameter, together with hemorrhagic degeneration in the uterine fundus. (B) View showing that the cut surface of the lesion was tan-brown in color, multicystic, and solid.

**Fig. 3.** Histologic and immunohistochemical findings. (A, B) Microscopic examination showing a biphasic tumor composed of both dilated glandular elements and abundant, hypercellular stromal elements. (C) The tumor shows expansile growth within the myometrium, with extensive myometrial invasion and focal infiltrates into the subserosa with expansile margins. (D) Proliferation of hypercellular spindle cells growing in a fascicular pattern, around benign endometrial glands. (E) The stromal cells show mild and focal moderate cytological atypia with occasional mitotic figures (arrows). (F–J) Immunohistochemical analysis showing that the glandular and stomal cells in tumor tissue are positive for estrogen receptor (F) and focally positive for p53 (G); that the stromal cells are positive for CD10 (H) and smooth muscle actin (I); and that the Ki-67 proliferation index is higher in the stromal component than in the epithelium (J).
of tumor recurrence, with negative radiologic findings, including transvaginal ultrasonography, chest and pelvic computed tomography, and whole body PET-CT. Her elevated serum CA125 level resolved immediately after surgery and has continued to remain in the normal range.

DISCUSSION

Uterine sarcomas of any type are rare when compared with uterine epithelial malignancies. Especially, Müllerian adenosarcomas are highly rare, accounting for only 3%–7% of all uterine sarcomas.1,12

Despite their rarity, these tumors affect women of all age groups.1 Although most adenosarcomas of the female genital tract occur in post-menopausal women (median age, 58 years), these tumors have been reported in perimenopausal women as well as in children as young as 10 years old.12 Clinically, the patients usually present with vaginal bleeding, but may also present with other nonspecific symptoms, such as pelvic pain and vaginal discharge; a large percentage of patients are asymptomatic.1,2

Although the eutopic endometrium of the uterus is the most common primary site, adenosarcomas may arise in the ovaries, fallopian tubes, cervix, or vagina, as well as at extragenital sites in rare cases.3 Adenosarcomas occurring outside the female genital tract likely represent tumors arising from preexisting endometriosis.3 In fact, approximately 1%–10% of these tumors are thought to arise from the malignant transformation of endometriosis.4 Endometrioid adenocarcinomas and clear cell carcinomas are the most common tumor types, occurring in 70% and 14% of patients, respectively.4 Only a limited number of stromal tumors including adenosarcoma have been reported to date.4

Furthermore, these tumors are thought to arise from the endometrial stroma, they can implant into the myometrium and coexist with diffuse adenomyosis or focal adenomyoma; however, only a few cases of intramural adenosarcoma arising from adenomyosis have previously been reported in the English literature.2,3,8,10,13-15 These include intramural adenosarcomas originating in the foci of adenomyosis,4 intramural adenomyoma,8 and deep subserosal adenomyoma2 with no evidence of eutopic endometrial origin. The detailed clinicopathologic characteristics of previously described patients are summarized in Table 1.

An essential feature of these adenosarcomas is their mixed nature, including both benign and neoplastic epithelial and malignant stromal elements.3 The epithelium, which may be dilated or slit-like in appearance, is mostly endometrioid type, usually composed of cuboidal or low columnar cells, although ciliated, mucinous, and occasionally squamous type epithelial cells have also been identified.1 Abundant eosinophilic cytoplasm may also be present in the epithelial cells.1 The epithelial cells are usually in active phase, showing mitoses or subnuclear vacuolation, in spite of the adjacent atrophic endometrium.3 Focal crowded glands with nuclear atypia may occur, which are indicators of atypical hyperplasia.3

The stromal component, which is usually low grade, is composed of spindle and/or round cells.1 In a minority of patients, including ours, the stromal component is uniformly cellular.7 The formation of a “periglandular cuffing” or “cambium” layer, in which the stroma surrounds the glandular elements, is one of the most characteristic features of adenosarcoma.1 This cellular zone is characterized by variable degrees of nuclear atypia and mitosis.3 Although most of these tumors have a mitotic count of ≥ 2/10 HPFs, some tumors have a lower count.3 The World Health Organization criteria consider stromal mitotic activity > 1/10 HPFs diagnostic of adenocarcinoma, whereas other classification systems use a cutoff of 4/10 HPFs.3 Moreover, many adenosarcomas include large areas of stromal fibrosis without mitotic activity.3 Stromal components of adenosarcomas are exclusively homologous, that are normally seen in the uterus.3 The most common mesenchymal elements consist of low-grade endometrial stromal or nonspecific fibroblastic components; the stromal cells in the periglandular area are often endometrial stromal type whereas that located away from the glands are typically of nonspecific fibrous type.3 Approximately one quarter of adenosarcomas show heterologous stromal components, comprising predominantly rhabdomyosarcoma; however, chondrosarcoma and liposarcoma may also be seen.16 Sex cord-like components, such as cords, solid nests, and/or hollow tubules, may be found within the stromal component.3 Occasionally, marked decidualization of the stroma may also be present, secondary to hormonal usage.3

Adenosarcoma is one of the mixed Müllerian tumors of the female genital tract, located between adenofibroma and carcinosarcoma (so called malignant mixed Müllerian tumor) on the spectrum.3 Because these tumors are rare, the precise molecular mechanism associated with their tumorigenesis is not fully understood.3 Some studies have reported low rates of TP53 mutations, and low-level amplification of MDM2 and other genes on chromosome band 12q14-15.17 Recently described alterations in adenosarcoma include MYBL1 amplification and ATRX mutation, each of which has been detected in 50% of adenosarcomas with sarcomatous overgrowth.17 Immunohistochemical analysis of the tumor in our patient showed an overexpression of TP53 in both epithelial and stromal cells. In a previous study, an adenosarcoma with sarcomatous overgrowth was found to have a hyperdiploid karyotype with multiple structural and numerical abnormalities involving
Table 1. Clinicopathologic features of adenosarcomas arising from adenomyosis

<table>
<thead>
<tr>
<th>Case No. (ref No.)</th>
<th>Clinical feature</th>
<th>Pathology</th>
<th>Treatment</th>
<th>Outcome</th>
<th>Remarks</th>
</tr>
</thead>
</table>
| 1\(^{15}\) Age: 51 yr  
Gyn hcx: unknown  
Clinical sign: unknown | Size: 4 cm  
Location: lateral wall of the uterine body  
Micro:  
Glands with no epithelial cell atypia  
Sarcomatous component with cell pleomorphism  
and a high mitotic count  
Accompanied by adenomyosis | Unknown | Unknown | - |
| 2\(^{23}\) Age: 20 yr  
Gyn hcx: null  
Clinical sign: a long-standing history of menorrhagia and vaginal bleeding | Size: unknown  
Location: right anterolateral portion  
Micro:  
Florid adenomyosis with extensive myometrial invasion, expansive growth within the myometrium, and intravascular invasion in the myometrium | Hysterectomy | Two years after surgery, no evidence of recurrent disease | Stromal overgrowth |
| 3\(^{3}\) Age: 46 yr  
Gyn hcx: para 1  
Clinical sign: vaginal bleeding | Size: unknown  
Location: subserosal mass arising from the posterior surface of the uterus  
Micro:  
Adenomyoma with focal predominant endometrial stroma and periglandular cuffs  
Endometrial stromal cells in the periglandular cuffs showing mild and focal moderate cytological atypia with sparse mitotic figures, including an occasional atypical form | Myomectomy | Unknown | - |
| 4\(^{4}\) Age: 38 yr  
Gyn hcx: gravida 1, para 0  
Clinical sign: chronic pelvic pain and dysmenorrhea | Size: 1.5 cm  
Location: right cornual area  
Micro:  
Irregular glands with benign epithelium surrounded by a hypercellular spindle cell stroma showing rare mitoses, mild nuclear hyperchromasia, and pleomorphism | Exploratory laparotomy, TAH, LSO, and omentectomy | Disease-free 30 mo after treatment | Heterologous element (rhabdomyosarcoma) |
| 5\(^{10}\) Age: 52 yr  
Gyn hcx: gravida 3, para 3  
Peri-menopausal | Size: uncheckable (no distinct mass formation)  
Location: uterine fundus  
Micro:  
Diffuse adenomyosis with focal stromal expansion, consisting of a hypercellular proliferation of moderately atypical spindle cells with mitotic activity around benign endometrial glands and infiltrating the anterior myometrium | Radical hysterectomy with BSO and lymph node dissection and debulking of the pelvic mass | Unknown | Extraterine pelvic mass (19 cm in diameter) diagnosed as adenosarcoma with rhabdomyosarcomatous differentiation and stromal overgrowth |

(Continued to the next page)
chromosomes 2, 8, 10, 13, 19, and 21. Finally, studies have also suggested that the use of tamoxifen may have a role in the pathogenesis of adenosarcoma.

Several types of uterine tumors, including adenofibroma, adenosarcoma, and carcinosarcoma, consist of mixtures of epithelial and stromal components. Because they include large hypocellular areas and infrequent mitoses, adenosarcomas are difficult to histologically differentiate from adenofibromas, endometriosis and adenomyosis. Especially on a background of adenomyosis, a conclusive pathologic differential diagnosis may be more difficult, because florid adenomyosis can prevent an accurate initial diagnosis. In typical adenomyosis, the volume of the endometrial stromal component is low relative to the volume of smooth muscle. Moreover, although endometrial stromal cells have mitotic activity, they show no evidence of significant cytological atypia. Similarly, stromal mitoses of ≥2 per 10 HPFs, marked stromal cellularity, and significant stromal cell atypia can differentiate Müllerian adenosarcoma from Müllerian adenofibroma. In one study, the diagnoses of eight adenomyomas were revised as adenofibroma, atypical polypoid adenomyoma, adenocarcinoma, and adenosarcoma. The adenofibroma contains variably cellular fibrous tissue in stroma, but lacks the smooth muscle and endometrial stromal tissue present in adenomyoma. In adenosarcoma, the endometrial stromal tissue predominates, not the smooth muscle component, and the glands are no longer evenly distributed but are more widely

<table>
<thead>
<tr>
<th>Table 2. The 2009 FIGO staging system for uterine adenosarcoma</th>
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FIGO, International Federation of Gynecology and Obstetrics.
The lack of a staging system for uterine adenosarcoma led to development of the FIGO staging system for endometrial carcinomas. The new FIGO staging system for uterine adenosarcoma, published in 2009, is identical to that for endometrial stromal sarcoma (Table 2) and is an improvement on the earlier generic application of the 1988 FIGO staging system for endometrial cancer to adenosarcoma.

Because of tumor involvement of the outer half of the myometrium, the tumor in our case would be staged as IC (deeply myoinvasive) according to the new FIGO staging system for uterine sarcomas. Stage IC tumors would normally be considered for adjuvant treatment. However, the FIGO staging applies to adenosarcomas that arise in the endometrium and invade into the myometrium. Thus, use of a staging system that does not consider adenosarcomas that arise ab initio within the myometrium, and are confined to this layer, would have resulted in overstaging of the adenosarcoma in our patient.

To date, there are no specific staging guidelines for tumors arising from adenomyosis. In a series of case reports, two previous cases of adenosarcomas were reported to be problematic in staging. The first was a tumor that arose in the eutopic endometrium with involvement of underlying adenomyosis but without myometrial invasion. The second was a tumor located in an intramural adenomyoma with no evidences of a primary lesion in the eutopic endometrium. It was suggested that these tumors should be noted as adenosarcoma, intramural stage I am1 or am2, whether the involved myometrium. Stage IC tumors would normally be considered for adjuvant treatment. However, the FIGO staging applies to adenosarcomas that arise in the endometrium and invade into the myometrium. Thus, use of a staging system that does not consider adenosarcomas that arise ab initio within the myometrium, and are confined to this layer, would have resulted in overstaging of the adenosarcoma in our patient.

In conclusion, this report describes the clinical and pathologic findings of a rare extraendometrial, intramural adenosarcoma arising from adenomyosis without eutopic endometrial involvement in a 40-year-old woman. Malignant transformation of adenomyosis should be considered in the differential diagnosis of a huge solid and cystic uterine mass. To avoid overstaging and inappropriate treatment, it is important to recognize that adenosarcomas can, although rarely, arise from adenomyosis. Collection of additional data and prospective studies will provide a clearer understanding of the malignant transformation of adenomyosis and may provide more evidence for an upgraded staging system.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.
Acknowledgments

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REFERENCES

Inhibition of mutated epidermal growth factor receptor (EGFR), EGFR tyrosine kinase inhibitor (TKI), is one of the most successful cancer targeted therapies.\(^1\) While this therapy has been beneficial for many non-small cell lung cancer (NSCLC) patients with activated EGFR mutations, almost all patients inevitably develop acquired resistance which limits the median response duration to around 1 year.\(^2\) Most of the mechanisms leading to EGFR TKI resistance are additional mutation or amplification of alternative pathways, and morphological transformation to small cell lung cancer (SCLC).\(^3\)

Recently, several authors reported another type of morphologic transformation for EGFR TKI resistance, namely, transformation from adenocarcinoma to squamous cell carcinoma.\(^4-6\) However, the reports were of limited value because they were based only on biopsied specimens. This report describes a case of lung adenocarcinoma with transformation to squamous cell carcinoma that was established histological evaluation of lobectomy specimen.

### CASE REPORT

A 40-year-old man with a 23-year smoking history received a medical check-up, which revealed a solitary pulmonary nodule. Chest computed tomography revealed a 17-mm-sized tumor at the peripheral posterior segment of the right upper lobe (clinically T1aN0M0). The patient underwent a right upper lobectomy with mediastinal lymph node dissection. Histological examination showed a 1.3-cm-sized poorly differentiated adenocarcinoma with solid and acinar growth patterns with visceral pleural invasion and no lymph node metastasis (pT2aN0). The tumor cells showed marked nuclear atypia, numerous mitoses (23/10 high power field [HPF]) and multifocal microscopic necrosis. The tumor was dissected and entirely embedded. We morphologically and immunohistochemically evaluated all tumor sections to rule out the possibility of adenosquamous carcinoma (Fig. 1A–E). The patient received adjuvant chemotherapy with combined vinorelbine and cisplatin.

Four years later, multiple metastatic lesions were found at the right lower lobe, right pleura, right fifth rib, and right femoral head. EGFR mutation analysis, using a previously resected lobectomy specimen, revealed a deletion at exon 19 of the EGFR gene. The patient was treated with afatinib, which is an EGFR TKI and radiation therapy. The patient showed partial remission of the tumor.

Two years later, the patient developed multiple metastatic lesions throughout the body, including a telangiectatic nodule on the scalp. A punch biopsy was performed and histologic evaluation of the biopsied specimen showed a tumorous lesion in the deep dermis. This tumor showed squamoid appearance with sheet-like growth patterns and evident intercellular bridges. Compared to the previous lobectomy specimen, the tumor cells were more bland with moderate nuclear atypia, some mitotic activity (13/10 HPF) and no necrosis. Immunohistochemical staining showed diffuse and strong p63 immunoreactivity and no thyroid transcription factor 1 immunoreactivity (Fig. 1F–J).
EGFR mutation analysis, using a biopsied specimen, revealed a deletion at exon 19 of the EGFR gene and an additional T790M point mutation. After several special and immunohistochemical stainings, this tumor was diagnosed as a metastatic carcinoma from the lung with squamous differentiation. The patient informed consent was waived by the Institutional Review Board of the Samsung Medical Center (2016-08-109).

DISCUSSION

Lung cancer is the most common cause of cancer deaths in Korea. Traditional therapy, including resection, platinum-based chemotherapy and radiation therapy, have only limited therapeutic value. Therefore, the 5-year survival rate of lung cancer has not changed significantly in the past 30 years.

EGFR TKI therapy, which specifically targets EGFR, was recently introduced and provided guidance in this situation. Targeting EGFR in patients with activating EGFR mutations has shown initial and significant success in practice. Unfortunately, the vast majority of patients develop resistance to the treatment, typically in less than 1 year. In this situation, understanding the mechanism of the resistance became very important.

Most of the mechanisms that lead to EGFR TKI resistance involve an additional mutation, such as a T790M mutation, or amplification of alternative pathways. In addition, morphological transformation is also a well-known mechanism. The most well known example of this interesting phenomenon is the transformation from NSCLC to SCLC.

Recently, several authors have described histologic transformation from lung adenocarcinoma to squamous cell carcinoma as a mechanism of resistance to EGFR TKI therapy. In four previously reported cases, all four patients were female and three of them had never been smokers. The most common EGFR mutation was an L858R point mutation and the most common acquired gene alteration was a T790M point mutation. All cases showed the same EGFR mutation in both primary
and metastatic carcinomas.

The mechanism of this morphologic transformation is still poorly understood. Possible explanations include (1) small population of squamous cell carcinoma phenotype cells are already present before the EGFR TKI therapy and selectively survive during EGFR TKI therapy; or (2) carcinoma cells acquire a different morphologic phenotype during EGFR TKI therapy.

Considering that primary adenosquamous carcinoma shows the same mutations in both adenocarcinoma and squamous cell carcinoma components, additional studies are needed and a solution is beyond the scope of this report.

In conclusion, lung primary adenocarcinoma can transform into squamous cell carcinoma after EGFR TKI therapy. We should be aware of this phenomenon to avoid misdiagnosis in practice.

Conflicts of Interest
No potential conflict of interest relevant to this article was reported.

REFERENCES


Bronchial Washing Cytology of Pulmonary Langerhans Cell Histiocytosis: A Case Report

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Pulmonary Langerhans cell histiocytosis (PLCH) is a rare histiocytic interstitial lung disease, characterized by abnormal infiltration of Langerhans cells (LCs) with lung parenchyma destruction. PLCH can occur at any age, but is reported mainly in adults (between 20 and 40 years), especially those with a history of cigarette smoking. The clinical presentation can vary from no symptoms to severe respiratory symptoms.1 PLCH is regarded as a reactive lesion rather than a neoplastic process, even though the proliferation of LCs is found in the lesion.2

PLCH is normally diagnosed from resected lung specimens. Cytological diagnostic methods such as bronchial washing or bronchoalveolar lavage (BAL) are used as a less invasive technique through bronchoscopy.3 Reports have indicated that BAL cytology can be helpful for diagnosing PLCH.4 However, the diagnosis of PLCH by bronchial washing cytology has not yet been reported. Herein, we present a case of PLCH diagnosed through bronchial washing cytology.

CASE REPORT

The 41-year-old male patient visited our hospital due to a cough that had persisted for 7 months. He had a history of 20 pack-year smoking and diabetes mellitus. Physical examination of the chest revealed fine crackle of both upper lung fields. There was no skin rash or palpable lymph node. The chest computed tomography (CT) showed multiple, irregularly-shaped cysts and centrilobular nodules of variable sizes in both lungs (Fig. 1).

Bronchial washing cytology, BAL, and wedge resection of the lung were performed. The BAL slides revealed a fraction rate of 59.6% monocytes and 9.4% eosinophils with about 6.5% LCs. The bronchial washing slides showed scattered non-cohesive large cells with abundant, granular, and eosinophilic cytoplasm. These cells had grooved or convoluted nuclei with fine chromatin, delicate nuclear membranes, and prominent nucleoli. Immunohistochemical staining for CD1a was positive in these cells (Fig. 2).

The cut surface of the wedge-resected lung specimen revealed numerous cystic spaces with whitish gray stellate fibrous scars. Microscopic examination indicated multiple cystic spaces of variable sizes with diffuse alveolar wall thickening, cellular infiltration, and fibrosis. The infiltrated cells had eosinophilic cytoplasm and grooved or infolded nuclei. These cells were proven to be LCs based on immunohistochemical staining, showing strong positive reaction for CD1a and S-100 protein (Fig. 3).

Bronchial washing cytology was performed again during follow-up. Some cells with the features described above were detected on the bronchial washing cytology. Therefore, we used the same approach to determine that the PLCH was still present.

DISCUSSION

When patients with ongoing PLCH present at a hospital, non-or less-invasive pulmonological approaches, such as chest CT, pulmonary function test, and fiberoptic bronchoscopy, are usually performed. The cytological specimens obtained from fiberoptic bronchoscopy are then sent to the pathology department, and these cytological specimens can be challenging for pathologists to diagnose. The cytological diagnosis of PLCH with bronchial washing can be difficult because it is a rare disease with unfamiliar

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features. Therefore, in this report, we focused on bronchial washing cytology findings of PLCH.

Some reports have presented cytological findings of Langerhans cell histiocytosis (LCH) that have arisen in various organs. Lee et al. reported a case of LCH in the mandible diagnosed with fine needle aspiration cytology and indicated that the smear revealed high cellularity, and that the LCs had abundant vacuolated cytoplasm with round or occasionally folded nuclei. The LC nuclei showed a fine granular chromatin pattern with prominent nucleoli. Ha et al. reported a case of LCH in lymph nodes using aspiration cytology similar to the case reported by Lee et al. In their study, the LCs had wrinkled or grooved nuclei with thin nuclear membranes and a delicate chromatin pattern. Multinucleated cells were also identified. In another report that applied fine needle aspiration LCH cytology, many of the cytological findings were similar, but the nucleoli were absent.

Another cytological method applied fiberoptic bronchoscopy for diagnosing PLCH is BAL fluid analysis. Takizawa et al. conducted BAL fluid analysis in PLCH and found a high mean cell fraction rate for macrophages (75.2%) and a variable eosinophil fraction rate (1% to 16%). The LCs revealed chromatin-poor nuclei with well-developed nucleoli. Previously described cytological findings with quantitative analysis of PLCH in BAL fluid are summarized in Table 1. In two studies with CD1a-positive cells in PLCH, BAL fluid obtained from patients with PLCH revealed more than 5% CD1a-positive cells. In the present case, BAL

Fig. 1. Chest computed tomography findings. There are multiple, variable-sized, irregularly-shaped cysts and centrilobular nodules in both lungs.

Fig. 2. Bronchial washing cytology. Bronchial washing cytology reveals many Langerhans cells. (A–C) The cells have abundant pale granular cytoplasm with a convoluted irregular nucleus and fine chromatin pattern (A, cell block; B, Thin prep; C, conventional smear). (D) Immunohistochemical staining for CD1a is positive in the Langerhans cells.
fluid cytology showed similar fraction rates and cytological features. The percentage of LCs was about 6.5%. Based on these results, the 5% cut-off value of CD1a-positive cells or LCs could be postulated in BAL fluid cytology to make a PLCH diagnosis.

In this report, we described bronchial washing cytology findings of PLCH. These findings are similar to other studies describing aspiration cytology and BAL. A common finding in all of the previous reports indicates that grooved or infolded nuclei with abundant cytoplasm could be used as diagnostic features for LCs.

The difference in the present case compared with other reports could be that prominent nucleoli were observed only in some cells. The Birbeck granules are known as diagnostic LCH structures on electron microscopy (EM). However, EM study requires additional time and costs. We did not perform EM to detect Birbeck granules since other authors have suggested that it is not essential for diagnosing LCH.

In conclusion, the possibility of LCH can be considered when bronchial washing cytology is mainly composed of Langerhans cells and eosinophils. In addition, more than 5% CD1a-positive cells in BAL fluid could be helpful for diagnosing PLCH. However, to date, there is no quantitative guideline for diagnosing PLCH. Therefore, additional cytological studies of PLCH, including quantitative analysis, are required to further identify and improve diagnostic approaches.

Conflicts of Interest
No potential conflict of interest relevant to this article was reported.

REFERENCES