

A multicenter study of interobserver variability in pathologic diagnosis of papillary breast lesions on core needle biopsy with WHO classification

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Background: Papillary breast lesions (PBLs) comprise diverse entities from benign and atypical lesions to malignant tumors. Although PBLs are characterized by a papillary growth pattern, it is challenging to achieve high diagnostic accuracy and reproducibility. Thus, we investigated the diagnostic reproducibility of PBLs in core needle biopsy (CNB) specimens with World Health Organization (WHO) classification. **Methods:** Diagnostic reproducibility was assessed using interobserver variability (kappa value, κ) and agreement rate in the pathologic diagnosis of 60 PBL cases on CNB among 20 breast pathologists affiliated with 20 medical institutions in Korea. This analysis was performed using hematoxylin and eosin (H&E) staining and immunohistochemical (IHC) staining for cytokeratin 5 (CK5) and p63. The pathologic diagnosis of PBLs was based on WHO classification, which was used to establish simple classifications (4-tier, 3-tier, and 2-tier). **Results:** On WHO classification, H&E staining exhibited 'fair agreement' (κ =0.21) with a 47.0% agreement rate. Simple classifications presented improvement in interobserver variability and agreement rate. IHC staining increased the kappa value and agreement rate in all the classifications. Despite IHC staining, the encapsulated/solid papillary carcinoma (EPC/SPC) subgroup (κ =0.16) exhibited lower agreement compared to the non-EPC/SPC subgroup (κ =0.35) with WHO classification, which was similar to the results of any other classification systems. **Conclusions:** Although the use of IHC staining for CK5 and p63 increased the diagnostic agreement of PBLs in CNB specimens. **Conclusions:** Although the use of IHC staining for CK5 and p63 increased the diagnostic agreement of PBLs in CNB specimens, WHO classification exhibited a higher discordance rate compared to any other classifications. Therefore, this result warrants further intensive consensus studies to improve the diagnostic reproducibility of PBLs with WHO classification.

Key Words: Papillary breast lesion; Core needle biopsy; Interobserver variability; Agreement rate

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380

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Papillary breast lesions (PBLs) encompass a broad spectrum of proliferative diseases that account for less than 3% of breast tumors [1-3]. The histologic features of PBLs include mass-like projections attached to the wall of the dilated ducts and have a fibrovascular stalk lined by epithelial cells. PBLs can be both benign and malignant lesions, representing less than 10% of benign breast lesions and less than 2% of all breast cancers [4,5], respectively.

Ultrasound-guided core needle biopsy (CNB) is universally used in the initial pathologic approach for suspicious radiologic findings in breast lesions. PBLs constitute approximately 4.5% to 10.7% of breast lesions diagnosed on CNB [6,7]. The 5th edition of the WHO classification of tumors of the breast is the most recently updated version for pathologically diagnosing PBLs [8]. Compared with the 4th edition of the WHO classification of tumors of the breast there have been little or no changes since 2012 in terms of the diagnostic criteria and classification of PBLs [8,9]. However, differential diagnosis of PBLs remains challenging due to the limited samples obtained from CNB. The difficulty in pathologic diagnosis of PBLs increases due to the broad spectrum of histological findings and subtle differences exemplifying each category [1-3]. Moreover, the lack of reliable and reproducible criteria of its diagnosis and classification may limit diagnostic accuracy [10].

Several studies have sought to promote the interpretation reproducibility of PBLs among pathologists as an endeavor to improve diagnostic accuracy. Immunohistochemical (IHC) staining has significantly increased diagnostic agreement rates among pathologists who have exhibited unsatisfactory findings on hematoxylin and eosin (H&E) staining [11-13]. These studies suggest that additional histopathologic modalities are potentially useful in increasing the diagnostic agreement rate. Nonetheless, these studies have limitations in generalization because their results were derived from analysis among very few pathologists from a single institution [11-13]. Moreover, there is little data about the agreement rate of PBLs based on the WHO classification.

To evaluate the diagnostic reproducibility of PBLs on CNB based on the WHO classification, we investigated the interobserver variability among 20 breast pathologists working in 20 medical institutions. We intended to compare the interobserver variability between H&E and IHC stains and specify the diagnostic pitfalls in the differential diagnosis of challenging cases.

MATERIALS AND METHODS

Study design and case selection

We evaluated the interobserver variability and agreement rates in 60 PBL cases on CNB among 20 breast pathologists affiliated with 20 medical institutions in Korea. Sixty PBL cases were recruited from 20 medical institutions that participated in this study. The consensus meeting of the Korean Breast Pathology Study Group (KBPSG) verified and determined the pathologic diagnosis of 60 PBL cases on CNB. Fig. 1 displays the composition of pathologic diagnoses in all 60 PBL cases. Each case constitutes one H&E and two IHC stained slides for both cytokeratin 5 (CK5) and p63. Initially, 60 H&E-stained slides were circulated to 20 breast pathologists for review. Subsequently, IHC stained slides for CK5 and p63 in the same 60 cases were circulated to the same 20 breast pathologists and re-reviewed. Interobserver variability and agreement rates were analyzed for the pathologic diagnosis of PBLs in H&E and IHC stains. Additionally, we conducted a detailed review of the challenging cases of differential diagnoses observed among our 60 PBL cases.

Diagnostic classification of PBLs

Pathologic classification of PBLs was conducted based on the 4th edition of the *WHO classification of tumors of the breast* [9]. In this classification [9], the PBLs were classified into 10 categories



Fig. 1. Composition of the pathologic diagnosis in all 60 papillary breast lesions. SPC, solid papillary carcinoma; IDP, intraductal papilloma; ADH, atypical ductal hyperplasia; DCIS, ductal carcinoma in situ; PCIS, papillary carcinomas in situ; EPC, encapsulated papillary carcinoma; SPC, solid papillary carcinoma.

comprising intraductal papillomas (IDP), IDPs with atypical ductal hyperplasia (ADH), IDPs with ductal carcinomas in situ (DCIS), IDPs with lobular carcinomas in situ (LCIS), papillary carcinomas in situ (PCIS), encapsulated papillary carcinomas (EPC), solid papillary carcinomas (SPC), EPCs with invasion, SPCs with invasion, and invasive papillary carcinomas (IPC). Of the WHO classification, intraductal papillary neoplasms (IDPN) were defined as a category including IDP, IDP with ADH, IDP with DCIS, IDP with LCIS, and PCIS. EPC and SPC were categorized into EPC/SPC.

In addition, using the WHO classification, we created simple classifications of PBL using 4-tier, 3-tier, and 2-tier systems as follows: 4-tier consisted of benign, atypical, in situ, and invasive; 3-tier consisted of benign, in situ, and invasive; 2-tier consisted of benign and malignant (Table 1). For instance, if EPC was diagnosed, it was categorized into in situ in the 4-tier system and malignant in the 2-tier system.

Immunohistochemistry

For each CNB specimen of cases, IHC staining for CK5 and p63 was performed. IHC staining for CK5 was conducted using antibodies against CK5 (XM26, Leica Biosystems, Newcastle upon Tyne, UK) with 1:200 antibody dilution and a detection kit (Ivew DAB kit, Ventana, Tucson, AZ, USA). IHC staining for p63 was performed using antibodies against p63 (BC4A4, Biocare Medical, Pacheco, CA, USA) with 1:100 antibody dilution and a detection kit (Ultraview DAB kit, Ventana). According to the manufacturer's protocol, all the procedures of IHC staining were processed by a Ventana BenchMark XT system (Ventana).

 Table 1. Diagnostic categories for PBL in the WHO, 4-tier, 3-tier, and 2-tier classifications

WHO	4-tier	3-tier	2-tier
Intraductal papilloma	Benign	Benign	Benign
IDP with ADH	Atypical		
IDP with DCIS	In situ	In situ	Malignant
IDP with LCIS			
Papillary carcinoma in situ			
Encapsulated papillary carci-			
noma			
Solid papillary carcinoma			
EPC with invasion	Invasive	Invasive	
SPC with invasion			
Invasive papillary carcinoma			

PBL, papillary breast lesion; WHO, World Health Organization; IDP, intraductal papilloma; ADH, atypical ductal hyperplasia; DCIS, ductal carcinoma in situ; LCIS, lobular carcinoma in situ; EPC, encapsulated papillary carcinoma; SPC, solid papillary carcinoma.

Statistical analysis

Fleiss's kappa values for interobserver variability were used in analyzing diagnostic reproducibility in H&E and IHC staining among 20 breast pathologists. Interobserver variability was classified into five categories (0.00–0.20, poor; 0.21–0.40, fair; 0.41– 0.60, moderate; 0.61–0.80, substantial; and 0.81–1.00, excellent agreement) to identify the level of reproducibility. Additionally, the average of the agreement rates in H&E and IHC staining of 60 cases was calculated in four diagnostic classifications. The agreement rate was determined by the proportion of pathologic diagnosis from 20 pathologists that was consistent with that from the consensus meeting of KBPSG. Statistical analyses were performed using STATA ver. 16.0 (StataCorp LP, College Station, TX, USA).

RESULTS

Interobserver variability and agreement rates in H&E and IHC staining in each classification are presented in Table 2. In the WHO classification, H&E staining exhibited 'fair agreement' (κ = 0.21). Kappa values increased inversely with the number of categories in the diagnostic classification (4-tier: κ =0.31, 3-tier: κ =0.42, and 2-tier: κ =0.44). IHC staining improved the interobserver variability in all classifications. In IHC staining, overt improvement in reproducibility was observed in 4-tier ('fair agreement' to 'moderate agreement') and 2-tier ('moderate agreement' to 'substantial agreement'). The agreement rate also exhibited similar findings with kappa values for interobserver variability. The agreement rate was generally higher in IHC staining compared to H&E staining in all classifications. Within the same staining methods, simpler diagnostic classification tended to have a higher agreement rate.

Fig. 2 shows the interobserver variability in H&E staining for all 60 PBL cases, IDPN, and EPC/SPC in each classification. There were 48 cases of IDPN and 12 cases of EPC/SPC in this

Table 2. The interobserver variability and agreement rates in H&E and IHC staining of 60 PBL cases in each diagnostic classification

Diagnostic classification	H&E staining		IHC staining	
	к	Agreement rate (%)	к	Agreement rate (%)
WHO	0.21	47.0	0.37	60.5
4-tier	0.31	63.3	0.51	76.4
3-tier	0.42	76.7	0.56	84.3
2-tier	0.44	80.0	0.62	87.5

H&E, hematoxylin and eosin; IHC, immunohistochemistry; PBL, papillary breast lesion; WHO, World Health Organization classification; 4-tier, 4-tier classification; 3-tier, 3-tier classification; 2-tier, 2-tier classification.



Fig. 2. Interobserver variability in the pathologic diagnoses of all 60 papillary breast lesions cases, intraductal papillary neoplasms (IDPN), and encapsulated/solid papillary carcinoma (EPC/SPC) with hematoxylin and eosin staining in all classifications. WHO, World Health Organization classification; 4-tier, 4-tier classification; 3-tier, 3-tier classification; 2-tier, 2-tier classification.



Fig. 3. Interobserver variability in the pathologic diagnoses of all 60 papillary breast lesions cases, intraductal papillary neoplasms (IDPN), and encapsulated/solid papillary carcinoma (EPC/SPC) with immunohistochemical staining in all classifications. WHO, World Health Organization classification; 4-tier, 4-tier classification; 3-tier, 3-tier classification; 2-tier, 2-tier classification.

study. Kappa values for IDPN and EPC/SPC were lower than that for all 60 PBL cases in all classifications. IDPN had lower reproducibility than all the 60 PBL cases, despite the same reproducibility ('fair agreement') in 4-tier. EPC/SPC exhibited the lowest kappa value with 'poor agreement' (WHO: $\kappa = 0.13$, 4-tier: $\kappa = 0.03$, 3-tier: $\kappa = 0.05$, and 2-tier: $\kappa = 0.05$).

IHC staining generally improved the interobserver reliability in all 60 PBL cases and IDPN in all classifications (Fig. 3). The reproducibility of IDPN improved to the same level of all 60 PBL cases except that in 2-tier (all 60 PBL cases: 'substantial agreement' and IDPN: 'moderate agreement'). However, the kappa values were lowest in EPC/SPC with 'poor agreement' (WHO: $\kappa = 0.16$, 4-tier: $\kappa = 0.04$, 3-tier: $\kappa = 0.05$, and 2-tier: $\kappa = 0.06$) similar to that in H&E staining, which demonstrated that IHC staining did not improve the diagnostic agreement of EPC/SPC in contrast with all 60 PBL cases and IDPN.

In our 60 cases, five cases were particularly challenging for differential diagnosis with a relatively high discordance rate (Table 3). The presence of apocrine metaplasia (Supplementary Fig. S1) and flat epithelial atypia-like features (Supplementary Fig. S2) made it difficult to distinguish benign from malignant intraductal lesions. Regarding the differential diagnosis between in situ and invasive lesions, we found three challenging cases including one large cystic mass with no myoepithelial cells along the papillae (Supplementary Fig. S3), one with a solid multinodular pattern and smooth contours (Supplementary Fig. S4), and one with a predominant solid multinodular and jigsaw pattern (Supplementary Fig. S5).

DISCUSSION

For 60 PBL cases obtained from CNB, we assessed the interobserver variability and agreement rates in pathologic diagnoses among 20 breast pathologists. In an analysis with the WHO classification, pathologic diagnosis in H&E staining showed 'fair agreement' ($\kappa = 0.21$) with an agreement rate of 47.0%. This result is comparable to those of previous studies in line with ours. In H&E staining for 57 cases of PBLs, three pathologists demonstrated a substantial agreement ($\kappa = 0.79$) in reproducibility and an 86% agreement rate with seven diagnostic categories [11]. Additionally, an analysis with five diagnostic categories indicated moderate agreement ($\kappa = 0.54$) and a 44% agreement rate in 129 PBL cases by H&E staining among four pathologists [12].

Compared with the previous results, it seems that our kappa values and agreement rates are relatively low. The plausible explanations for this finding may be the number of pathologists and the complexity of the diagnostic categories. Our study was performed to assess interobserver variability within 10 diagnostic categories among 20 pathologists. The number of pathologists and diagnostic categories is greater than those of other studies conducted with three pathologists with seven categories [11] and four pathologists with five categories [12].

It seems that the greater number of pathologists makes it harder to obtain a consistent diagnosis for any lesion compared to fewer pathologists. Moreover, a more complicated diagnostic category contributes to lower reproducibility as found in our study. We observed improved reproducibility in simple diagnostic categories, showing the highest kappa value (0.44) and agreement rate (80.0%) in 2-tier. Additionally, the characteristics of diagnostic classification may contribute to the change of reproducibility in our study. The WHO classification features

Table 3. Examples of challenging cases and their challenging points with diagnostic agreement rates in IHC staining among 20 breast pathologists

Challenging case	Challenging point (%)	Diagnosis (agreement rate, n/20)
Apocrine metaplasia	Benign (55)	IDP (30%, 6/20)
		IDP with ADH (25%, 5/20)
	Malignant (45)	IDP with DCIS (35%, 7/20)
		PCIS (10%, 2/20)
Flat epithelial atypia-like features	Benign (30)	IDP with ADH (30%, 6/20)
	Malignant (70)	IDP with DCIS (35%, 7/20)
		PCIS (20%, 4/20)
		EPC (15%, 3/20)
Large cystic pattern with fibrous capsule but no or rare myoepithelial cells	In situ (95)	IDP with DCIS (5%, 1/20)
		PCIS (70%, 14/20)
		EPC (20%, 4/20)
	Invasive (5)	IPC (5%, 1/20)
Solid multinodular pattern with smooth contours but no or rare myoepithelial cells	In situ (65)	EPC (5%, 1/20)
		SPC in situ (60%, 12/20)
	Invasive (35)	SPC invasive (30%, 6/20)
		IPC (5%, 1/20)
Solid multinodular and jigsaw pattern with ragged contours but no myoepithelial cells	In situ (75)	PCIS (20%, 4/20)
		EPC (40%, 8/20)
		SPC in situ (15%, 3/20)
	Invasive (25)	SPC invasive (10%, 2/20)
		IPC (15%, 3/20)

IHC staining, immunohistochemical staining for CK5 and p63; IDP, intraductal papilloma; ADH, atypical ductal hyperplasia; DCIS, ductal carcinoma in situ; PCIS, papillary carcinoma in situ; EPC, encapsulated papillary carcinoma; IPC, invasive papillary carcinoma; SPC, solid papillary carcinoma.

determinacy in diagnosis. However, the pathology category classification (B1-B5) published by the UK National Health Service Breast Screening Programme (NHSBSP) allows for probability in differential diagnosis [14]. In practice, the use of these reporting systems exhibited higher reproducibility ($\kappa = 0.54$) compared to our study ($\kappa = 0.21$) [12]. Therefore, it is assumed that the adoption of this diagnostic classification would lead to the higher reproducibility in our cases.

Elmore et al. [15] investigated the concordance rate of the pathologic diagnosis of non-PBLs on CNB among 115 pathologists recruited from eight U.S. states with consensus-derived reference diagnoses. Their study showed that the overall concordance rate was 75.3% (95% confidence interval, 73.4% to 77.0%; 5,194 of 6,900 interpretations). The diagnostic concordance rate on CNB is lower in PBLs (63.3% in our study and 44% in the previous study [12]) compared to non-PBLs (75.3%) with similar diagnostic categories despite fewer pathologists, indicating more complicated diagnostic difficulty in PBLs.

Multicenter studies are thought to be superior to single-center studies in presenting generalized results in breast pathology. However, to my knowledge, there has been no multicenter study examining the reproducibility of PBLs [15]. In contrast, our study was conducted for 20 pathologists from 20 multiple medical institutions, conferring more generalizability on our findings in PBLs. Additionally, it is noted that our results were derived from breast pathologists. One study indicated that breast pathologists are more accurate in diagnosing CNB guided PBLs compared to non-breast pathologists [13]. In that study, interobserver variability was 'fair agreement' ($\kappa = 0.38$) between the breast pathologists and non-breast pathologists. Therefore, our results suggest the difficulty in diagnosing PBLs even in breast pathologists, warranting improving diagnostic accuracy for PBLs.

IHC staining increases diagnostic accuracy and improves interobserver variability [16]. CK5 is used in distinguishing between hyperplastic and neoplastic epithelial proliferation in PBLs [17,18]. p63 is a nuclear protein that is specific for myoepithelial cells without manifestation in blood vessels and myofibroblasts [19,20]. IHC staining helps to make an accurate diagnosis on PBLs through the ability of CK5 and p63 in identifying monoclonal epithelial proliferation and the presence of myoepithelial cells [17,21], respectively. Our study also showed that the application of IHC staining generally improved interobserver variability and agreement rates in diagnosing PBLs. Nonetheless, it is noted that the utilization of IHC staining is limited in EPC/SPC with very low kappa values.

EPC and SPC are distinctive variants of PCIS, each accounting for <1% of breast carcinomas [22]. Morphological differentiation in H&E staining has a decisive role in diagnosing EPC/ SPC because IHC staining is less helpful. Despite differential points including cystic versus solid and single versus multiple in morphology [23], some cases on CNB practically exhibit overlaps like a transition from single to multiple ductal lesions and cystic to solid appearance with a gradual cystic filling of proliferation [24]. Additionally, definitive cut-off criteria have yet to be determined, which may decrease the diagnostic agreement rates among pathologists. Supplementary Table S1 presents the distribution of histologic patterns and pathologic diagnoses of 12 EPC/SPC cases based on 2-tier or 4-tier classifications, revealing that most pathologists diagnosed most EPC/SPC cases as malignant in 2-tier classification. Nonetheless, it is interesting that kappa values remained low in 2-tier classification for EPC and SPC. This was attributed to the technical limitations of the formula used to calculate kappa values. In cases where the observed agreement was asymmetrically lopsided, kappa values can be drastically lowered due to increased chance agreement rates [25]. Therefore, the tipping effect of lopsided pathologic diagnoses by 20 pathologists induced low kappa values even in 2-tier classification.

We intended to describe five challenging histologic patterns of PBL with diagnostic pitfalls even in IHC staining, specifically apocrine metaplasia, flat epithelial atypia-like features, large cystic masses with no myoepithelial cells along the papillae, and predominant solid multinodular masses with smooth contours or jigsaw patterns. Benign PBLs are often exaggerated by the presence of apocrine metaplasia [10]. Apocrine metaplasia is characterized by abundant eosinophilic cytoplasm with CK5 (-) and a lack of myoepithelial cells with p63 (-) [26,27]. Therefore, the first case with apocrine metaplasia confounded the distinction between benign and malignant intraductal lesions, leading to a diagnostic disagreement even in IHC staining. The WHO classification defined flat epithelial atypia as columnar cell lesions with nuclear atypia [9]. In contrast with non-PBLs, no definite concept of flat epithelial atypia associated with PBLs has been suggested or proposed until now. In the second case with flat epithelial atypia-like features, we observed a high proportion of diagnosis in IDP with ADH (30%) and IDP with DCIS (35%). This heterogeneous diagnosis may be attributable to the difficulty in determining the size (\geq 3 mm in DCIS or < 3 mm in ADH) of histologically identical epithelial proliferation [10]. The third case with a large cystic pattern revealed that PCIS was the most common diagnosis (70%), followed by EPC (20%). EPC is histologically similar to PCIS in some ways, but EPC is characterized by a single cystic or nodular pattern without myoepithelial cells along the papillae and at the periphery, occasionally forming a thick fibrous capsule [10]. The helpful point of distinguishing PCIS from EPC is the presence of myoepithelial cells at the periphery of the PCIS [28]. However, our case presented completely CK5 (–) and sparsely and focally p63 (+) at the periphery, potentially leading to diagnostic disagreement. It is important to differentiate between in situ and invasive lesions for the management and prognostication of PBLs [24]. In the fourth case with a solid multinodular pattern, there were smooth contours and focal suspected microinvasions without immunoreactivity of both CK5 and p63. Although SPC occupied 90% of diagnoses, it was classified as both in situ (60%) and invasive (30%). The fifth case with a solid multinodular and jigsaw pattern was challenging to distinguish between in situ and invasive lesions. Moreover, a solid multinodular pattern suggested SPC, but a fragmented lesion may be misinterpreted as one that fell out of a cystic lesion reminiscent of EPC. Limited materials and fragmented samples in CNB specimens may be the main factors contributing to these disagreements of diagnoses. Interestingly, all 12 cases of EPC/SPC were categorized to the latter three histologic patterns for not only challenging but also helping differential diagnoses as shown in Supplementary Table S1. In papillary carcinomas that were difficult to differentiate between in situ and invasive lesions, such as the above fourth and fifth cases, it is recommended to diagnose them as PCIS or of uncertain invasiveness on CNB to avoid overtreatment, especially in the current era of preoperative (neoadjuvant) chemotherapy.

Our study has some limitations. First, we could not include very rare cases such as IDP with LCIS, EPC with invasion, and IPC because of their extreme rarity especially on CNB. Therefore, PBL cases included in this study actually belong to seven categories as shown in Fig. 1 in contrast to 10 categories of the WHO classification. Second, our cases did not have information on the final diagnosis from excisional biopsy or surgical resection. Although the consensus meeting of KBPSG proposed the pathologic diagnosis of 60 PBL cases, their diagnoses were not likely to guarantee the correctness of the diagnosis. Third, there was no clinical and radiologic information about the 60 PBL cases in our raw data. The absence of this information may be an obstacle in determining an accurate diagnosis and prognosis. Fourth, we did not use the most recently updated version of the WHO classification published in 2019. Nonetheless, because there was no difference between the 4th and 5th editions in diagnosing and classifying PBLs, the concern for the discrepancy between the two versions is minimal [8,9].

In conclusion, our study demonstrated that interobserver variability in the pathologic diagnosis of PBLs was unsatisfactory among 20 breast pathologists from 20 multiple medical institutions. Although IHC staining improved interobserver variability and agreement rates in diagnosing PBLs, diagnostic reproducibility was still limited in specific cases including EPC/SPC. Therefore, more intensive consensus studies are necessary to improve the diagnostic agreement and categorization of PBLs with the WHO classification. Further studies should continue to develop effective modalities in distinguishing PBLs especially on CNB.

Supplementary Information

The Data Supplement is available with this article at https://doi.org/10.4132/jptm.2021.07.29.

Ethics Statement

This study was approved by the National Cancer Center Institutional Review Board with a waiver of informed consent (NCC2018-0214).

Availability of Data and Material

The datasets generated or analyzed during the study are available from the corresponding author on reasonable request.

Code Availability

Not applicable.

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Conflicts of Interest

S.Y.P., editor-in-chief of the *Journal of Pathology and Translational Medicine*, were not involved in the editorial evaluation or decision to publish this article. All remaining authors have declared no conflicts of interest.

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